# FINAL REPORT

# VINELAND CHEMICAL SUPERFUND SITE: POST REMEDIAL ACTION SAMPLING – SEPTEMBER 2007 OPERABLE UNITS #3 AND #4 VINELAND, NEW JERSEY

# Prepared for:



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# LIST OF ABBREVIATIONS, ACRONYMS, AND UNITS

ADR Automated Data Review

ASTM American Society for Testing and Materials

°C Degrees Celsius COC Chain of Custody

DESA Division of Environmental Science and Assessment

DGPS Differential Global Positioning System

EA Engineering, Science, and Technology, Inc.

EDDs Electronic Data Deliverables

EM Engineer Manual

ft Foot/Feet

HNO<sub>3</sub> Nitric acid

in Inch(es)

MDL Method Detection Limit

mg/Kg Milligram(s) Per Kilogram (ppm)

mg/L Milligram(s) Per Liter

mL Milliliter(s)

MS/MSD Matrix Spike/Matrix Spike Duplicate

NAD83 North American Datum 1983

NJ New Jersey ND Non-detect

PCB Polychlorinated Biphenyl

ppb Part(s) Per Billion (µg/kg or µg/L)
ppm Part(s) Per Million (mg/Kg or mg/L)
ppt Part(s) Per Thousand (g/kg or g/L)

QA Quality Assurance QC Quality Control

RL Reporting Limit ROD Record of Decision

SOP Standard Operating Procedure

μg/L Microgram(s) Per Liter (ppb)

μm Micrometer(s)

USACE U.S. Army Corps of Engineers

USEPA U.S. Environmental Protection Agency

#### **EXECUTIVE SUMMARY**

The Vineland Chemical site is a 54-acre manufacturing facility located in Cumberland County, New Jersey (NJ) (Figure ES-1). The facility was involved in the production of arsenical herbicides, fungicides, and biocides since 1949. Arsenical feedstock compounds were historically stored in unprotected piles that resulted in soil and groundwater contamination in the vicinity of the site. Runoff during storm events and the recharge of arsenic-bearing groundwater has contaminated the adjacent watershed, including soil, sediment, and surface waters of nearby waterways such as Blackwater Branch, Maurice River, and Union Lake (Figure ES-1). Four long-term, remedial phases at the site focus on source control, migration management, and cleanup of the rivers and Union Lake sediments, which was the subject of a Record of Decision (ROD) in 1989 (USEPA 1989). Two general areas of consideration for the study include public health and remedial actions. The current phase of remediation at the site involves removing the contaminated soils/sediments of the Blackwater Branch and the floodplain east of Mill Road and adjacent to the site. This excavation has the potential to stir up sediments and impact the waterways downstream. A monitoring program has been initiated that includes baseline (preexcavation) sampling, during excavation/construction sampling, and post-construction sampling to determine the status of exposure and impacts to human health exposure pathways. The first survey, the May 2006 Baseline Sampling, was conducted prior to the initiation of planned remedial excavation activities in the Blackwater Branch (EA 2006a). The second survey, the November 2006 Periodic Sampling, was conducted during excavation activities at the site. The third survey, the September 2007 Post Remedial Action Sampling, was conducted following the completion of excavation for remedial action activities.

This report presents the results of the Post Remedial Action Sampling that was conducted in September 2007. The Post Remedial Action Sampling was designed to identify, analyze, and evaluate the arsenic concentrations in sediments, soil, and water collected at ten locations in and near waterways located adjacent to the site following the Blackwater Branch excavation activities as part of the third round of sampling. EA Engineering, Science, and Technology, Inc. (EA) was contracted by the U.S. Army Corps of Engineers (USACE) - Philadelphia District to conduct sediment, soil, and surface water sampling at ten locations along Blackwater Branch, the Maurice River, and Union Lake. The arsenic concentration in each of the samples was measured by the U.S. Environmental Protection Agency (USEPA) Region II Laboratory located in Edison, New Jersey. The Sampling and Analysis Plan (SAP) (EA 2006b) described the sampling and data-gathering methods for the project and followed guidance provided by the USACE Engineer Manual (EM) 200-1-3 Requirements for Preparation of Sampling and Analysis Plans (1994).

The following types of samples were collected and analyzed for arsenic during the September 2007 Post Remedial Action Sampling:

- Surficial sediment samples (0-0.5 ft depth increment beneath the water/sediment interface) co-located with the surface water samples collected at either midstream (for the river reaches) or at greater than 200 ft from the shoreline (for the lake stations);
- Surficial sediment nearshore (shore) samples (0-0.5 ft depth increment beneath the water/sediment interface) collected 2-10 ft below the waterline;
- Surface water collected prior to sediment collection or disturbance at each site;

- Surface water collected following agitation of sediment upstream from each sampling point;
- Beach soils collected approximately 6-10 ft above the waterline, and;
- Sediment borings collected to refusal with analysis of one to three depth intervals (0-1 ft, 1-2 ft, and approximately 2-4 ft), depending upon refusal depth.

Detected arsenic concentrations in water samples were compared to the USEPA Drinking Water Criterion for arsenic of 10 parts per billion (ppb or  $\mu g/L$ ), and the results for detected arsenic concentrations in sediment and soil were compared to the Site Clean-up Level of 20 parts per million (ppm or mg/Kg) for arsenic in solids. The Site Clean-up Level of 20 ppm is based upon the New Jersey Residential Clean-up Standard for Arsenic. The water, soil, and sediment results for each station from the September 2007 Post Remedial Action Sampling were also compared to the May 2006 Baseline Sampling results (EA 2006a) and the November 2006 Periodic Sampling results (EA 2007). Additionally, the water, soil, and sediment results from the September 2007 Post Remedial Action Sampling beach stations were compared to historical arsenic data collected from five beach stations during the year 1992 and from 1994 through 1999.

The September 2007 Post Remedial Action Sampling also included analyzing sediment, soil, and water samples for nine Polychlorinated Biphenyl (PCB) aroclors. The aroclor concentrations were summed and the total PCB concentrations in water samples were compared to the USEPA Drinking Water Maximum Contaminant Level for total PCBs of 0.5 (ppb or  $\mu g/L$ ), and the total PCB concentrations in sediment and soil were compared to the Soil Remediation Standard of 0.2 (ppm or mg/Kg; 200 ppb or  $\mu g/Kg$ ) for total PCBs in solids. The Soil Remediation Standard of 0.2 ppm is based upon the New Jersey Residential Direct Contact Soil Remediation Standard for total PCBs.

# ES.1 ARSENIC RESULTS

Figure ES-2 presents arsenic concentrations for each of the three sampling events: the May 2006 Baseline Sampling, the November 2006 Periodic Sampling, and the September 2007 Post Remedial Action Sampling.

Overall, results of the September 2007 Post Remedial Action Sampling indicated more exceedences of the arsenic criterion for agitated water samples than in previous sampling events. Arsenic concentrations in agitated water samples exceeded the 10 ppb criterion at seven of the ten sampling locations. Both the May 2006 Baseline and the November 2006 (during excavation) Periodic Sampling events exhibited only three locations with agitated water exceedences. The arsenic concentration in agitated water (3,800 ppb) collected in September 2007 from Station 1 (West of Mill Rd) was substantially higher than any other water sample tested during the three sampling events.

As indicated in the May 2006 and November 2006 Sampling events, the two stations located immediately downstream of the Vineland site, Station 1 (West of Mill Rd.) and Station 2 (West of Rte. 55) had the highest concentrations of arsenic in sediment and had the greatest number of concentrations that exceeded the Site Clean-up Level of 20 ppm for arsenic (Figure 3-1). Unlike the May 2006 and November 2006 Sampling events, the arsenic at stations located further

downstream, including Station 7 (Sherman Ave), Station 8 (North End of Union Lake), and Station 9 (Union Lake), more frequently exceeded the water and soil criteria than in previous sampling events. Although located furthest downstream of the Vineland site, the stations located along Union Lake, including Station 8 (North End of Union Lake) and Station 9 (Union Lake Beach) had arsenic concentrations in sediments that exceeded the Site Clean-up Level of 20 ppm. These trends in Union Lake may be attributable to the proportion of fine silt/clays that were observed in the sediment samples; arsenic is strongly sorbed onto fine particulates, including silt (Bodek et. al 1988). The arsenic that originates from upstream sources may be transported downstream via particulates which settle out in the lake depositional areas. Previous reports for the site have stated that sediment in the Maurice River and Union Lake contains a high content of organic matter (USEPA 1999). Importantly, with the exception of Station 7 (Sherman Ave), the arsenic concentrations that exceeded the Site Clean-up Level of 20 ppm were for lake sediments collected greater than 200 ft from the shoreline. In addition, arsenic concentrations from the five beach locations [Station 4 (Alliance Beach), Station 5 (Almond Beach), Station 6 ("BareA" Beach), Station 9 (Union Lake Beach), and Station 10 (South End Union Lake Beach)] were either < 1.0 mg/Kg or below the analytical detection limit.

Below Station 2 (West of Rte. 55), additional water flow from the Maurice River and other tributaries flowing into the Maurice River may transport arsenic that is bound to fine particulates further downstream. Previously in the May 2006 and November 2006 Sampling events, arsenic concentrations in sediments, water, and beach soil did not exceed criteria at Station 4 (Alliance Beach), Station 5 (Almond Beach), or Station 6 ("BareA" Beach). However, in September 2007, the first exceedence of applicable criteria occurred at Station 5 (Almond Beach) in the agitated water sample and at Station 6 ("BareA" Beach) in both a surface water sample and a nearshore (shore) sediment sample.

Ten total sediment boring samples were collected from six of the ten sampling stations. Four of the six stations that were sampled for sediment borings were collected only for the first depth interval (0-1 ft / closest to surface) and resulted in either no detection or low detection of arsenic; grain size analyses in the first depth interval for these stations indicated that sand was the dominant substrate. Two of the six stations, Station 1 (West of Mill Rd) and Station 2 (West of Rte. 55), were sampled in three depth intervals for the sediment borings. The general trends observed for these stations resulted in arsenic greater than the 20 ppm criterion for the first depth interval (0-1 ft deep), higher arsenic concentrations for the second depth interval (1-2 ft deep), and concentrations less than the 20 ppm criterion in the deepest third depth interval (2-3 or 2-4 ft deep). Grain size data for Station 2 (West of Rte. 55) indicated that fine grained material (silts/clays) dominated the first (0-1 ft) depth interval, while the grain size data for Station 1 (West of Mill Rd) indicated that sand dominated the first (0-1 ft) depth interval.

Arsenic results for each sampling location and matrix (sediment, water, beach soil) are presented in Figure ES-2 and summarized as follows:

# Station 1 – West of Mill Rd (see Figures 3-1 and 3-2)

Arsenic concentrations in the agitated water sample exceeded the 10 ppb USEPA Drinking Water Criterion by a factor of 380; at 3,800  $\mu$ g/L, the arsenic concentration at Mill-Wat2 was the single highest concentration measured during the September 2007 Post Remedial Action

Sampling. Arsenic concentrations in the nearshore (shore) surficial sediment exceeded the Site Clean-up Level of 20 ppm by a factor of 6.5 and in the sediment borings by a factor of 4.7 (Boring-1) and by a factor of 23 (Boring-2). Arsenic concentrations in neither the mid-stream surficial sediment (0-0.5 ft depth increment) nor the third-depth sediment boring (Boring-3) exceeded the Site Clean-up Level of 20 ppm

# Station 2 – West of Rte 55 (see Figures 3-1 and 3-2)

Both of the arsenic concentrations in the surface water and agitated water sample exceeded the 10 ppb USEPA Drinking Water Criterion by a factor of 1.4 and 70, respectively. Arsenic concentrations also exceeded the Site Clean-up Level of 20 ppm in the mid-stream surficial sediment (0-0.5 ft depth increment) by a factor of 85, in the nearshore (shore) surficial sediment by a factor of 170, and in the sediment borings by a factor of 95 (Boring-1) and by a factor of 180 (Boring-2). The third boring (Boring-3) was the only sample that did not exceed the Site Clean-up Level of 20 ppm.

# Station 3 – BWB & Maurice Confluence (see Figures 3-1 and 3-3)

None of the arsenic concentrations detected in sediment samples exceeded the Site Clean-up Level of 20 ppm. Arsenic was not detected in either of the water samples.

# Station 4 – Alliance Beach (see Figures 3-1 and 3-3)

Arsenic was not detected in any of the sediment, water, or beach soil samples at Station 4.

# Station 5 – Almond Beach (see Figures 3-1 and 3-4)

None of the arsenic concentrations detected in sediment samples exceeded the Site Clean-up Level of 20 ppm. Arsenic was not detected in beach soils, in the nearshore (shore) surficial sediment, in the surface water sample, or the one sediment boring (0-1 ft) that was collected; arsenic was detected at 1.3 mg/Kg in the mid-stream surficial sediment (0-0.5 ft depth increment). The arsenic concentration in the agitated water sample exceeded the 10 ppb USEPA Drinking Water Criterion by a factor of 1.6.

# Station 6 – "BareA" Beach (see Figures 3-1 and 3-5)

The arsenic concentration in the nearshore (shore) surficial sediment was 290 mg/Kg, which exceeded the Site Clean-up Level of 20 ppm by a factor of 14.5. Arsenic was detected at 2.3 mg/Kg in the mid-stream surficial sediment (0-0.5 ft depth increment). Arsenic was not detected in beach soils or in the agitated (Wat-2) water sample. However, the arsenic concentration in the surface water sample was 13  $\mu$ g/L, which exceeded the 10 ppb USEPA Drinking Water Criterion by a factor of 1.3.

### Station 7 – Sherman Ave. (see Figures 3-1 and 3-6)

Arsenic concentrations in the mid-stream surficial sediment (0-0.5 ft depth increment) sample and both water samples exceeded applicable criteria. The arsenic concentration in the mid-stream surficial sediment (0-0.5 ft depth increment) was 110 mg/Kg, which exceeded the Site Clean-up Level of 20 ppm by a factor of 5.5. The arsenic concentrations in the surface water (12  $\mu$ g/L) and the agitated water (100  $\mu$ g/L) sample exceeded the 10 ppb USEPA Drinking Water Criterion by factors of 1.2 and 10, respectively. Arsenic was detected at 5.5 mg/Kg in the nearshore (shore) surficial sediment.

# Station 8 – North End of Union Lake (see Figures 3-1 and 3-7)

Both of the arsenic concentrations in the surficial and nearshore (shore) sediment samples exceeded the Site Clean-up Level of 20 ppm. The arsenic concentration in the surficial sediment (0-0.5 ft depth increment) collected 200 ft offshore was 30 mg/Kg and in the nearshore (shore) surficial sediment was 320 mg/Kg, which exceeded the Site Clean-up Level of 20 ppm by factors of 1.5 and 16, respectively. Arsenic was not detected in the surface water, but was detected in the agitated water sample (22  $\mu$ g/L) and exceeded the 10 ppb USEPA Drinking Water Criterion by a factor of 2.2.

# Station 9 – Union Lake Beach (see Figures 3-1 and 3-8)

Arsenic was detected in each of the soil/sediment samples, including the one sediment boring (0-1 ft) collected at Station 9. Arsenic was detected at 360 mg/Kg in the surficial sediment (0-0.5 ft depth increment) collected 200 ft offshore and exceeded the Site Clean-up Level of 20 ppm by a factor of 18. Arsenic was detected at 2.1 mg/Kg in the nearshore (shore) surficial sediment, at 1.0 mg/Kg in the beach (soil) sample, and at 1.4 mg/Kg in the sediment boring (Boring-1). Arsenic was not detected in the non-agitated surface water, but was detected in the agitated water sample (15 μg/L) and exceeded the 10 ppb USEPA Drinking Water Criterion by a factor of 1.5.

# Station 10 – South End of Union Lake Beach (see Figures 3-1 and 3-8)

None of the detected arsenic concentrations in sediment samples exceeded the Site Clean-up Level of 20 ppm. Arsenic was detected in the surficial sediment (0-0.5 ft depth increment) collected 200 ft offshore at 18 mg/Kg, in the nearshore (shore) surficial sediment at 6.5 mg/Kg, and in the sediment boring (Boring-1) at 12 mg/Kg. Arsenic was not detected in the beach (soil) sample. Arsenic was not detected in the surface water, but was detected in the agitated water sample ( $28 \mu g/L$ ) and exceeded the 10 ppb USEPA Drinking Water Criterion by a factor of 2.8.

# ES.2 COMPARISONS TO HISTORICAL ARSENIC DATA

During 1992 and from 1994 through 1999, water, soil, and sediment samples were collected in the vicinity of and downstream of the Vineland site at beach stations for arsenic analyses (USEPA/ERTC 1999). Data were collected from five beach locations including Alliance Beach (Station 4), Almond Beach (Station 5), "BareA" Beach (Station 6), Union Lake Beach (Station 9), and South End Union Lake Beach (Station 10). These data were compared to the May 2006 Baseline Sampling, the November 2006 Periodic Sampling, and the September 2007 Post Remedial Action Sampling for surface water, beach soils, and surficial sediment arsenic concentrations.

### Surface Water Data

Throughout the period of 1992 and 1994-1999, arsenic concentrations in surface waters at Station 4 (Alliance Beach), Station 5 (Almond Beach), and Station 6 ("BareA" Beach) were variable and substantially exceeded the current USEPA Drinking Water Criterion of 10 ppb. Arsenic concentrations in surface waters at Station 9 (Union Lake Beach) slightly declined from 1996 (above criterion) through 1999 (below criterion). The arsenic concentration in surface water at Station 10 (South End of Union Lake Beach) was above the criterion in both 1998 and 1999. Surface water data from samples collected in both May 2006 and November 2006

indicated that arsenic was not detected in the surface waters at each of these five previously sampled locations (Stations 4, 5, 6, 9, and 10), with the exception of Station 10 (South End of Union Lake Beach) in May 2006, when arsenic in the agitated water sample was equivalent to the USEPA Drinking Criterion. However, in September 2007, there were several exceedences of the arsenic criterion in surface water. The majority of these exceedences corresponded to agitated water samples.

# Beach Soil Data

None of the beach soil samples collected in 1992, 1994 through 1999, and 2006 exceeded the Site Clean-up Level of 20 ppm for arsenic at Station 4 (Alliance Beach), Station 5 (Almond Beach), Station 6 ("BareA" Beach), Station 9 (Union Lake Beach), and Station 10 (South End of Union Lake Beach). Detected concentrations in May 2006, November 2006, and September 2007 were either comparable to or lower than those previously reported for each of the five stations and none of the 2006 or 2007 samples exceeded the Site Clean-up Level of 20 ppm.

# Surface Sediment Data

Throughout the period of 1992 and 1994-1999, arsenic concentrations were below the Site Clean-up Level of 20 ppm at each of the five stations, with the exception of Station 6 ("BareA" Beach) in 1998. Results from samples collected in May 2006 indicated that arsenic concentrations in surficial sediment (collected greater than 200 ft from the shoreline) at Station 9 (Union Lake Beach) and Station 10 (South End of Union Lake Beach) were substantially higher than concentrations previously reported in 1992 and 1994-1999. Results from samples collected in both November 2006 and September 2007 indicated that arsenic concentrations in surficial sediment (collected greater than 200 ft from the shoreline) at Station 9 (Union Lake Beach) were substantially higher than concentrations previously reported in 1992 and 1994-1999, but comparable to concentrations reported in May 2006 at this station. These changes could potentially be attributable to downstream transport of arsenic bound to fine grained materials (i.e., silts) and their subsequent accumulation in depositional areas of the lake.

# ES.3 PCB RESULTS

None of the 20 surface water samples analyzed had detected concentrations of PCB aroclors. Two of the 36 sediment samples, boring-2 and boring-3 at Station 2 (West of Rte 55), had detected concentrations for Aroclor-1254 of 9.4 and 4.1 ppb ( $\mu g/Kg$ ), respectively. However, neither of these sediment concentrations exceeded the 200 ppb ( $\mu g/Kg$ ) Soil Remediation Standard for total PCBs.

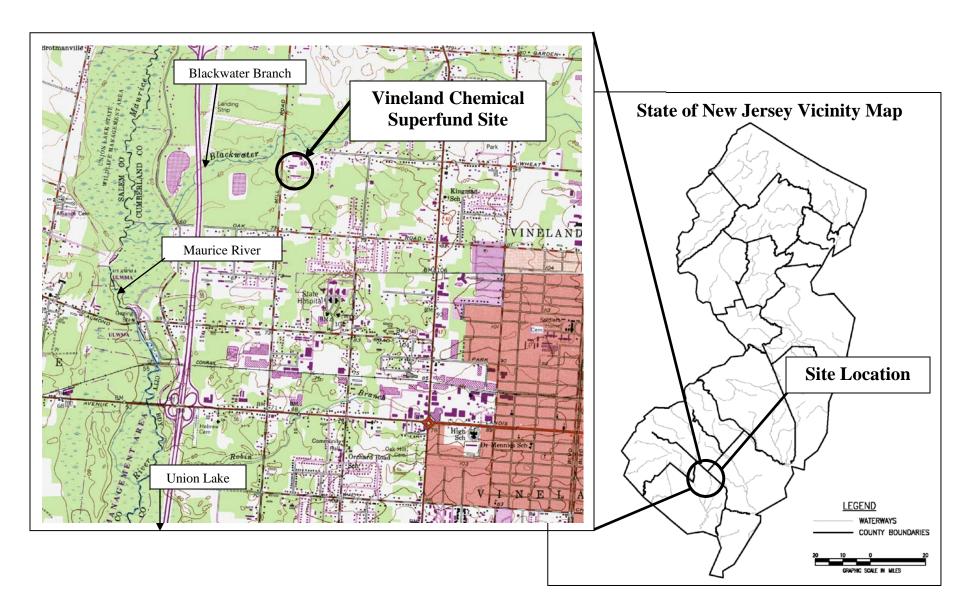
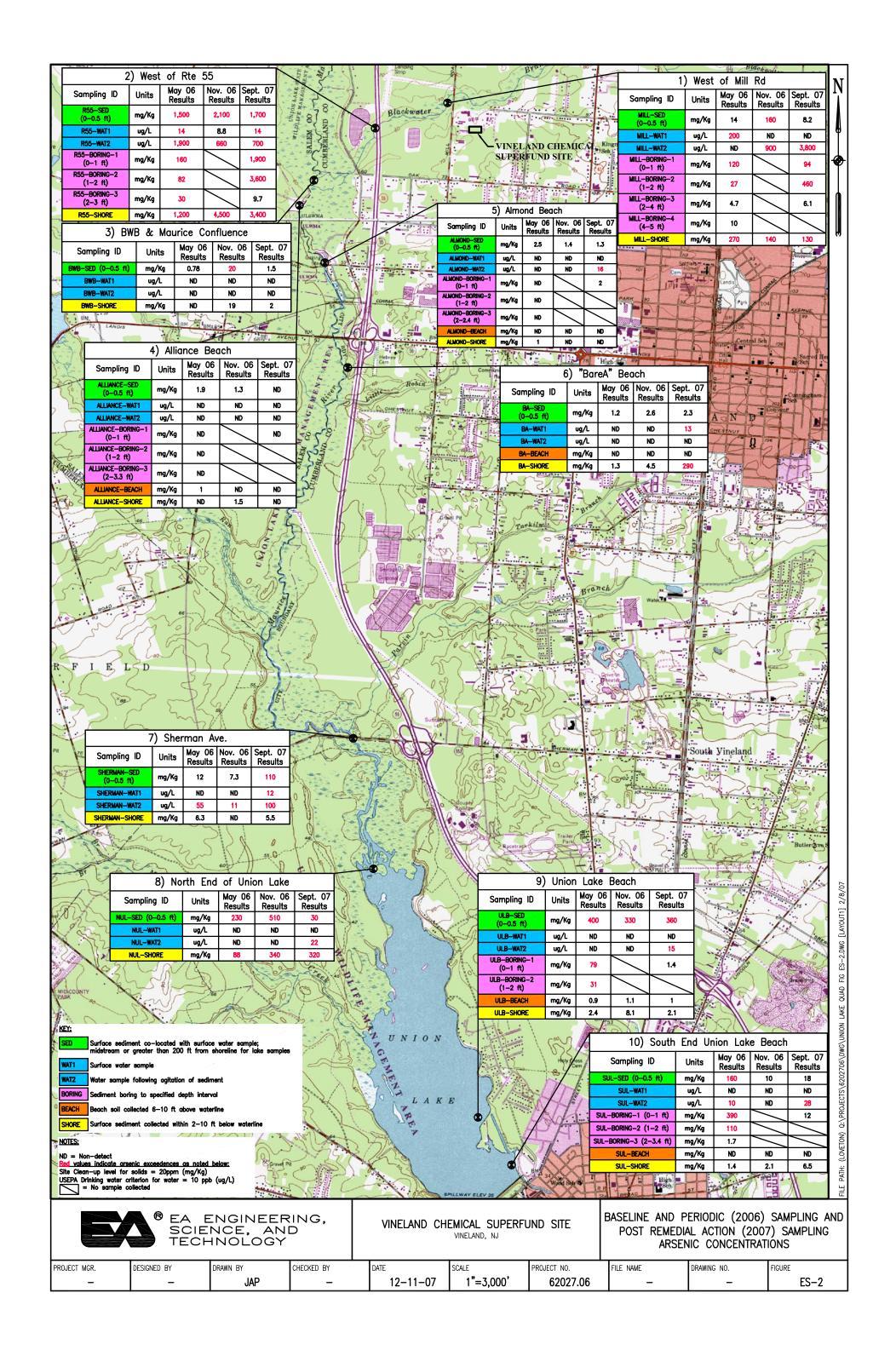


Figure ES-1. Vineland Chemical Superfund Site Location Map, Cumberland County, NJ



# 1. INTRODUCTION

This report presents results from an arsenic survey in the vicinity of Vineland Chemical Company Superfund Site in Cumberland County, New Jersey (NJ) that was conducted on 10 through 13 September 2007 in accordance with the Sampling and Analysis Plan, Vineland Chemical Superfund Site: Baseline Sampling and Monitoring Program, Operable Units #3 and #4, Vineland, New Jersey (EA 2006b). These data will be used to document the arsenic concentrations in nearby waterways that have been impacted by previous operations of the site following the remediation activities. Results for soils and sediments will be compared to the Site Clean-up Level of 20 parts per million (ppm or mg/Kg) and results for surface water will be compared to the U.S. Environmental Protection Agency (USEPA) Drinking Water Criterion of 10 parts per billion (ppb or µg/L) for the protection of human health. These data (referred to as the September 2007 Post Remedial Action Sampling) represent the third of three sampling and monitoring events that were planned to assess the potential impacts of remedial activities at the site.

In addition to the analysis of arsenic, the September 2007 Post Remedial Action Sampling included the analysis of sediment, soil, and water samples for polychlorinated biphenyl (PCB) aroclors. Total PCB concentrations in water samples were compared to the USEPA Drinking Water Maximum Contaminant Level for total PCBs of 0.5 (ppb or  $\mu g/L$ ), and the results for detected total PCB concentrations in sediment and soil were compared to the Soil Remediation Standard of 0.2 ppm for total PCBs in solids. The Soil Remediation Standard of 0.2 ppm is based upon the New Jersey Residential Direct Contact Soil Remediation Standard for total PCBs.

# 1.1 PROJECT BACKGROUND

Previous studies have shown that the Vineland Chemical Company Superfund Site (site) has arsenic contamination in the soils, sediments, and ground water. The site manufactured arsenicbased herbicides from 1950 to 1994 on a 54-acre site in a residential and industrial area of the City of Vineland, NJ. The site is located adjacent and upstream from nearby waterways that include the Blackwater Branch, Maurice River, and Union Lake (Figure 1-1). The soil, sediment, and water of these waterbodies have been impacted by the operations of the site. Beginning in 1982, and in response to State actions, the Vineland Chemical Company instituted some cleanup actions and modified the production process. The site clean-up is being addressed in two stages, including immediate actions and long-term remedial phases. Four long-term, remedial phases will focus on source control, migration management, and cleanup of the rivers and Union Lake sediments, which was the subject of a Record of Decision (ROD) in 1989 The current phase of remediation at the site involves removing the contaminated soils/sediments of the Blackwater Branch and the floodplain east of Mill Road and adjacent to the site. This excavation has the potential to stir up sediments and impact the waterways downstream. Therefore, baseline (pre-excavation), during excavation, and postremedial action sampling/monitoring is required.

Two general areas of consideration for the study include public health and remedial actions. The monitoring and sampling program is being completed to determine the status of exposure and

impacts to human health exposure pathways. Results from sampling efforts will determine the extent of contamination in the surrounding areas prior to excavation activities, during excavation, and post-excavation. The first survey, the May 2006 Baseline Sampling (EA 2006a), was conducted prior to the initiation of planned remedial excavation activities in the Blackwater Branch. The second survey, the November 2006 Periodic Sampling (EA 2007), was conducted during excavation activities at the site. This third survey, the September 2007 Post Remedial Action Sampling, was conducted following the completion of excavation for remedial action activities.

A three-year period of monitoring and sampling will be implemented at the completion of Operational Unit #1 remediation activities. This program will provide data to assist with determination of the impacts associated with ongoing remedial activities, including the removal of contaminated soil and sediments and the pump and treat groundwater program to facilitate evaluation of further remedial action in the river areas and Union Lake.

# 1.2 PROJECT LOCATION

The Vineland Chemical site is a 54-acre manufacturing facility located in Vineland, Cumberland County, NJ (Figure 1-1). The site is located in south-central NJ, approximately 40 miles from Wilmington, Delaware and approximately 35 miles from Atlantic City, NJ. The facility was involved in the production of arsenical herbicides, fungicides, and biocides since 1949. Arsenical feedstock compounds were historically stored in unprotected piles. This resulted in soil and groundwater contamination in the vicinity of the site. Runoff during storm events and the recharge of arsenic-bearing groundwater has contaminated the adjacent watershed, including nearby waterways such as Blackwater Branch, Maurice River, and Union Lake.

# 1.3 PROJECT PURPOSE AND OBJECTIVES

Determination of arsenic concentrations in the sediments, soil, and water in the vicinity of the site is necessary in order to provide information about the environmental conditions at the site during and following the Blackwater Branch excavation activities to assess potential human exposure to arsenic and to document the extent of contamination. This third sampling and monitoring effort, the September 2007 Post Remedial Action Sampling, documents the levels of arsenic concentrations in the sediment, soil, and water following the excavation process, and compares current (September 2007) arsenic concentrations to the May 2006 Baseline Sampling (EA 2006a), the November 2006 Periodic Sampling (EA 2007) and the historic (1992 and 1994 through 1999) arsenic concentrations at the site. Additionally, a three-year period of sampling and monitoring will be conducted to determine the impacts of ongoing remedial activities including removal of contaminated soil and sediments and pump and treat groundwater program to facilitate evaluation of further remedial action in the River Areas and Union Lake. In addition to the analysis of arsenic, the September 2007 Post Remedial Action Sampling also included the analysis of sediment, soil, and water samples for PCB aroclors. The September 2007 Post Remedial Action Sampling program consisted of the following tasks:

• Sediment, soil, and water sample collection at 10 locations;

- Sediment boring sample collection at 6 locations;
- Analytical testing of sediment, soil, and water samples for arsenic concentrations;
- Analytical testing of sediment, soil, and water samples for PCB aroclor concentrations;
- Data report preparation and submittal.

### 1.4 EXPERIMENTAL DESIGN

The executing agency for this project is the U.S. Army Corps of Engineers (USACE), North Atlantic Division, Philadelphia District. This investigation was designed to identify, analyze, and evaluate the arsenic concentrations in sediments, soil, and water collected at ten locations in and near waterways located adjacent to the site. EA Engineering, Science, and Technology, Inc. (EA) was contracted by the USACE - Philadelphia District to conduct sediment, soil, and surface water sampling at ten locations along Blackwater Branch, the Maurice River, and Union Lake. Arsenic concentrations in each of the samples were measured by the USEPA Region II Laboratory located in Edison, NJ. The Sampling and Analysis Plan (EA 2006) described the sampling and data-gathering methods for the project and followed guidance provided by the USACE Engineer Manual (EM) 200-1-3 Requirements for Preparation of Sampling and Analysis Plans (1994). Grain size analyses for the sediment cores were conducted by E2CR, a geotechnical firm located in Baltimore, MD. Analytical testing for PCBs was conducted by DataChem Laboratories located in Salt Lake City, UT.

# 1.5 REPORT ORGANIZATION

This report contains a comprehensive summary of field activities and the results of the sediment, soil and water analyses. Field sampling techniques and analytical methodologies for arsenic and PCB aroclor analyses are provided in Chapter 2, results of the arsenic analyses are provided in Chapter 3, and results of the PCB aroclor analyses are provided in Chapter 4. A summary of findings and a comparison to historical data is provided in Chapter 5. References cited are provided in Chapter 6. Appendix A presents the analytical results and accompanying Chain-of-Custody (COC) forms from the arsenic analyses; Appendix B presents the grain size analysis results and accompanying COC forms; Appendix C presents the analytical results and accompanying COC forms for the PCB aroclor analyses; Appendix D provides a copy of the field logbook; Appendix E presents the Standard Operating Procedures (SOPs) for the laboratory analysis of arsenic; Appendix F presents the SOPs for the laboratory analysis of PCB aroclors; and Appendix G presents the historical arsenic data results from the years 1992 and from 1994 through 1999.

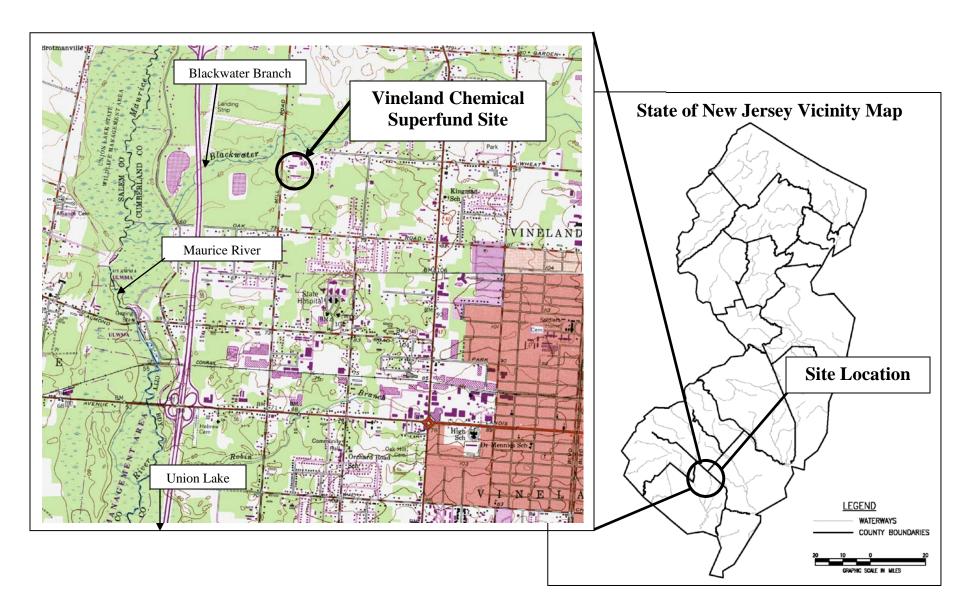


Figure 1-1. Vineland Chemical Superfund Site Location Map, Cumberland County, NJ

# 2. METHODOLOGY

This September 2007 Post Remedial Action Sampling event was carried out in accordance with the *Sampling and Analysis Plan Vineland Chemical Superfund Site: Baseline Sampling and Monitoring Program* (EA 2006b). Collection of the water, soil, and sediment quality samples was initiated on 10 September and continued through 13 September 2007.

Two water samples (the first representing a sample without bottom disturbance and the second representing a sample following agitation/disturbance of the bottom sediments) were collected from each of ten stations (20 total water samples). In addition, one surficial sediment sample was collected from mid-stream (or 200 ft from offshore at lake locations) at each of ten stations, one nearshore (shore) sediment sample was collected from each of ten stations, and one beach soil sample was collected from each of five stations, and sediment cores (boring) samples were collected from six stations (Table 2-1). Each sediment, soil, and water sample was analyzed for total arsenic concentrations and PCB aroclor concentrations. In addition, eight of the ten sediment core (boring) samples were submitted for grain size analyses.

# 2.1 SAMPLING OBJECTIVES

The third sampling effort included surficial sediment sampling, sediment boring sampling, surface water sampling, nearshore (shore) sediment sampling, and beach soil sampling. Table 2-1 provides the sampling locations and number of samples collected as part of the September 2007 sampling event for the project. Table 2-2 provides the sampling locations, coordinates of the sampling efforts, and sample identification. The overall objectives of the field sampling were to:

- Collect two surface water samples (mid-stream and mid-depth) of the water column (one prior to sediment disturbance and one following sediment disturbance) at each of 10 locations for arsenic and PCB aroclor analysis;
- Collect one shallow sediment sample (0-0.5 ft depth increment beneath the water/sediment interface) at 10 locations at either mid-stream (for upper sampling locations approx. 2-3 ft from shoreline) or greater than 200 ft from the shoreline (lake sampling locations) for arsenic and PCB aroclor analysis;
- Collect sediment borings at six locations for arsenic and PCB aroclor analysis;
- Collect one shallow, nearshore (shore) sediment sample (0-0.5 ft depth increment beneath the water/sediment interface) at 10 locations approximately 2-10 ft below the waterline for arsenic and PCB aroclor analysis;
- Collect one beach soil sample at five locations approximately 6-10 ft above waterline for arsenic and PCB aroclor analysis;

- Collect and transfer sediment, soil, and water samples to appropriate, laboratory-prepared containers and preserve/hold samples for analysis according to protocols that ensure sample integrity;
- Measure and record *in situ* water quality information (temperature, conductivity, salinity, dissolved oxygen, and pH) at each surface water sampling location;
- Submit equipment blanks, duplicates, and matrix spike/matrix spike duplicates for analytical testing; and
- Complete appropriate COC documentation.

# 2.2 SAMPLING LOCATION DETERMINATION

Sampling locations were provided by USACE-Philadelphia District and correspond to locations that were sampled in previous investigations (USEPA/ERTC 1999, EA 2006a, EA 2007). Sampling locations and northing and easting coordinates [NJ State Plane North American Datum 1983 (NAD83)] are provided in Table 2-2. Positioning in the field was determined using a Trimble ProXR Differential Global Positioning System (DGPS), which utilizes the United States Coast Guard Differential Beacon System to obtain sub-meter accuracy. Sample locations and a brief description are included below:

# **Sample Location: Description of Sampling Location:**

1) West of Mill Rd	Along Blackwater Branch, immediately downstream of site		
2) West of Rte 55	Along Blackwater Branch, further downstream of site		
3) BWB & Maurice Confluence	At the Blackwater Branch and Maurice River confluence		
4) Alliance Beach	Privately owned and located along the Maurice River and upstream of Almond Beach		
5) Almond Beach	Along the Maurice River, publicly maintained beach area, approximately 100-150 ft long		
6) "BareA" Beach	Along the Maurice River, downstream of Almond Beach, unmaintained public day-use area		
7) Sherman Ave.	Along the Maurice River, at the Sherman Avenue Bridge		
8) North End of Union Lake	In the northern section of Union Lake		
9) Union Lake Beach	Privately maintained beach area, downstream of site (access at Union Lake Sailing and Tennis Club)		
10) South End Union Lake Beach	In the southern section of Union Lake, north of the spillway		

# 2.3 SAMPLE VOLUME REQUIREMENTS

The sample volume requirements are detailed in Table 2-1 for arsenic analyses. Arsenic analysis of sediments and beach soils required 250 grams of sediment per sample. Water samples required 250 milliliters (ml) per sample for arsenic analysis. PCB aroclor analysis of sediments required approximately 400 grams of sediment per sample, and PCB aroclor analysis of water samples required approximately 2-liters (L) of water per sample. For arsenic and PCB aroclor analyses, a total (not including field duplicates, matrix spike, and matrix spike duplicates) of ten sediment samples, ten shore samples, five beach soil samples and ten sediment boring samples were collected for the September 2007 Post Remedial Action Sampling effort. For arsenic and PCB aroclor analyses, 20 water samples (not including field duplicates, equipment blanks, matrix spike, and matrix spike duplicates) were collected. Additionally, sufficient volume of sediment from ten sediment boring samples were collected and submitted for grain size analyses.

# 2.4 IN SITU WATER QUALITY MEASUREMENTS

Water quality measurements were recorded *in situ* at each of the ten stations using a YSI water quality probe. Measurements were recorded at the same locations where water samples were collected for chemical analysis (mid-stream/mid-depth of the water column). The following parameters were recorded in the field log book:

- Sampling location number
- Sampling data and time
- Station depth
- Weather conditions
- Water temperature [degrees (<sup>0</sup>) Celsius]
- Conductivity (mS/cm)
- Salinity [parts per thousand (ppt)]
- pH
- Dissolved oxygen [milligrams per liter (mg/L)]

A summary of the water quality data is provided in Table 2-3. Copies of the field logbook are provided in Appendix D.

# 2.5 SAMPLE COLLECTION, STORAGE, AND TRANSPORT

Upon completion of sample collection, samples were shipped via overnight delivery to the USEPA Region II Laboratory in Edison, NJ for arsenic analyses, to DataChem Laboratories for PCB aroclor analyses, and to E2CR for grain size analyses. Samples were shipped on ice and maintained at 4<sup>0</sup> Celsius. COCs accompanied the samples and documented the dates and times of sample collections for arsenic analyses are included in Appendix A, the grain size analysis results and accompanying COC forms are included in Appendix B, and the analytical results and accompanying COC forms for the PCB aroclor analyses are included in Appendix C. Samples were received at the USEPA laboratory on 14 September 2007 for arsenic analyses, at DataChem

Laboratories on 12 and 14 September and 1 November 2007, and at E2CR on 14 September 2007.

One sample, Almond-Boring-2, was listed on the COC, but was not received by the USEPA laboratory. Two QC samples, BA-beach-MSD and Almond-shore-DUP, were received broken by the USEPA laboratory with no recoverable sample.

# 2.5.1 Surface Water Samples

Surface water samples were collected from ten locations along Blackwater Branch, the Maurice River and Union Lake for arsenic and PCB aroclor analyses. At each location, one surface water sample was collected as a mid-stream, mid-water column sample prior to any disturbance of bottom sediment. The second surface water sample was collected at the same location as above after disturbance of the bottom sediment. This "disturbed" sample was used to simulate potential human exposure to arsenic contaminated surface water with suspended sediment during recreational contact (i.e., beach use, wading, and swimming).

The "disturbance" to the sediment was conducted by wading in the water in a region 0-10 ft upstream of the sampling location for approximately 30 seconds. The "disturbed" water sample was collected from mid-depth of the water column immediately following the disturbance of the bottom sediments.

Water samples were collected using an ISCO peristaltic pump with dedicated Tygon tubing. Water samples were transferred directly to pre-cleaned 250 ml plastic bottles preserved with nitric acid. Samples were kept on ice and maintained at 4<sup>o</sup> Celsius.

# 2.5.2 Shallow Sediment Samples

Two types of shallow sediment samples were collected, including in-stream and nearshore (shore) sediment samples for arsenic and PCB aroclor analyses. The shallow sediment samples were collected using a decontaminated stainless-steel Ponar grab sampler for both arsenic and PCB aroclor analyses. Samples were homogenized in the field using stainless steel bowls and spoons immediately following sample collection. The homogenized sediment samples were then transferred directly to 4 ounce glass jars for arsenic analyses and 9 ounce jars for PCB aroclor analyses; both sample types were kept on ice and maintained at 4<sup>0</sup> Celsius. The stainless steel bowls and spoons were decontaminated following the process described in Section 2.6.

# 2.5.2.1 In-Stream Sediment Samples

In-stream sediment samples were collected from ten locations along Blackwater Branch, the Maurice River and Union Lake. Sediment samples were co-located with surface water samples. The shallow sediment samples were collected from the 0-0.5 ft depth increment beneath the water/sediment interface. These samples were collected at either midstream (river locations) or at a distance of greater than 200 ft from the shoreline (lake locations).

# 2.5.2.2 Nearshore (Shore) Sediment Samples

Nearshore (shore) sediment samples were collected from ten locations along Blackwater Branch, the Maurice River and Union Lake 2-10 ft feet below the waterline. Similar to the shallow sediment sample collection, the shore sediment sample was collected from the 0-0.5 ft depth increment beneath the water/sediment interface and used to simulate potential human exposure to arsenic contaminated sediment during recreational activities/recreational contact (i.e., beach wading, playing in shallow near shore water, a special concern regarding children).

# 2.5.3 Deep Sediment Cores (Borings)

Deeper sediment samples were obtained using a vibracoring system at six locations along Blackwater Branch, the Maurice River, and Union Lake for arsenic and PCB aroclor analyses (Table 2-4). These sediment samples were co-located with water sampling stations, when possible. Sediment core samples were targeted for 10 ft below the sediment surface or until refusal. Due to differing refusal depths at each vibracoring location, each sampling station had different total sample depth. In addition, some locations sampled during the September 2007 Post Remedial Action Sampling met refusal at shallower depths than the May 2006 Baseline Sampling. For the first two feet of the organic sediment layer below the water, samples were analyzed at 1-foot intervals. Following the 0-1 ft and 1-2 ft depth intervals, the depth interval(s) submitted for analysis varied based upon the depth of refusal. Table 2-4 details the sediment core sampling depths by location. Due to equipment refusal, the following core depth interval samples were not recovered in September of 2007 (compared to May 2006): Mill-Boring-4 (4-5 ft), Alliance-Boring-2 (1-2 ft), Alliance-Boring-3 (2-3.3 ft), Almond-Boring-3 (2-2.4 ft), ULB-Boring-2 (1-2 ft), SUL-Boring-2 (1-2 ft), and SUL-Boring-3 (2-3.4 ft).

Sediment samples for the project were collected using a vibracoring system supplied by EA Engineering. The vibracoring system uses an aluminum core barrel with an outside diameter of 3 inches. Coring operations in the Blackwater Branch and Maurice River were conducted by hand using a concrete vibrator (attached to the aluminum barrel) and a tripod for retrieving the core sample. Vibracoring was conducted by lowering the barrel to the sediment surface and vibrating to the required depth. After the core penetrated to a sufficient depth (until refusal), the core barrel was retrieved and brought back to the surface. The core barrel was then capped at both ends, sealed, and labeled.

Cores were collected during each workday and stored in a cooled, insulated container accompanying the field crew. After completion of coring activities, the sediment cores were transported to EA in Sparks, Maryland, where they were logged and sub-sampled for testing. The cores were stored in refrigeration units at EA (maintained at 4°C) until they were processed. Each core was sectioned into depth intervals using a hacksaw with decontaminated stainless steel hacksaw blades. Depth-interval sediment samples from the cores were homogenized using stainless steel bowls and spoons. Homogenized samples were then transferred directly to 9 ounce glass jars and were kept on ice and maintained at 4° Celsius. The stainless steel hacksaw blades, bowls and spoons were decontaminated following the process described in Section 2.6.

#### 2.5.4 Beach Soils

Beach soils were sampled from five locations along the Maurice River and Union Lake for arsenic and PCB aroclor analyses. Figure 2-1 provides the location of the beach sampling points. Sampling points were located at Station 4 (Alliance Beach), Station 5 (Almond Beach), Station 6 ("BareA" Beach), Station 9 (Union Lake Beach), and Station 10 (South End Union Lake Beach).

Samples were collected at the closest shore area adjacent to sediment sampling locations and approximately 6-10 ft above the waterline using a stainless steel spoon/shovel. A grab surface soil sample from a depth of 0-0.5 ft was collected and transferred to a stainless steel bowl and homogenized with a stainless steel spoon. The homogenized sediment samples were then transferred directly to 4 ounce glass jars for arsenic analyses and 9 ounce jars for PCB aroclor analyses; both sample types were kept on ice and maintained at 4<sup>0</sup> Celsius. The stainless steel shovel, bowls, and spoons were decontaminated following the process described in Section 2.6.

# 2.5.5 Equipment Blanks

Equipment blanks were collected to determine the extent of contamination, if any, from the sampling equipment used as part of the project. A total of eight equipment blanks (Tables 2-1 and 2-5) were collected for the September 2007 Post Remedial Action Sampling, which included the following:

- One blank per sampling day (two samples collected) for shallow sediment sampling/beach sampling equipment (i.e., grab sampler and stainless steel bowls and spoons),
- One blank per sampling event/phase (one sample collected) for dedicated water collection equipment (i.e., water pump tubing), and
- One blank per sampling event/phase for dedicated boring equipment (i.e., core liner).

Equipment blanks were collected by pouring deionized water, which is provided by EA's Ecotoxicology Laboratory, over sampling equipment that was decontaminated using the procedure outlined in Section 2.6. The rinsate water was placed in laboratory-prepared containers, submitted to the analytical laboratory, and tested for the same chemical parameters as the sediments and site water. Equipment blanks were sent with the surface water, sediment, and beach soil samples to the USEPA Region II laboratory for arsenic analyses and were submitted to DataChem laboratory for PCB aroclor analyses.

# 2.5.6 Field Duplicates

Field duplicate samples were collected simultaneously from the same sampling locations as sediment and water samples and are used as measures of matrix homogeneity and sampling precision (Table 2-5). A total of nine duplicate samples were collected as individual co-located samples, and they were homogenized separately. Seven field duplicate samples were collected at random locations for sediment and two field duplicate samples were collected at random locations for water.

# 2.5.7 Matrix Spike / Matrix Spike Duplicate Samples

A matrix spike (MS) is a field sample to which a known amount of analyte is added before sample preparation and analysis to evaluate the potential effects of matrix interference. Analyte concentrations in the spiked and unspiked sample are used to calculate percent recovery as a measure of matrix interference. A matrix spike duplicate (MSD) is a duplicate of the MS sample. Additional volumes of sediment and water were collected at random locations and included four sets of MS/MSD for sediment and soil samples and one set of MS/MSD for water samples (Table 2-5).

# 2.6 EQUIPMENT DECONTAMINATION PROCEDURES

Equipment that came into direct contact with sediment and beach soil during sampling was decontaminated prior to deployment in the field to minimize cross-contamination. This included aluminum core liners, core caps, stainless steel spoons, and processing equipment (spoons, knives, bowls, extruder, etc.). While performing the decontamination procedure, phthalate-free nitrile gloves were used to prevent phthalate contamination of the sampling equipment or the samples.

The decontamination procedure is described below:

- Rinse equipment using clean tap or site water
- Wash and scrub with non-phosphate detergent (Alconox or other laboratory-grade detergent)
- Rinse with tap water
- Rinse with 1 percent nitric acid (HNO<sub>3</sub>)
- Rinse with distilled or de-ionized water
- Rinse with methanol followed by hexane
- Rinse with distilled or de-ionized water

Waste liquids were contained during decontamination procedures and transferred to EA's facility in Sparks, Maryland, for disposal.

## 2.7 SAMPLE CHAIN-OF-CUSTODY AND DOCUMENTATION

# 2.7.1 Field Logbook

Field notes were recorded in a permanently bound, dedicated field logbook. A log of sampling activities, station locations, water depths, and core recoveries were recorded in the log in

indelible ink. Personnel names, local weather conditions, and other applicable field sampling program information were also recorded.

Sample location coordinates, approximate water depth, and weather conditions at each sampling location were recorded. In addition, water quality was measured and recorded at each station using an electronic water quality monitoring instrument. Information was recorded in indelible ink. Copies of the project logbook are provided in Appendix D.

# 2.7.2 Sample Identification

A sample numbering system was utilized for the sediment, soil, and water samples. The sample numbering system provided communication between the sample processing operation and the laboratory performing the desired analyses. Surface water, shallow sediment, and beach soil samples were identified by site name, sample type, and date of collection. See table below for sample identification by locations:

Sample Location:	<b>Sample Identification:</b>
1) West of Mill Rd	Mill-
2) West of Rte 55	R55-
3) BWB & Maurice Confluence	BWB-
4) Alliance Beach	Alliance-
5) Almond Beach	Almond-
6) "BareA" Beach	BA-
7) Sherman Ave.	Sherman-
8) North End of Union Lake	NUL-
9) Union Lake Beach	ULB-
10) South End Union Lake Beach	SUL-

The following sample descriptors were then used to denote sample types:

- Shore shallow sediment collected within 2-10 ft below the waterline;
- Sed shallow sediment co-located with the surface water sample;
- Boring deep sediment cores collected from target depth intervals ranging from 0-4 ft;
- Wat1 surface water sample collected prior to sediment and/or core collection;
- Wat2 surface water sample collected after sediment and/or core collection (following agitation and disturbance of the sediments);
- Beach beach soil collected at the closest area adjacent to sediment sampling locations (approximately 6-10 ft above the waterline).

For example, sample Mill-Shore-date (MMDDYY) indicated a shallow sediment sample collected within 2 feet of the shoreline at the station located west of Mill Road. Each sample name was then followed with a date consisting of day, month, and year of sample collection to enable differentiation between future sampling and monitoring events that will be scheduled at the site as part of the remediation activities.

Field Duplicate water and sediment samples were submitted to the laboratory as blind duplicates. The site name and collection date were not designated as part of the sample identifier. Duplicate samples were designated with an identifier (i.e., DUP) and number (i.e., 1, 2, 3, etc.). For example, DUP-1 was designated as the first duplicate sample collected from a random station. DUP-2 was then designated as the next (or second) duplicate sample collected from a separate random station. Locations where duplicate samples were collected and the corresponding sample ID were recorded in the field logbook for future cross-referencing with sample laboratory results. The cross-referenced sampling locations for the field duplicates are included in Table 2-5.

MS/MSD sediment, soil, and water samples were designated with identifiers added after the site name and sample type. For example, Mill-Shore-MS-*date* indicated a matrix spike shoreline sample from the station located West of Mill Road. The following descriptors were used for matrix spike and matrix spike duplicate samples:

- MS matrix spike sample
- MSD matrix spike duplicate

Equipment blanks were identified by type of blank, number of each type, and date (Table 2-5). For example, PBlank-02-date represented the second rinsate blank for the Ponar grab sampler and bowls and spoons used for shallow sediment sampling. The following descriptors were used to denote equipment blanks:

- PBlank Ponar grab sampler and bowls/spoons for shallow sediment samples
- BSBlank Bowls and spoons used for beach sampling
- BrlBlank dedicated aluminum core barrel for core sampling
- TTBlank dedicated tygon tubing blank for water sampling

# 2.7.3 Deep Sediment Core (Borings) Identification

Sediment cores were collected at six locations during the September 2007 Post Remedial Action Sampling. Sediment cores were labeled in the field showing sample location, date, time of collection, and orientation (top and bottom).

Upon processing, the sediment sample IDs from the cores corresponded to the depth intervals (0-1 ft, 1-2 ft, etc.). For example:

- Mill-Boring-1 corresponded to the 0-1-ft interval,
- Mill-Boring-2 corresponded to the 1-2 ft interval,

The sample Mill-Boring-2 corresponded to the second 1-foot interval.

# 2.7.4 Sample Documentation

# 2.7.4.1 Sample Labels

Both the individual sediment cores and the processed sediment were labeled. Sample containers for the processed sediment and water samples were labeled with the following information:

- Client name
- Project number
- Sample ID
- Station location
- Date and time of collection
- Sampler's initials
- Type of analyses required

# 2.7.4.2 Chain-of-Custody Records

Sediment, soil, and water samples collected in the field and at EA's processing facility were documented on a COC form. This COC accompanied the samples to the analytical and geotechnical laboratories. The COC indicated the date and time of sample collection and was signed by appropriate personnel. Copies of the COCs that accompanied the analytical testing for arsenic are provided in Appendix A, the grain size analysis COC forms are included in Appendix B, and the accompanying COC forms for the PCB aroclor analyses are included in Appendix C.

# 2.7.5 Documentation Procedures

Documentation was initialed by the author and dated. Corrections to documentation were made with a single line through the error with the author's initials and date.

# 2.8 ANALYTICAL METHODS

Analytical testing for arsenic was conducted by the USEPA Region II Division of Environmental Science & Assessment (DESA) Laboratory Branch located in Edison, NJ. Analytical testing for PCBs was conducted by DataChem Laboratories located in Salt Lake City, UT. Grain size analyses were conducted by E2CR, a geotechnical firm located in Baltimore, MD.

# 2.8.1 Analytical Methods, Laboratory Quality Control, and Detection Limits

Samples obtained during the September 2007 Post Remedial Action Sampling were analyzed for total arsenic using SW846 3050B/6010B for solids (sediment and soil samples) and SW846 3020A/6010B for water samples. Table 2-1 summarizes analytical information (total number of samples, QA/QC samples, sample volumes, sample holding times, and preservatives) for the project. The target detection limits (TDL)/screening values and laboratory reporting limits (RL) for arsenic in the water and soils were as follows:

Matrix	Target Detection Limit (TDL) / Screening Value for Arsenic	Laboratory Reporting Limit (RL) for Arsenic	
Water	10 ppb (USEPA Drinking Water Criterion)	8 μg/L (ppb)	
Solid	20 ppm (Site Clean-up Level)*	0.73 to 0.8 mg/Kg (ppm)	

<sup>\*</sup>The Site Clean-up Level of 20 ppm is based upon the New Jersey Residential Clean-up Standard for Arsenic.

Copies of the USEPA Region II DESA Laboratory Branch SOPs for sample digestion and for analysis of metals are provided in Appendix E, as well as laboratory Quality Control (QC) and Quality Assurance (QA) procedures.

Samples obtained during the September 2007 Post Remedial Action Sampling were analyzed for PCB aroclors using the SOP Hazardous Waste (HW)-37/AROCLOR, organic analysis of low/medium concentration of aroclor organic compounds SOM01.2. The target detection limits (TDL)/screening values and laboratory reporting limits (RL) for PCB aroclors in the water and sediments/soils were as follows:

Matrix	Target Detection Limit (TDL) / Screening Value for total PCBs	Laboratory Reporting Limit (RL) for total PCBs
Water	0.5 ppb (USEPA Drinking Water Maximum Contaminant Level)	0.5 μg/L (ppb)*
Solid	200 ppb (Soil Remediation Standard)**	17 to 170 μg/Kg (ppb)*

<sup>\*</sup>Based upon 1,000 ml for water and based upon 30 g for soil.

Grain size analysis was conducted using method American Society for Testing and Materials (ASTM) D422 (ASTM 1995) for eight of the ten collected samples from the sediment cores.

# 2.8.2 Data Validation and Electronic Data Deliverables

Data validation was conducted by the USEPA Region II for both the arsenic and PCB aroclor analyses. In addition, USEPA DESA provided Electronic Data Deliverables (EDDs) that comply with Automated Data Review (ADR), and submitted the results directly to USACE-Philadelphia.

<sup>\*\*</sup>The Soil Remediation Standard of 0.2 ppm (200 ppb) is based upon the New Jersey Residential Direct Contact Soil Remediation Standard for total PCBs.

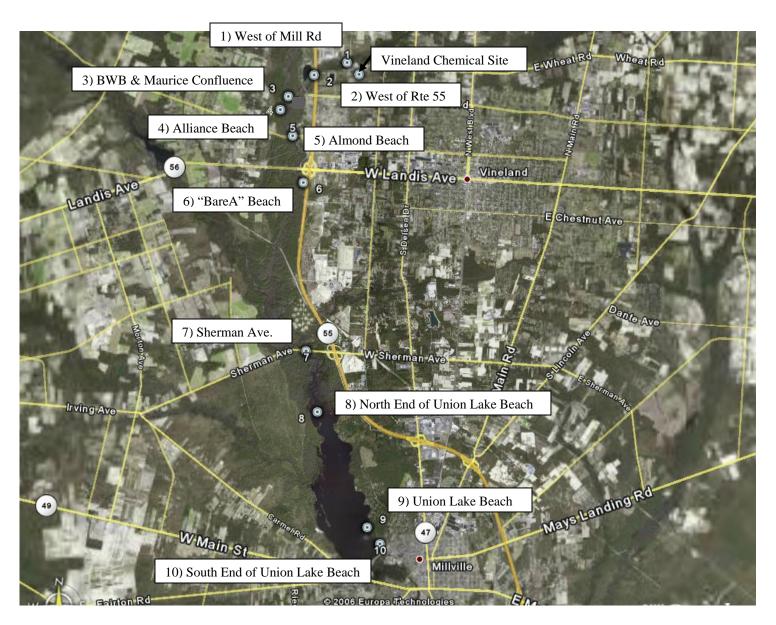


Figure 2-1. Sampling Locations in Vicinity of Vineland Chemical Superfund Site, September 2007

TABLE 2-1. SUMMARY OF SEDIMENT, SOIL, AND WATER SAMPLES COLLECTED IN VICINITY OF VINELAND CHEMICAL SUPERFUND SITE, SEPTEMBER 2007

### VINELAND POST REMEDIAL ACTION SAMPLING AND MONITORING PROGRAM, OPERABLE UNITS #3 AND #4

		Type and Number of Samples				
Sample Location (Name)	Sample Date	Sediment (sed)	Water (wat1/wat2)	Boring±	Beach (beach)	Shore (shore)
		Arsenic/PCB aroclor	Arsenic/PCB aroclor	Arsenic/PCB aroclor	Arsenic/PCB aroclor	Arsenic/PCB aroclor
1) West of Mill Rd (MILL)	9/10/2007	1/1	2/2	3/3	-	1 + MS, MSD/1
2) West of Rte 55 (R55)	9/11/2007	1 + MS, MSD/1	2/2	3/3	-	1/1
3) BWB & Maurice Confluence (BWB)	9/11/2007	1 + DUP/1 + DUP + MS, MSD	2 + MS, MSD/2	-	-	1 + DUP/1+ DUP
4) Alliance Beach (ALLIANCE)	9/11/2007	1/1	2/2	1/1	1 + DUP/1+ DUP	1/1
5) Almond Beach (ALMOND)	9/11/2007	1/1	2/2	2*/2	1/1	1 + DUP*/1+ DUP
6) "BareA" Beach (BA)	9/10/2007	1 + DUP/1+ DUP	2/2	-	1 + MS, MSD*/1	1/1
7) Sherman Ave. (SHERMAN)	9/10/2007	1 + DUP/1 + DUP	2/2	-	-	1/1
8) North End of Union Lake (NUL)	9/12/2007	1/1	2 + DUP/2 + DUP	-	-	1/1
9) Union Lake Beach (ULB)	9/12/2007	1/1	2 + DUP/2 + DUP	1/1	1 + MS, MSD/1	1/1
10) South End Union Lake Beach (SUL)	9/12/2007	1/1	2/2	1/1	1 + DUP/1+ DUP	1/1
NUMBER OF ARSENIC/PCB SAMPLES	-	10/10	20/20	10/11	5/5	10/10
NUMBER OF QC SAMPLES	-	5/4	4/2	0	6/2	4/2
TOTAL NUMBER OF ARSENIC SAMPLES	-	15	24	10	11	14
TOTAL NUMBER OF PCB SAMPLES		14	22	11	7	12
Arsenic Samples:		Sediment	Water	Boring	Beach	Shore
Sample Volume		1 x 250 g	1 x 250 ml	1 x 250 g	1 x 250 g	1 x 250 g
Holding Time		6 months	6 months	6 months	6 months	6 months
Preservative/Temperature		4°C	$HNO_3$ to $pH < 2$ , $4^{\circ}C$	4°C	4°C	4°C
Extraction/Analysis Methodologies		EPA 200.7 Rev 4.4/6010	EPA 200.7 Rev 4.4/6010	EPA 200.7 Rev 4.4/6010	EPA 200.7 Rev 4.4/6010	EPA 200.7 Rev 4.4/6010
PCB Aroclor Samples:		Sediment	Water	Boring	Beach	Shore
Sample Volume		1 x 400 g	1 x 2-L	1 x 400 g	1 x 400 g	1 x 400 g
Holding Time		14 days	7 days	14 days	14 days	14 days
Preservative/Temperature		4°C	4°C	4°C	4°C	4°C
Extraction/Analysis Methodologies		EPA SOM ARO(SOM01.2)	EPA SOM ARO(SOM01.2)	EPA SOM ARO(SOM01.2)	EPA SOM ARO(SOM01.2)	EPA SOM ARO(SOM01.2)

 $\pm Boring\ processing\ date = 9/13/2007$ 

\*Samples were collected but not analyzed by the laboratory due to sample loss or container breakage during shipment.

# NOTE:

QC sample duplicates were collected and analyzed for each media at a rate of approximately 10% per sample matrix per analysis per sample event.

Sediment/soil equipment blanks= 1 blank each per day for beach soil and sediment + 1 blank for dedicated boring equipment

Aqueous equipment blank = 1 blank total for dedicated water pump/tubing

Blank Equipment Samples (8) Collected for Arsenic and for PCB aroclors (7\*\*):

BRLBLANK-01: Dedicated aluminum core barrel for core sampling BSBLANK-01, 02, 03: Bowls and spoons used for beach sampling

PBLANK-01, 02, 03: Ponar grab sampler and bowls/spoons for shallow sediment samples

TTBLANK-01: Dedicated tygon tubing blank for water sampling

<sup>\*\*</sup>BSBLANK-03 not collected for PCB aroclors

# TABLE 2-2. COORDINATES FOR WATER, SOIL, AND SEDIMENT SAMPLES COLLECTED IN VICINITY OF VINELAND CHEMICAL SUPERFUND SITE, SEPTEMBER 2007

# VINELAND POST REMEDIAL ACTION SAMPLING AND MONITORING PROGRAM, OPERABLE UNITS #3 AND #4

Sample Location	Sample Identification	Northing (ft)*	Easting (ft)*
	MILL-SED	247692.5	334013.9
1) West of Mill Rd	MILL-WAT1, 2	247692.5	334013.9
1) West of Mill Ru	MILL-SHORE	247687.2	334024.5
	MILL-BORING-1, 2, 3	247692.5	334013.9
	R55-SED	246768	331373.3
2) West of Dec 55	R55-WAT1, 2	246768	331373.3
2) West of Rte 55	R55-SHORE	246759	331364.9
	R55-BORING-1, 2, 3	246768.2	331358.3
	BWB-SED	244865.4	329111.8
3) BWB & Maurice Confluence	BWB-WAT1, 2	244865.4	329111.8
	BWB-SHORE	244849.9	329105.7
	ALLIANCE-SED	243928.2	328599.8
	ALLIANCE-WAT1, 2	243928.2	328599.8
4) Alliance Beach	ALLIANCE-SHORE	243944.2	328587.6
	ALLIANCE-BEACH	243966.7	328571.7
	ALLIANCE-BORING-1	243928.2	328599.8
	ALMOND-SED	241822.5	329518.8
	ALMOND-WAT1, 2	241822.5	329518.8
5) Almond Beach	ALMOND-SHORE	241833.2	329545
	ALMOND-BEACH	241833.3	329554.9
	ALMOND-BORING-1,2	241822.5	329518.8
	BA-SED	237980.8	330352.4
C) "Dans A" Dansla	BA-WAT1, 2	237980.8	330352.4
6) "BareA" Beach	BA-SHORE	237991.4	330368.4
	BA-BEACH	238011.2	330394.8
	SHERMAN-SED	224390.3	330557
7) Sherman Ave.	SHERMAN-WAT1, 2	224390.3	330557
	SHERMAN-SHORE	224384.3	330561.1
	NUL-SED	219756.6	331231.3
8) North End of Union Lake	NUL-WAT1, 2	219756.6	331231.3
	NUL-SHORE	219729.7	331302.9
	ULB-SED	210474.8	335193.8
	ULB-WAT1, 2	210474.8	335193.8
9) Union Lake Beach	ULB-BEACH	210346.1	335387.4
	ULB-SHORE	210356.3	335358.2
	ULB-BORING-1	210358.9	335205.7
	SUL-SED	208687.7	336375.6
	SUL-WAT1, 2	208687.7	336375.6
10) South End Union Lake Beach	SUL-SHORE	208799.2	336532.1
	SUL-BEACH	208812.2	336555.9
	SUL-BORING-1	208746.2	336485.3

<sup>\*</sup>Coordinates are in NJ State Plane (ft), NAD 83

TABLE 2-3. IN SITU WATER QUALITY MEASUREMENTS COLLECTED IN VICINITY OF VINELAND CHEMICAL SUPERFUND SITE, SEPTEMBER 2007, NOVEMBER 2006, MAY 2006, AND MAY 1999\* VINELAND POST REMEDIAL ACTION SAMPLING AND MONITORING PROGRAM, OPERABLE UNITS #3 AND #4

Sample Location	Date and Time of Sample	Sample Depth	Water Temperature (degrees C)	Salinity (ppt)	Dissolved Oxygen (mg/L)	рН	Conductivity (mS/cm)
1) West of Mill Rd	9/10/07 1730	Surface	22.3	0.06	8.9	7.6	0.121
2) West of Rte 55	9/11/07 0943	Surface	18.8	0.09	8.5	8.0	0.195
3) BWB & Maurice Confluence	9/11/07 1200	Surface	22.7	0.06	7.4	7.7	0.138
4) Alliance Beach	9/11/07 1404	Surface	22.9	0.07	7.3	7.5	0.140
5) Almond Beach	9/11/07 1538	Surface	22.8	0.07	7.4	7.2	0.140
6) "BareA" Beach	9/10/2007 1420	Surface	23.8	0.07	8.1	7.8	0.150
7) Sherman Ave.	9/10/07 1530	Surface	22.7	0.09	7.4	7.2	0.193
8) North End of Union Lake	9/12/07 1121	Surface	20.3	0.08	7.3	7.7	0.176
9) Union Lake Beach	9/12/07 1214	Surface	25.2	0.07	7.0	7.6	0.159
10) South End of Union Lake Beach	9/12/2007 1447	Surface	25.3	0.07	7.2	7.6	0.160
,					i		
1) West of Mill Rd	11/02/06 0855	Surface	12.9	0.04	5.4	6.7	0.073
2) West of Rte 55	11/02/06 0940	Surface	12.8	0.05	7.6	6.6	0.089
3) BWB & Maurice Confluence	11/02/06 1047	Surface	12.3	0.04	7.6	6.1	0.068
4) Alliance Beach	11/02/06 1144	Surface	12.5	0.04	7.9	6.2	0.069
5) Almond Beach	11/01/06 1655	Surface	13.0	0.04	5.5	6.0	0.070
6) "BareA" Beach	11/01/06 1600	Surface	12.9	0.04	5.6	6.3	0.071
7) Sherman Ave.	11/01/06 1443	Surface	13.1	0.06	5.8	7.0	0.106
8) North End of Union Lake	11/01/06 1140	Surface	12.3	0.06	6.0	6.7	0.092
9) Union Lake Beach	11/01/06 1230	Surface	11.3	0.06	6.4	7.7	0.087
10) South End of Union Lake Beach	11/01/06 1300	Surface	12.9	0.06	6.8	8.3	0.093
1) West of Mill Rd	5/23/06 0840*	Surface	12.7	0.08	9.8	7.1	0.137
2) West of Rte 55	5/23/06 1808*	Surface	17.4	0.08	10.2	6.9	0.145
3) BWB & Maurice Confluence	5/25/06 0849*	Surface	17	0.05	8.8	7.2	0.098
4) Alliance Beach	5/23/06 1340*	Surface	17	0.07	9.9	6.9	0.126
5) Almond Beach	5/23/06 1453*	Surface	17.6	0.07	10.3	6.9	0.123
6) "BareA" Beach	5/23/06 1535*	Surface	17.7	0.05	10.1	6.8	0.101
7) Sherman Ave.	5/23/06 1710*	Surface	17.7	0.08	9.5	7.1	0.148
8) North End of Union Lake	5/24/06 1157*	Surface	15.8	0.07	10	6.9	0.116
9) Union Lake Beach	5/24/06 1318*	Surface	19	0.06	10.1	7.3	0.117
10) South End of Union Lake Beach	5/24/06 1642*	Surface	20	0.06	10.5	7.7	0.111
4) Alliance Beach	5/1999**	unknown	15.1	0.0	8.9	6.3	0.079
5) Almond Beach	5/1999**	unknown	13.5	0.0	8.7	6.4	0.077
6) "BareA" Beach	5/1999**	unknown	14.6	0.0	9.7	5.8	0.079
9) Union Lake Beach	5/1999**	unknown	16.9	0.0	9.7	5.7	0.091
10) South End of Union Lake Beach	5/1999**	unknown	15.3	0.0	9.4	5.9	0.093

<sup>\*\*</sup>May 1999 data were referenced from (USEPA /ERTC 1999) citation

### TABLE 2-4. SUMMARY OF SEDIMENT CORE SAMPLES COLLECTED IN VICINITY OF VINELAND CHEMICAL SUPERFUND SITE, SEPTEMBER 2007 VINELAND POST REMEDIAL ACTION SAMPLING AND MONITORING PROGRAM, OPERABLE UNITS #3 AND #4

Sample Location	Date	Time	Sample IDs	Corresponding Sample Depths
	9/13/07	1300	Mill-Boring-1*	0-1 ft
1) West of Mill Rd	9/13/07	1301	Mill-Boring-2	1-2 ft
	9/13/07	1302	Mill-Boring-3*	2-2.9 ft
	9/13/07	1215	R55-Boring-1*	0-1 ft
2) West of Rte 55	9/13/07	1216	R55-Boring-2	1-2 ft
	9/13/07	1217	R55-Boring-3*	2-4 ft
4) Alliance Beach	9/13/07	1240	Alliance-Boring-1*	0-1 ft
5) Almond Beach	9/13/07	1155	Almond-Boring-1*	0-1 ft
9) Union Lake Beach	9/13/07	1250	ULB-Boring-1*	0-1 ft
10) South End Union Lake Beach	9/13/07	1128	SUL-Boring-1*	0-1 ft

<sup>\*</sup>denotes depth interval submitted for grain size analysis

# TABLE 2-5. DUPLICATE, EQUIPMENT BLANK, AND MATRIX SPIKE SAMPLES COLLECTED IN VICINITY OF VINELAND CHEMICAL SUPERFUND SITE,

### **SEPTEMBER 2007**

### VINELAND POST REMEDIAL ACTION SAMPLING AND MONITORING PROGRAM, OPERABLE UNITS

	Arsenic and PCB	Aroclor Station ID		
Sample Location	CI- ID	<b>Duplicate Cross-</b>	Date Collected	Matrix
_	Sample ID	Referenced Station		
	DUP-1	NUL-Wat1	9/12/2007	Aqueous
	DUP-2	ULB-Wat2	9/12/2007	Aqueous
	DUP-3	SUL-Beach	9/12/2007	Sediment
	DUP-4	Sherman-Sed	9/10/07	Sediment
Field Duplicate	DUP-5	BA-Sed	9/10/07	Sediment
	DUP-6	Almond-Shore	9/11/2007	Sediment
	DUP-7	BWB-Shore	9/11/2007	Sediment
	DUP-8	BWB-Sed	9/11/2007	Sediment
	DUP-9	Alliance-Beach	9/11/2007	Sediment
	PBlank-01*	N/A	9/10/07	Aqueous
	PBlank-02*	N/A	9/11/2007	Aqueous
	PBlank-03*	N/A	9/12/2007	Aqueous
Equipment Plenk	BSBlank-01*	N/A	9/10/07	Aqueous
Equipment Blank	BSBlank-02*	N/A	9/11/2007	Aqueous
	BSBlank-03	N/A	9/12/2007	Aqueous
	BrlBlank-01*	N/A	9/12/2007	Aqueous
	TTBlank-01*	N/A	9/12/2007	Aqueous
	Mill-Shore-MS	N/A	9/10/07	Sediment
	Mill-Shore-MSD	N/A	9/10/07	Sediment
	R55-Sed-MS	N/A	9/11/2007	Sediment
	R55-Sed-MSD	N/A	9/11/2007	Sediment
Matrix Spikes (MS) and Matrix	BWB-Wat2-MS	N/A	9/11/2007	Aqueous
Spike Duplicates (MSD)**	BWB-Wat2-MSD	N/A	9/11/2007	Aqueous
	BA-Beach-MS	N/A	9/10/07	Sediment
	BA-Beach-MSD	N/A	9/10/07	Sediment
	ULB-Beach-MS	N/A	9/12/2007	Sediment
	ULB-Beach-MSD	N/A	9/12/2007	Sediment

<sup>\*</sup>Equipment blanks collected for PCB aroclors

<sup>\*\*</sup>For PCB aroclor analyses, the MS/MSD sample was collected at station BWB-sed

### 3. ARSENIC RESULTS

The September 2007 Post Remedial Action Sampling arsenic results for each station and sampling matrix are presented in Figure 3-1. Arsenic results by individual station locations are presented in Figures 3-2 through 3-8. Detected arsenic concentrations in water samples were compared to the USEPA Drinking Water Criterion for arsenic of 10 parts per billion (ppb or µg/L), and the results for detected arsenic concentrations in sediment and soil were compared to the Site Clean-up Level of 20 parts per million (ppm or mg/Kg). The Site Clean-up Level of 20 ppm is based upon the New Jersey Residential Clean-up Standard for Arsenic. The laboratory report and the accompanying COC forms are provided in Appendix A. Appendix C and D present the SOPs for the laboratory analyses.

### 3.1 WATER

### 3.1.1 *In Situ* Water Quality

Water quality measurements were recorded *in situ* at each of the ten locations where water samples were collected for chemical analysis (mid-stream/mid-depth of the water column). As stated previously in Section 2, the *in situ* water quality data recorded in September 2007 are presented in Table 2-3. Table 2-3 also includes *in situ* water quality data collected during the November 2006 (EA 2007), May 2006 (EA 2006a), and May 1999 (USEPA/ERTC 1999) field collection at the site for comparison. The *in situ* water quality results were within the expected range of parameters for a freshwater system in New Jersey. Water temperature ranged from 18.8 to 25.3 degrees Celsius, the salinity ranged from 0.06 to 0.09 parts per thousand (ppt), dissolved oxygen ranged from 7.0 to 8.9 mg per liter (mg/L), pH ranged from 7.0 to 8.9, and the conductivity ranged from 0.121 to 0.195 mS/cm. As seen in Table 2-3 and with the exception of seasonal temperature and DO concentrations, the water quality parameters collected in September 2007 were similar to the water quality parameters recorded at the same locations in November 2006, May 2006, and May 1999.

#### 3.1.2 Surface Water

Surface water samples were collected from ten locations along Blackwater Branch, the Maurice River, and Union Lake (Figures 3-1 through 3-8). At each location, one surface water sample was collected prior to any disturbance of bottom sediment (referred to as sample one – Wat1) and the second surface water sample was collected at the same location as above after disturbance of the bottom sediment (referred to as sample two – Wat2), intended to simulate potential human exposure to arsenic during recreational contact. Therefore, a total of 20 surface water samples were collected from ten locations in the vicinity of the site. Ten of the 20 surface water samples analyzed had detected concentrations of arsenic that were above the 10 ppb (μg/L) USEPA Drinking Water Criterion for arsenic. Seven of the ten exceedences occurred in the agitated water samples, with arsenic concentrations ranging from 15 ppb at Station 9 (Union Lake Beach) to 3,800 at Station 1 (West of Mill Rd). The other three surface water exceedences occurred at Station 1 (West of Mill Rd), Station 2 (West of Rte 55), and Station 6 ("BareA" Beach), with concentrations ranging from 12 ppb to 14 ppb. At Station 1 (West of Mill Rd), Station 5 (Almond Beach), Station 8 (North End of Union Lake), Station 9 (Union Lake Beach)

and Station 10 (South End of Union Lake), the surface water sample (Wat1) had undetected concentrations of arsenic and the agitated water sample (Wat2) had detected concentrations of arsenic above the 10 ppb ( $\mu$ g/L) USEPA Drinking Water Criterion for arsenic. At Station 2 (West of Rte 55) and Station 7 (Sherman Ave), arsenic was detected in both water samples (surface and agitated water samples) above the 10 ppb USEPA Drinking Water Criterion for arsenic. Arsenic was not detected in surface water of in the agitated water samples at Station 3 (BWB & Maurice Confluence), and Station 4 (Alliance Beach), or in any of the equipment blanks (Table 3-1).

### 3.2 SEDIMENT AND SOIL

### 3.2.1 Shallow Sediment

Two types of shallow and nearshore (shore) sediment samples were collected, including instream and nearshore (shore) sediment samples (Figures 3-1 through 3-8). The results are presented in the following paragraphs and Table 3-2. The shallow sediment samples were intended to simulate potential human exposure to arsenic contaminated sediment during recreational contact (i.e., beach wading, playing in shallow near shore water, a special concern regarding children).

### 3.2.1.1 In-Stream Sediment

In-stream sediment samples (0-0.5 ft depth increment beneath the water/sediment interface) were collected from ten locations along Blackwater Branch, the Maurice River, and Union Lake (Figures 3-1 through 3-8). Arsenic was detected in the shallow sediment samples for nine of the ten locations, ranging from 1.3 mg/Kg to 1,700 mg/Kg (Table 3-2). At four locations, in-stream arsenic concentrations in sediment exceeded the Site Clean-up Level of 20 ppm (mg/Kg): Station 2 (West of Rte 55), Station 7 (Sherman Ave), Station 8 (North End of Union Lake), and Station 9 (Union Lake Beach). Arsenic was not detected in shallow sediment at Station 4 (Alliance Beach).

### 3.2.1.2 Nearshore (Shore) Sediment

Nearshore (shore) sediment samples were collected from ten locations along Blackwater Branch, the Maurice River and Union Lake, 2-10 ft below the waterline, representing the 0-0.5 ft depth increment beneath the water/sediment interface (Figures 3-1 through 3-8). Arsenic was detected in eight of the ten shore sediment samples, ranging from 2 mg/Kg to 3,400 mg/Kg (Table 3-2). At four locations, nearshore arsenic concentrations in sediment exceeded the Site Clean-up Level of 20 ppm for arsenic in solids. Arsenic exceeded the 20 ppm criterion at Station 1 (West of Mill Rd), Station 2 (West of Rte 55), Station 6 ("BareA" Beach) and Station 8 (North End of Union Lake).

### **3.2.2** Deep Sediment Cores (Borings)

### 3.2.2.1 Grain Size Analysis

Grain size analysis was conducted for eight of the ten samples collected from the sediment cores (one sample from each of four coring locations and two samples from each of two locations). Table 3-3 and Figure 3-9 presents the results of the particle size distribution for each of the five stations where sediment cores were collected. The grain size analyses indicate that seven of the eight sediment core samples were predominantly comprised of sand (> 70 %). One sediment core, R55-Boring-1 (0-1 ft), was predominantly comprised of silt and clay (88%).

### **3.2.2.2** Arsenic Concentrations

A total of ten depth interval sediment samples were obtained from six locations along Blackwater Branch, the Maurice River, and Union Lake (Figures 3-1 through 3-4 and Figure 3-8). Three depth intervals were analyzed for Station 1 (West of Mill Rd) and Station 2 (West of Rte. 55); the first depth interval (0-1 ft) was analyzed for Station 4 (Alliance Beach), Station 5 (Almond Beach), Station 9 (Union Lake Beach), and Station 10 (South End of Union Lake Beach). Hard-packed sand precluded the collection of deeper sediments due to refusal of the coring device. Arsenic concentrations were low (either not detected or substantially below the Site Clean-up Level of 20 ppm) in the 0-1 ft interval for Station 4 (Alliance Beach), Station 5 (Almond Beach), Station 9 (Union Lake Beach), and Station 10 (South End of Union Lake Beach). Station 1 (West of Mill Rd) and Station 2 (West of Rte. 55) had arsenic concentrations that exceeded the Site Clean-up Level of 20 ppm in the 0-1 ft and 1-2 ft depth intervals; the samples greater than 2 ft in depth at these stations exhibited arsenic concentrations below the Site Clean-up Level of 20 ppm. Grain size analysis indicated that sand dominated the first (0-1 ft) depth interval at Station 1(West of Mill Rd), while fine grained material (silts/clays) dominated the first (0-1 ft) depth interval at Station 2 (West of Rte. 55). The high concentration of arsenic at Station 2 may be correlated to the silt content. Arsenic is strongly sorbed onto soils and sediments, including silt (Bodek et. al 1988).

### 3.2.3 Beach Soils

Beach soils were sampled from a total of five locations along the Maurice River and Union Lake (Figures 3-1, 3-3, 3-4, 3-5, and 3-8). Arsenic was only detected at one of the five beach locations (Station 9 - Union Lake Beach) at a concentration of 1.0 mg/Kg, which is well below the Site Clean-up Level of 20 ppm for arsenic in solids (Table 3-2).

### 3.3 QA/QC RESULTS

The results for the QA/QC samples, including equipment blanks and field duplicates are provided in Tables 3-1 and 3-2, respectively and are discussed in the following subsections.

### 3.3.1 Equipment Blanks

Arsenic was not detected at concentrations above the Method Detection Limit (MDL -  $8 \mu g/L$ ) in any of the equipment blanks. Therefore, it is unlikely that any contamination can be attributed to sampling equipment or collection and handling.

### 3.3.2 Field Duplicates

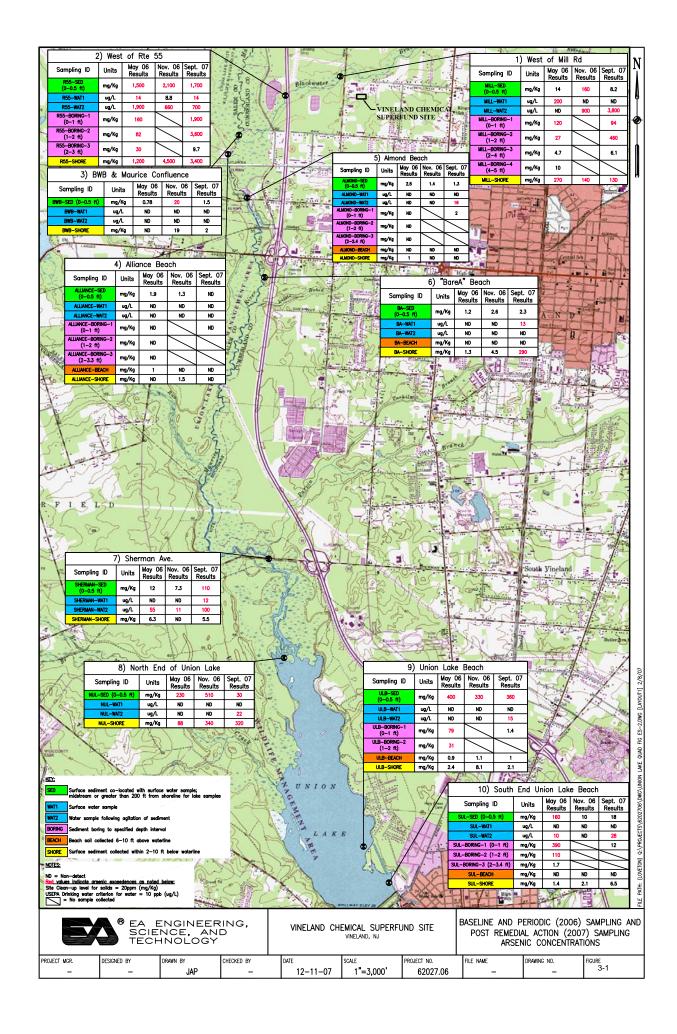
Field duplicate samples were collected simultaneously from the same sampling locations as sediment and water samples. The two field duplicate samples collected at random locations for water had arsenic results equivalent to the corresponding co-located water samples. Similarly, the seven field duplicate samples that were collected at random locations for sediment were also had arsenic results nearly equivalent to the corresponding co-located sediment samples.

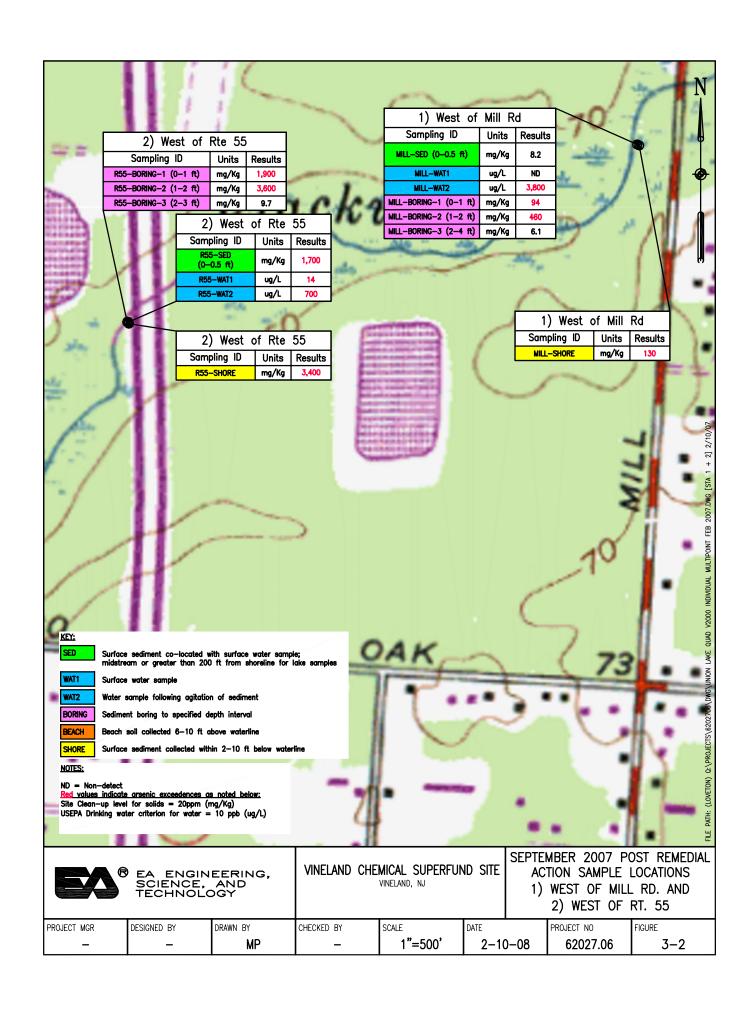
Type of Sample	<b>Duplicate # / Result</b>	Matching Sampling ID / Result
Water	$\overline{\text{DUP-1}/(\text{U})}$	NUL-wat1 / (U)
Water	$DUP-2 / (270 \mu g/L)$	ULB-wat2 / $(15 \mu g/L)$
Soil	DUP-3 / (U)	SUL-beach / (U)
Sediment	DUP-4 / (140  mg/Kg)	Sherman-sed / (110 mg/Kg)
Sediment	DUP-5 / (1.2  mg/Kg)	BA-sed / $(2.3  mg/Kg)$
Sediment	DUP-6 / (U)	Almond-shore / (U)
Sediment	DUP-7 / (2.5  mg/Kg)	BWB-shore / (2 mg/Kg)
Sediment	DUP-8 / (U)	BWB-sed / $(1.5 mg/Kg)$
Soil	DUP-9 / (U)	Alliance-beach / (U)
77 7		

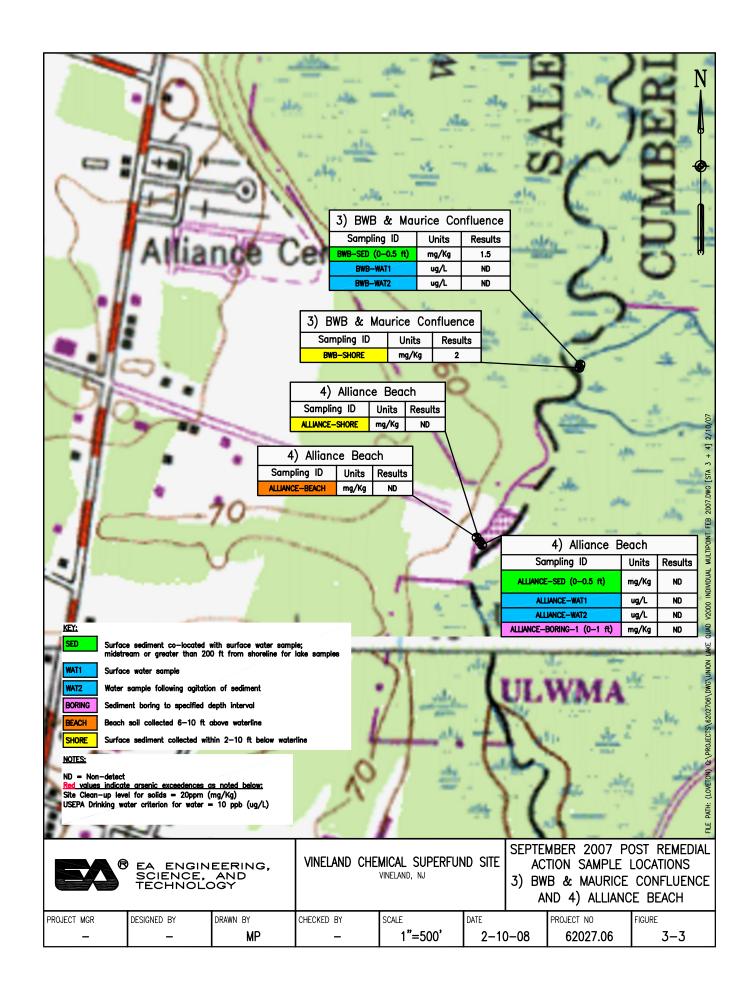
U = undetected arsenic concentration

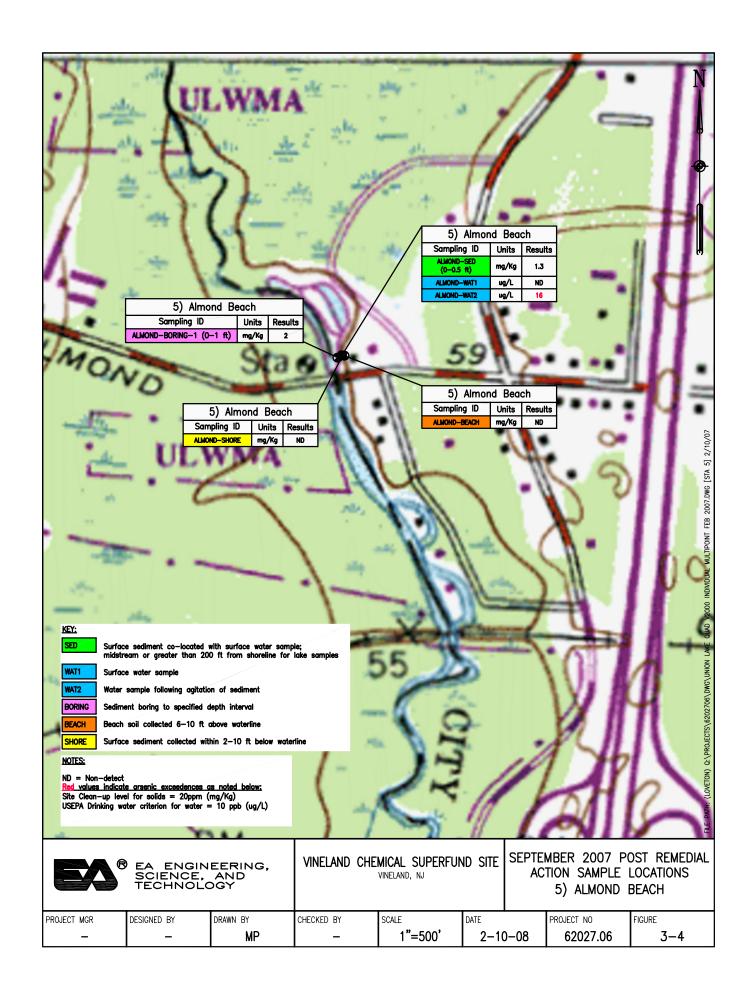
### 3.3.3 MS/MSD Samples

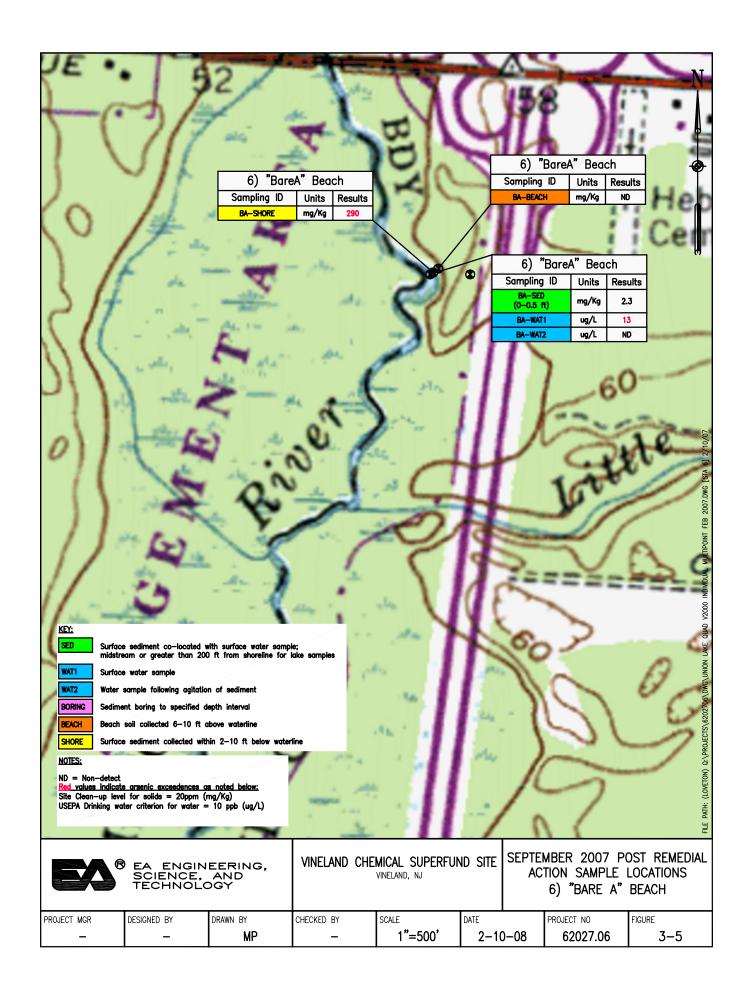
The Laboratory's established QC criteria were met for MS and MSD samples, including aqueous samples, soil samples, and sediment samples. These data were validated by the USEPA Region II DESA Laboratory.

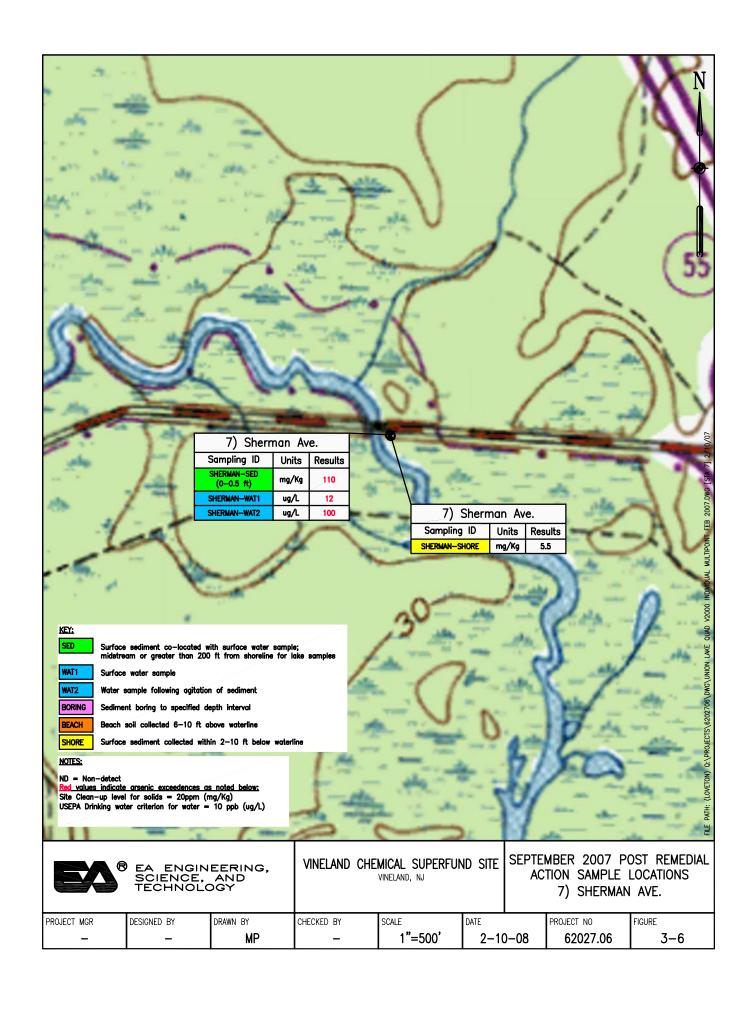


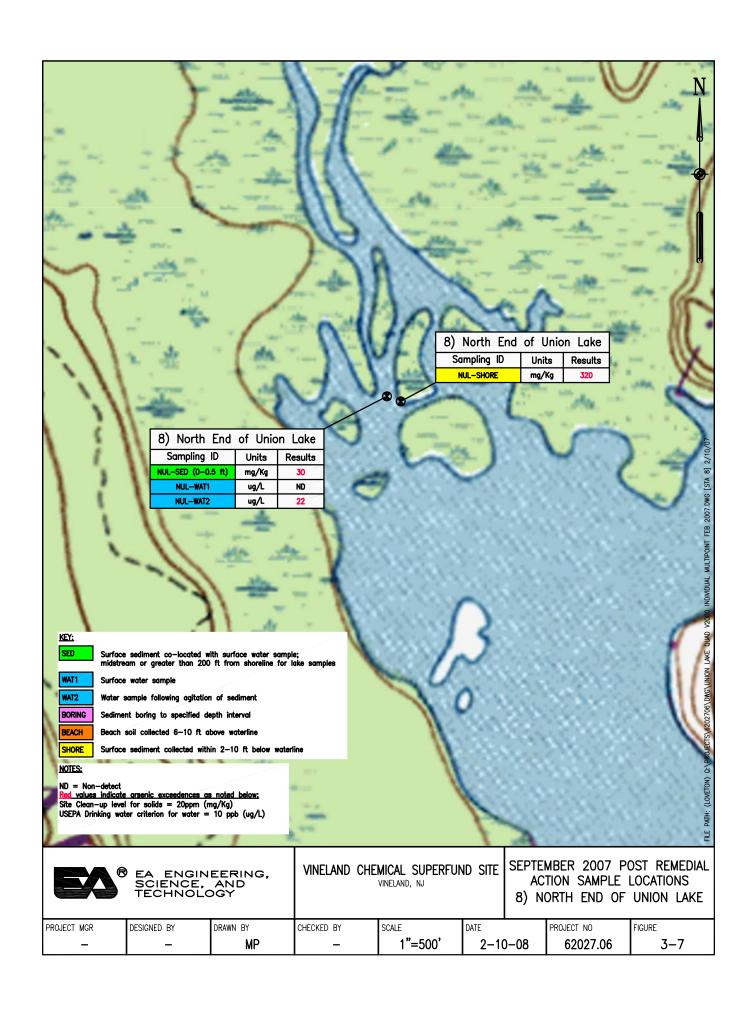


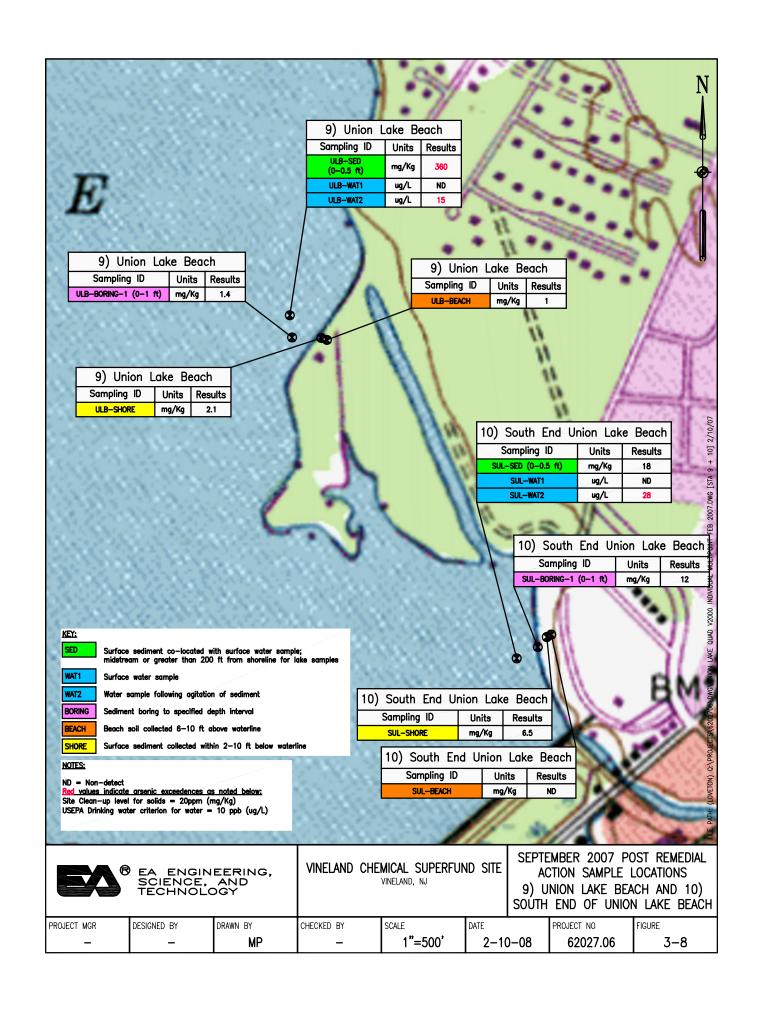












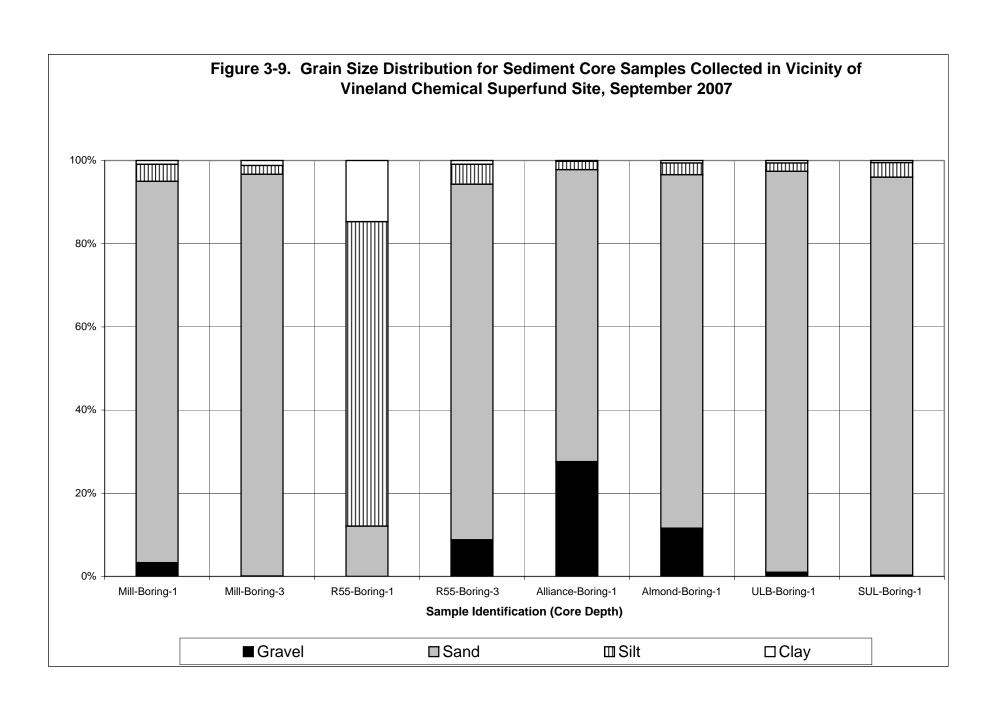


TABLE 3-1. ARSENIC RESULTS IN WATER SAMPLES COLLECTED IN VICINITY OF VINELAND CHEMICAL SUPERFUND SITE, SEPTEMBER 2007

Sample Location	Station ID	Units	Average RL	Result
1) West of Mill Rd	MILL-WAT1	ug/L	8	U
1) West of Mili Rd	MILL-WAT2	ug/L		3,800
2) West of Rte 55	R55-WAT1	ug/L		14
2) West of Rie 33	R55-WAT2	ug/L		700
3) BWB & Maurice Confluence	BWB-WAT1	ug/L	8	U
3) BWB & Maurice Confidence	BWB-WAT2	ug/L	8	U
1) Allianas Basah	ALLIANCE-WAT1	ug/L	8	U
4) Amance Beach	ALLIANCE-WAT2	ug/L	8	U
5) Almond Doogh	ALMOND-WAT1	ug/L	8	U
3) Almond Beach	ALMOND-WAT2	ug/L		16
6) "Doro A" Doooh	BA-WAT1	ug/L		13
o) BareA Beach	BA-WAT2	ug/L	8	U
7) Sharman Ava	SHERMAN-WAT1	ug/L		12
7) Sherman Ave.	SHERMAN-WAT2	ug/L		100
9) North End of Union Lake	NUL-WAT1	ug/L	8	U
8) North End of Official Lake	NUL-WAT2	ug/L		22
0) Union Laka Basah	ULB-WAT1	ug/L	8	U
9) Ullon Lake Beach	ULB-WAT2	ug/L		15
10) Couth End Union Lake Deach	SUL-WAT1	ug/L	8	U
10) South End Union Lake Beach	SUL-WAT2	ug/L		28
Dumlicates	DUP-1 (NUL-WAT1)*	ug/L	8	U
Duplicates	DUP-2 (ULB-WAT2)*	ug/L		270
	PBlank-01	ug/L	8	U
	PBlank-02	ug/L	8	U
	PBlank-03	ug/L	8	U
Alliance Beach Almond Beach BareA" Beach Sherman Ave. North End of Union Lake Union Lake Beach South End Union Lake Beach plicates	BSBlank-01	ug/L	8	U
Equipment brank	BSBlank-02	ug/L	8	U
	BSBlank-03	ug/L	8	U
	BrlBlank-01	ug/L	8	U
	TTBlank-01	ug/L	8	U

**RL** = laboratory reporting limit (average)

NOTE: Bold values represent detected arsenic concentrations; shaded values exceed the USEPA Drinking Water Criterion of 10 ug/L (ppb) U = arsenic was analyzed, but not detected

<sup>\*</sup>Denotes cross-referenced sample location of blind duplicate sample

TABLE 3-2. ARSENIC RESULTS IN SOIL AND SEDIMENT SAMPLES COLLECTED IN VICINITY OF VINELAND CHEMICAL SUPERFUND SITE, SEPTEMBER 2007

Sample Location	Station ID	Units	Matrix	Average RL	Result
	MILL-SED	mg/Kg	Sediment		8.2
	MILL-SHORE	mg/Kg	Sediment		130
1) West of Mill Rd	MILL-BORING-1	mg/Kg	Sediment		94
	MILL-BORING-2	mg/Kg	Sediment		460
	MILL-BORING-3	mg/Kg	Sediment		6.1
	R55-SED	mg/Kg	Sediment		1,700
	R55-SHORE	mg/Kg	Sediment		3,400
2) West of Rte 55	R55-BORING-1	mg/Kg	Sediment		1,900
	R55-BORING-2	mg/Kg	Sediment		3,600
	R55-BORING-3	mg/Kg	Sediment		9.7
2) DWD 9 Marris Conflant	BWB-SED	mg/Kg	Sediment		1.5
3) BWB & Maurice Confluence	BWB-SHORE	mg/Kg	Sediment		2
	ALLIANCE-BEACH	mg/Kg	Soil	0.75	U
4) A11' D1-	ALLIANCE-SED	mg/Kg	Sediment	0.72	U
4) Alliance Beach	ALLIANCE-SHORE	mg/Kg	Sediment	0.78	U
	ALLIANCE-BORING-1	mg/Kg	Sediment	0.77	U
	ALMOND-BEACH	mg/Kg	Soil	0.78	U
5) Al., 1 D 1	ALMOND-SED	mg/Kg	Sediment		1.3
5) Almond Beach	ALMOND-SHORE	mg/Kg	Sediment	0.74	U
	ALMOND-BORING-1	mg/Kg	Sediment		2
	BA-BEACH	mg/Kg	Soil	0.76	U
6) "BareA" Beach	BA-SED	mg/Kg	Sediment		2.3
	BA-SHORE	mg/Kg	Sediment		290
7) Cl	SHERMAN-SED	mg/Kg	Sediment		110
7) Sherman Ave.	SHERMAN-SHORE	mg/Kg	Sediment		5.5
8) North End of Union Lake	NUL-SED	mg/Kg	Sediment		30
8) North End of Union Lake	NUL-SHORE	mg/Kg	Sediment		320
	ULB-BEACH	mg/Kg	Soil		1
9) Union Lake Beach	ULB-SED	mg/Kg	Sediment		360
9) Union Lake Beach	ULB-SHORE	mg/Kg	Sediment		2.1
	ULB-BORING-1	mg/Kg	Sediment		1.4
	SUL-BEACH	mg/Kg	Soil	0.8	U
10) South End Union Lake Beach	SUL-SED	mg/Kg	Sediment		18
10) South End Union Lake Beach	SUL-SHORE	mg/Kg	Sediment		6.5
	SUL-BORING-1	mg/Kg	Sediment		12
	DUP-3 (SUL-BEACH)*	mg/Kg	Soil	0.77	U
	DUP-4 (SHERMAN-SED)*	mg/Kg	Sediment		140
	DUP-5 (BA-SED)*	mg/Kg	Sediment		1.2
Duplicate	DUP-6 (ALMOND-SHORE)*	mg/Kg	Sediment	0.72	U
	DUP-7 (BWB-SHORE)*	mg/Kg	Sediment		2.5
	DUP-8 (BWB-SED)*	mg/Kg	Sediment	0.78	U
	DUP-9 (ALLIANCE-BEACH)*	mg/Kg	Soil	0.73	U

RL = laboratory reporting limit (average)

NOTE: Bold values represent detected arsenic concentrations; shaded values are equivalent to or exceed the Site Clean-up Level of 20 mg/Kg (ppm)
U = arsenic was analyzed, but not detected

<sup>\*</sup>Denotes cross-referenced sample location of blind duplicate sample

TABLE 3-3. PARTICLE SIZE DISTRIBUTION FOR SEDIMENT CORES COLLECTED IN VICINITY OF VINELAND CHEMICAL SUPERFUND SITE, SEPTEMBER 2007

Sample IDs (Core Depth)		Particle	e Size (%)		Dominant Material
Sample IDs (Core Deptil)	Gravel	Sand	Silt	Clay	Dominant Material
Mill-Boring-1	3.3	91.7	4.1	0.9	Sand
Mill-Boring-3	0.1	96.6	2.1	1.2	Sand
R55-Boring-1	0	12.1	73.2	14.7	Silt
R55-Boring-3	8.8	85.5	4.8	0.9	Sand
Alliance-Boring-1	27.6	70.2	2	0.2	Sand
Almond-Boring-1	11.6	85	2.8	0.6	Sand
ULB-Boring-1	1	96.4	2	0.6	Sand
SUL-Boring-1	0.3	95.7	3.5	0.5	Sand

#### 4. PCB RESULTS

The September 2007 Post Remedial Action Sampling included the analysis of nine PCB aroclors for 29 total aqueous samples (including 20 water samples, seven aqueous equipment blanks, and two aqueous field duplicates) and for 43 total soil/sediment samples (36 soil/sediment samples and seven field duplicates). The aroclor concentrations were summed [Non-detect (ND) = 0] and the total PCB concentrations in water samples were compared to the USEPA Drinking Water Maximum Contaminant Level for total PCBs of 0.5 (ppb or  $\mu g/L$ ), and the total PCB concentrations in sediment and soil were compared to the Soil Remediation Standard of 0.2 (ppm or  $\mu g/Kg$ ; 200 ppb or  $\mu g/Kg$ ) for total PCBs in solids. The Soil Remediation Standard of 0.2 ppm is based upon the New Jersey Residential Direct Contact Soil Remediation Standard for total PCBs. The analytical results report for total PCBs and the accompanying COC forms are provided in Appendix C. Appendix E presents the SOPs for the laboratory analyses of PCB aroclors.

### 4.1 WATER

None of the 20 surface water samples analyzed had detected concentrations of PCB aroclors (Table 4-1). Therefore, none of the agitated water samples collected had a total PCB concentration that exceeded the 0.5 ppb ( $\mu g/L$ ) USEPA Drinking Water Maximum Contaminant Level for total PCBs.

### 4.2 SEDIMENT AND SOIL

A total of 36 sediment and soil samples were collected and submitted for analysis of PCB aroclors (Table 4-2). Two of the 36 samples, boring-2 and boring-3 at Station 2 (West of Rte 55), had detected concentrations for Aroclor-1254 of 9.4 and 4.1 ppb ( $\mu g/Kg$ ), respectively. However, neither of the concentrations exceeded the 200 ppb ( $\mu g/Kg$ ) Soil Remediation Standard for total PCBs.

### 4.3 QA/QC RESULTS

For the aqueous samples, none of the seven blank samples or the two field duplicates had detected concentrations of PCB aroclors. For the sediment and soil samples, none of the seven field duplicates had detected concentrations of PCB aroclors. The Laboratory's established QC criteria were evaluated during the data validation process for MS and MSD samples, including those for aqueous samples, soil samples, and sediment samples. One sediment sample, R55-Sed, received an "R" qualifier, which indicated that the concentration was rejected during QC/validation analysis and that the presence or absence of PCB aroclors could not be verified.

TABLE 4-1. PCB AROCLOR CONCENTRATIONS (ug/L) AND TOTAL PCBs\*\* IN WATER SAMPLES COLLECTED IN VICINITY OF VINELAND CHEMICAL SUPERFUND SITE, SEPTEMBER 2007

								R	ESULTS				
Sample Location	Station ID	Units	$\mathbf{RL}$	Aroclor-	Total PCBs								
				1016	1221	1232	1242	1248	1254	1260	1262	1268	(ND = 0)
1) West of Mill Rd	MILL-WAT1	ug/L	0.5	0.5 U	0								
1) West of Willi Ku	MILL-WAT2	ug/L	0.5	0.5 U	0								
2) West of Rte 55	R55-WAT1	ug/L	0.5	0.5 U	0								
2) West of Rie 33	R55-WAT2	ug/L	0.5	0.5 U	0								
3) BWB & Maurice Confluence	BWB-WAT1	ug/L	0.5	0.5 U	0								
3) BWB & Maurice Confluence	BWB-WAT2	ug/L	0.5	0.5 U	0								
4) Alliance Beach	ALLIANCE-WAT1	ug/L	0.5	0.5 U	0								
4) Amance Beach	ALLIANCE-WAT2	ug/L	0.5	0.5 U	0								
5) Almond Beach	ALMOND-WAT1	ug/L	0.5	0.5 U	0								
3) Almond Beach	ALMOND-WAT2	ug/L	0.5	0.5 U	0								
6) "BareA" Beach	BA-WAT1	ug/L	0.5	0.5 U	0								
bareA beach	BA-WAT2	ug/L	0.5	0.5 U	0								
7) Sherman Ave.	SHERMAN-WAT1	ug/L	0.5	0.5 U	0								
// Sherman Ave.	SHERMAN-WAT2	ug/L	0.5	0.5 U	0								
8) North End of Union Lake	NUL-WAT1-120907	ug/L	0.5	0.5 U	0								
North End of Union Lake	NUL-WAT2-120907	ug/L	0.5	0.5 U	0								
9) Union Lake Beach	ULB-WAT1-120907	ug/L	0.5	0.5 U	0								
9) Ollion Lake Beach	ULB-WAT2-120907	ug/L	0.5	0.5 U	0								
10) South End Union Lake Beach	SUL-WAT1-120907	ug/L	0.5	0.5 U	0								
10) South End Offion Lake Beach	SUL-WAT2-120907	ug/L	0.5	0.5 U	0								
Duplicates	NUL-WAT1 (DUP)*	ug/L	4.5	4.5 UJ	0								
Duplicates	ULB-WAT2 (DUP)*	ug/L	5.0	5 UJ	0								
	PBLANK-01-100907	ug/L	0.5	0.5 U	0								
	PBLANK-02-110907	ug/L	0.5	0.5 U	0								
	PBLANK-03-120907	ug/L	0.5	0.5 U	0								
Equipment Blank	BSBLANK-01-100907	ug/L	0.5	0.5 U	0								
	BSBLANK-02-110907	ug/L	0.5	0.5 U	0								
	BRBLANK-120907	ug/L	0.5	0.5 U	0								
	TTBLANK-120907	ug/L	0.5	0.5 U	0								

**RL** = laboratory reporting limit

U = PCB aroclors were analyzed, but not detected.

**UJ** = PCB aroclors were not detected above the reported sample quantitation limit. However, the reported quantitation limit is approximate and may or may not represent the actual limit of quantitation necessary to accurately and precisely measure the analyte in the sample.

<sup>\*</sup>Denotes cross-referenced sample location of blind duplicate sample

<sup>\*\*</sup>Total PCB aroclors calculated by summing concentrations of the nine aroclors using ND = 0

TABLE 4-2. PCB AROCLOR CONCENTRATIONS (ug/Kg) AND TOTAL PCBs\*\* IN SOIL AND SEDIMENT SAMPLES COLLECTED IN VICINITY OF VINELAND CHEMICAL SUPERFUND SITE, SEPTEMBER 2007

					RESULTS									
Sample Location	Station ID	Units	Matrix	RL	Aroclor-	Total PCBs								
•					1016	1221	1232	1242	1248	1254	1260	1262	1268	(ND = 0)
	MILL-SED	ug/Kg	Sediment	21	21 U	0								
	MILL-SHORE	ug/Kg	Sediment	32	32 U	0								
1) West of Mill Rd	MILL-BORING-1	ug/Kg	Sediment	28	28 U	0								
	MILL-BORING-2	ug/Kg	Sediment	47	47 U	0								
	MILL-BORING-3	ug/Kg	Sediment	20	20 U	1262         1268           21 U         21 U           32 U         32 U           28 U         28 U           47 U         47 U	0							
	R55-SED	ug/Kg	Sediment	170	170 R	NA								
	R55-SHORE	ug/Kg	Sediment	90	90 UJ	0								
2) West of Rte 55	R55-BORING-1	ug/Kg	Sediment	130	130 UJ	0								
	R55-BORING-2	ug/Kg	Sediment	41	45 U	9.4 JN	45 U	45 U	45 U	9.4				
	R55-BORING-3	ug/Kg	Sediment	diment         90         90 UJ         130 UJ         141 UJ         15 U         15 U         15 U         15 U         15 U         15 U	19 U	4.1								
0) DWD 0 M : G G	BWB-SED	ug/Kg	Sediment	20	20 U	0								
3) BWB & Maurice Confluence	BWB-SHORE	ug/Kg	Sediment	21	21 U	0								
	ALLIANCE-BEACH	ug/Kg	Soil	20	20 U	0								
A) AH: D I	ALLIANCE-SED	ug/Kg	Sediment	19	19 U	0								
4) Alliance Beach	ALLIANCE-SHORE	ug/Kg	Sediment	21	21 U	0								
	ALLIANCE-BORING-1	ug/Kg	Sediment	18	18 U	0								
	ALMOND-BEACH	ug/Kg	Soil	17	17 U	0								
	ALMOND-SED	ug/Kg	Sediment	19	19 U	0								
5) Almond Beach	ALMOND-SHORE	ug/Kg	Sediment	21	21 U	0								
•	ALMOND-BORING-1	ug/Kg	Sediment	18	18 U	0								
	ALMOND-BORING-2	ug/Kg	Sediment	17	17 U	21 U 18 U 17 U 17 U 18 U	0							
	BA-BEACH	ug/Kg	Soil	17	17 U	0								
6) "BareA" Beach	BA-SED	ug/Kg	Sediment	18	18 U	0								
	BA-SHORE	ug/Kg	Sediment	120	120 UJ	0								
7) 61	SHERMAN-SED	ug/Kg	Sediment	120	120 UJ	0								
7) Sherman Ave.	SHERMAN-SHORE	ug/Kg	Sediment	20	20 U	0								
8) North End of Union Lake	NUL-SED-120907	ug/Kg	Sediment	38	38 U	0								
8) North End of Union Lake	NUL-SHORE-120907	ug/Kg	Sediment	130	130 UJ	0								
	ULB-BEACH-120907	ug/Kg	Soil	17	17 U	0								
9) Union Lake Beach	ULB-BORING-1	ug/Kg	Sediment	19	19 U	0								
9) Ullon Lake Beach	ULB-SED-120907	ug/Kg	Sediment	76	76 UJ	0								
	ULB-SHORE-120907	ug/Kg	Sediment	21	21 U	0								
	SUL-BEACH-120907	ug/Kg	Soil	17	17 U	0								
10) South End Union Lake Beach	SUL-BORING-1	ug/Kg	Sediment	19	19 U	0								
10) South End Union Lake Beach	SUL-SED-120907	ug/Kg	Sediment	22	22 U	0								
	SUL-SHORE-120907	ug/Kg	Sediment	20	20 U	0								
	BWB-SED (DUP)*	ug/Kg	Sediment	39	39 UJ	0								
	BWB-SHORE (DUP)*	ug/Kg	Sediment	41	41 UJ		41 UJ	0						
	ALLIANCE-BEACH (DUP)*	ug/Kg	Soil	39	39 UJ		0							
Duplicates	ALMOND-SHORE (DUP)*	ug/Kg	Sediment	39	39 UJ	0								
	BA-SED (DUP)*	ug/Kg	Sediment	39	39 UJ	0								
	SHERMAN-SED (DUP)*	ug/Kg	Sediment	130	130 UJ	0								
	SUL-BEACH (DUP)*	ug/Kg	Soil	34	34 UJ	0								

RL = laboratory reporting limit

NOTE: Bold values represent detected PCB aroclor concentrations

U = PCB aroclors were analyzed, but not detected.

UJ = PCB aroclors were not detected above the reported sample quantitation limit. However, the reported quantitation limit is approximate and may or may not represent the actual limit of quantitation necessary to accurately and precisely measure the analyte in the sample.

J = The analyte was positively identified; the associated numerical value is the approximate concentration of the analyte in the sample.

JN = the analysis indicates that the presence of an analyte that has been "tentatively identified" and the associated numerical value represents its approximate concentration.

R = the sample results are rejected due to serious deficiencies in the ability to analyzed the sample and meet quality control criteria; the presence or absence of the analyte cannot be verified.

NA = results are not applicable due to rejection of data during data validation process

<sup>\*</sup>Denotes cross-referenced sample location of blind duplicate sample

<sup>\*\*</sup>Total PCB aroclors calculated by summing concentrations of the nine aroclors using ND = 0

### 5. SUMMARY AND COMPARISONS TO PREVIOUS DATA

Overall, results of the September 2007 Post Remedial Action Sampling indicated more exceedences of the arsenic criterion for agitated water samples than in previous sampling events. Arsenic concentrations in agitated water samples exceeded the 10 ppb criterion at seven of the ten sampling locations. Both the May 2006 Baseline and the November 2006 (during excavation) Periodic Sampling events exhibited only three locations with agitated water exceedences. The arsenic concentration in agitated water (3,800 ppb) collected in September 2007 from Station 1 (West of Mill Rd) was substantially higher than any other water sample tested during the three sampling events.

As indicated in the May 2006 and November 2006 Sampling events, the two stations located immediately downstream of the Vineland site, Station 1 (West of Mill Rd.) and Station 2 (West of Rte. 55) had the highest concentrations of arsenic in sediment and had the greatest number of concentrations that exceeded the Site Clean-up Level of 20 ppm for arsenic (Figure 3-1). Unlike the May 2006 and November 2006 Sampling events, the arsenic at stations located further downstream, including Station 7 (Sherman Ave), Station 8 (North End of Union Lake), and Station 9 (Union Lake), more frequently exceeded the water and soil criteria than in previous sampling events. Although located furthest downstream of the Vineland site, the stations located along Union Lake, including Station 8 (North End of Union Lake) and Station 9 (Union Lake Beach) had arsenic concentrations in sediments that exceeded the Site Clean-up Level of 20 ppm. These trends in Union Lake may be attributable to the proportion of fine silt/clays that were observed in the sediment samples; arsenic is strongly sorbed onto fine particulates, including silt (Bodek et. al 1988). The arsenic that originates from upstream sources may be transported downstream via particulates which settle out in the lake depositional areas. Previous reports for the site have stated that sediment in the Maurice River and Union Lake contains a high content of organic matter (USEPA 1999). Importantly, with the exception of Station 7 (Sherman Ave), the arsenic concentrations that exceeded the Site Clean-up Level of 20 ppm were for lake sediments collected greater than 200 ft from the shoreline. In addition, arsenic concentrations from the five beach locations [Station 4 (Alliance Beach), Station 5 (Almond Beach), Station 6 ("BareA" Beach), Station 9 (Union Lake Beach), and Station 10 (South End Union Lake Beach)] were either < 1.0 mg/Kg or below the analytical detection limit.

Below Station 2 (West of Rte. 55), additional water flow from the Maurice River and other tributaries flowing into the Maurice River may transport arsenic that is bound to fine particulates further downstream. Previously in the May 2006 and November 2006 Sampling events, arsenic concentrations in sediments, water, and beach soil did not exceed criteria at Station 4 (Alliance Beach), Station 5 (Almond Beach), or Station 6 ("BareA" Beach). However, in September 2007, the first exceedence of applicable criteria occurred at Station 5 (Almond Beach) in the agitated water sample and at Station 6 ("BareA" Beach) in both a surface water sample and a nearshore (shore) sediment sample.

Ten total sediment boring samples were collected from six of the ten sampling stations. Four of the six stations that were sampled for sediment borings were collected only for the first depth interval (0-1 ft / closest to surface) and resulted in either no detection or low detection of arsenic; grain size analyses in the first depth interval for these stations indicated that sand was the

dominant substrate. Two of the six stations, Station 1 (West of Mill Rd) and Station 2 (West of Rte. 55), were sampled in three depth intervals for the sediment borings. The general trends observed for these stations resulted in arsenic greater than the 20 ppm criterion for the first depth interval (0-1 ft deep), higher arsenic concentrations for the second depth interval (1-2 ft deep), and concentrations less than the 20 ppm criterion in the deepest third depth interval (2-3 or 2-4 ft deep). Grain size data for Station 2 (West of Rte. 55) indicated that fine-grained material (silts/clays) dominated the first (0-1 ft) depth interval, while the grain size data for Station 1 (West of Mill Rd) indicated that sand dominated the first (0-1 ft) depth interval.

Overall, for all stations and matrices, during the May 2006 Baseline Sampling event, the frequency of arsenic detection was 57 percent, with 32 percent of analyzed samples exceeding the applicable criteria for arsenic. Comparatively, during the November 2006 Periodic Sampling event, the frequency of arsenic detection was 53 percent, with 27 percent of analyzed samples exceeding the applicable criteria for arsenic. During the September 2007 Post Remedial Action Sampling event, the frequency of arsenic detection was 68 percent, with 42 percent of analyzed samples exceeding the applicable criteria for arsenic. The increased frequency of detection and frequency of exceeding applicable criteria that were observed in the September 2007 Post Remedial Action Sampling could potentially be attributed to the excavation activities associated with the remedial actions currently occurring at the site.

None of the 20 surface water and agitated water samples analyzed had detected concentrations of PCB aroclors. Two of the 36 sediment samples, boring-2 and boring-3 at Station 2 (West of Rte 55), had detected concentrations for Aroclor-1254 of 9.4 and 4.1 ppb ( $\mu g/Kg$ ), respectively. However, neither of these two sediment/soil samples exceeded the 200 ppb ( $\mu g/Kg$ ) Soil Remediation Standard for total PCBs.

### 5.1 Summary of September 2007 Post Remedial Action Sampling Arsenic Results by Station

### Station 1 – West of Mill Rd (see Figures 3-1 and 3-2)

Arsenic concentrations in the agitated water sample exceeded the 10 ppb USEPA Drinking Water Criterion by a factor of 380; at 3,800  $\mu$ g/L, the arsenic concentration at Mill-Wat2 was the single highest concentration measured during the September 2007 Post Remedial Action Sampling. Arsenic concentrations in the nearshore (shore) surficial sediment exceeded the Site Clean-up Level of 20 ppm by a factor of 6.5 and in the sediment borings by a factor of 4.7 (Boring-1) and by a factor of 23 (Boring-2). Arsenic concentrations in neither the mid-stream surficial sediment (0-0.5 ft depth increment) nor the third-depth sediment boring (Boring-3) exceeded the Site Clean-up Level of 20 ppm

### Station 2 – West of Rte 55 (see Figures 3-1 and 3-2)

Both of the arsenic concentrations in the surface water and agitated water sample exceeded the 10 ppb USEPA Drinking Water Criterion by a factor of 1.4 and 70, respectively. Arsenic concentrations also exceeded the Site Clean-up Level of 20 ppm in the mid-stream surficial sediment (0-0.5 ft depth increment) by a factor of 85, in the nearshore (shore) surficial sediment by a factor of 170, and in the sediment borings by a factor of 95 (Boring-1) and by a factor of 180 (Boring-2). The third boring (Boring-3) was the only sample that did not exceed the Site

Clean-up Level of 20 ppm.

### Station 3 – BWB & Maurice Confluence (see Figures 3-1 and 3-3)

None of the arsenic concentrations detected in sediment samples exceeded the Site Clean-up Level of 20 ppm. Arsenic was not detected in either of the water samples.

### Station 4 – Alliance Beach (see Figures 3-1 and 3-3)

Arsenic was not detected in any of the sediment, water, or beach soil samples at Station 4.

### Station 5 – Almond Beach (see Figures 3-1 and 3-4)

None of the arsenic concentrations detected in sediment samples exceeded the Site Clean-up Level of 20 ppm. Arsenic was not detected in beach soils, in the nearshore (shore) surficial sediment, in the surface water sample, or the one sediment boring (0-1 ft) that was collected; arsenic was detected at 1.3 mg/Kg in the mid-stream surficial sediment (0-0.5 ft depth increment). The arsenic concentration in the agitated water sample exceeded the 10 ppb USEPA Drinking Water Criterion by a factor of 1.6.

### Station 6 – "BareA" Beach (see Figures 3-1 and 3-5)

The arsenic concentration in the nearshore (shore) surficial sediment was 290 mg/Kg, which exceeded the Site Clean-up Level of 20 ppm by a factor of 14.5. Arsenic was detected at 2.3 mg/Kg in the mid-stream surficial sediment (0-0.5 ft depth increment). Arsenic was not detected in beach soils or in the agitated (Wat-2) water sample. However, the arsenic concentration in the surface water sample was 13  $\mu$ g/L, which exceeded the 10 ppb USEPA Drinking Water Criterion by a factor of 1.3.

### Station 7 – Sherman Ave. (see Figures 3-1 and 3-6)

Arsenic concentrations in the mid-stream surficial sediment (0-0.5 ft depth increment) sample and both water samples exceeded applicable criteria. The arsenic concentration in the mid-stream surficial sediment (0-0.5 ft depth increment) was 110 mg/Kg, which exceeded the Site Clean-up Level of 20 ppm by a factor of 5.5. The arsenic concentrations in the surface water (12  $\mu$ g/L) and the agitated water (100  $\mu$ g/L) sample exceeded the 10 ppb USEPA Drinking Water Criterion by factors of 1.2 and 10, respectively. Arsenic was detected at 5.5 mg/Kg in the nearshore (shore) surficial sediment.

### Station 8 – North End of Union Lake (see Figures 3-1 and 3-7)

Both of the arsenic concentrations in the surficial and nearshore (shore) sediment samples exceeded the Site Clean-up Level of 20 ppm. The arsenic concentration in the surficial sediment (0-0.5 ft depth increment) collected 200 ft offshore was 30 mg/Kg and in the nearshore (shore) surficial sediment was 320 mg/Kg, which exceeded the Site Clean-up Level of 20 ppm by factors of 1.5 and 16, respectively. Arsenic was not detected in the surface water, but was detected in the agitated water sample (22  $\mu$ g/L) and exceeded the 10 ppb USEPA Drinking Water Criterion by a factor of 2.2.

### Station 9 – Union Lake Beach (see Figures 3-1 and 3-8)

Arsenic was detected in each of the soil/sediment samples, including the one sediment boring (0-1 ft) collected at Station 9. Arsenic was detected at 360 mg/Kg in the surficial sediment (0-0.5 ft depth increment) collected 200 ft offshore and exceeded the Site Clean-up Level of 20 ppm by a factor of 18. Arsenic was detected at 2.1 mg/Kg in the nearshore (shore) surficial sediment, at 1.0 mg/Kg in the beach (soil) sample, and at 1.4 mg/Kg in the sediment boring (Boring-1). Arsenic was not detected in the non-agitated surface water, but was detected in the agitated water sample (15 μg/L) and exceeded the 10 ppb USEPA Drinking Water Criterion by a factor of 1.5.

### Station 10 – South End of Union Lake Beach (see Figures 3-1 and 3-8)

None of the detected arsenic concentrations in sediment samples exceeded the Site Clean-up Level of 20 ppm. Arsenic was detected in the surficial sediment (0-0.5 ft depth increment) collected 200 ft offshore at 18 mg/Kg, in the nearshore (shore) surficial sediment at 6.5 mg/Kg, and in the sediment boring (Boring-1) at 12 mg/Kg. Arsenic was not detected in the beach (soil) sample. Arsenic was not detected in the surface water, but was detected in the agitated water sample (28 µg/L) and exceeded the 10 ppb USEPA Drinking Water Criterion by a factor of 2.8.

### 5.2 Comparison of September 2007 Arsenic Results to May 2006 and November 2006 Arsenic Results

The current phase of remediation at the site involves removing the contaminated soils/sediments of the Blackwater Branch and the floodplain east of Mill Road and adjacent to the site. As stated previously, the baseline sampling was conducted in May 2006 and the November 2006 Periodic Sampling was conducted during the excavation/construction activities. The results from the previous sampling events are compared to the results from the September 2007 Post Remedial Action Sampling.

In the May 2006 Baseline Sampling, the November 2006 Periodic Sampling, and the September 2007 Post Remedial Action Sampling, arsenic concentrations were analyzed in sediments, soil, and water collected at the same ten locations in and near waterways located adjacent to the site. Both the May 2006 Baseline Sampling and the September 2007 Post Remedial Action Sampling included the collection and analysis of deep sediment cores; the November 2006 Periodic Sampling effort did not include the collection and analysis of deep sediment cores. With the exception of core sampling and analysis, the sampling scheme and sampling locations for the May 2006 Baseline Sampling, the November 2006 Periodic Sampling, and the September 2007 Post Remedial Action Sampling were identical. The following paragraphs and Table 5-1 compare the results of the May 2006 Baseline Sampling, the November 2006 Periodic Sampling, and the September 2007 Post Remedial Action Sampling for sediment, soil, and water samples.

### May and November 2006 and September 2007 Arsenic Surface Water Data

The May 2006 results indicated that five of 20 surface water samples exceeded the applicable criterion, the November 2006 results indicated that only three of 20 surface water samples exceeded the applicable criterion, and the September 2007 results indicated that ten of 20 surface water samples exceeded the applicable criterion. Surface water arsenic concentrations that were equivalent to or exceeded the USEPA Drinking Water Criterion of  $10 \mu g/L$  (ppb) ranged from 10 to 1,900  $\mu g/L$  for the May 2006 sampling, from 11 to 900  $\mu g/L$  for the November 2006

sampling, and from 12 to 3,800  $\mu$ g/L for the September 2007 sampling. Data for each of the three sampling events indicated that the stations with the highest arsenic concentrations were located directly downstream of the site – Station 1 (West of Mill Rd.) and Station 2 (West of Rte. 55). Generally, for the November 2006 Periodic Sampling, only the agitated water samples (versus surface water samples) exceeded the USEPA Drinking Water Criterion of 10  $\mu$ g/L (ppb). In September 2007, there were substantially more exceedences of arsenic in the agitated water samples at the majority of the stations. First-time exceedences of the USEPA Drinking Water Criterion for arsenic in surface water occurred at Station 5 (Almond Beach), Station 8 (North End of Union Lake), and Station 9 (Union Lake Beach).

### May and November 2006 and September 2007 Arsenic Mid-Stream Sediment (0-0.5ft) Data

The May 2006 results indicated that four of 10 sediment samples exceeded the applicable criterion, the November 2006 results indicated that five of 10 sediment samples exceeded the applicable criterion, and the September 2007 results indicated that four of 10 sediment samples exceeded the applicable criterion. Arsenic concentrations that exceeded the Site Clean-up Level of 20 ppm ranged from 160 to 1,500 mg/Kg for the May 2006 results, ranged from 20 to 2,100 mg/Kg for the November 2006 results, and ranged from 30 to 1,700 mg/Kg for the September 2007 results. For each of the three sampling events, the stations with the highest arsenic concentrations were located directly downstream of the site – Station 1 (West of Mill Rd.) or Station 2 (West of Rte. 55) – and at the stations located north and along Union Lake – Station 7 (Sherman Ave), Station 8 (North End of Union Lake), and Station 9 (Union Lake Beach). Consistently, Station 2 (West of Rte. 55) had the absolute highest arsenic concentrations in midstream sediment (0-0.5 ft depth increment beneath the water/sediment interface) for each of the three sampling events; the May 2006 arsenic concentration was 1,500 mg/Kg, the November 2006 arsenic concentration was 2,100 mg/Kg, and the September 2007 arsenic concentration was 1,700 mg/Kg.

### May and November 2006 and September 2007 Arsenic Nearshore (Shore) Sediment (0-0.5ft) Data

Shore stations from Station 1 (West of Mill Rd.), Station 2 (West of Rte. 55), and Station 8 (North End of Union Lake) exceeded the Site Clean-up Level of 20 ppm for each of the three sampling events. In September 2007, the shore sample for Station 6 ("BareA" Beach) also exceeded the criterion. Arsenic concentrations in the shore sediments that exceeded the Site Clean-up Level of 20 mg/Kg (ppm) ranged from 88 to 1,200 mg/Kg, 140 to 4,500 mg/Kg, and 110 to 3,400 mg/Kg for the May 2006, November 2006, and September 2007 events, respectively. Data from each of the three sampling events indicate that the stations with the highest arsenic concentrations were located directly downstream of the site – Station 1 (West of Mill Rd.) and Station 2 (West of Rte. 55) and at Station 8 (North End of Union Lake). In September 2007, for the first time, an arsenic exceedence of 290 mg/Kg occurred at Station 6 ("BareA" Beach). Similar to the mid-stream sediment arsenic results, Station 2 (West of Rte. 55) had the absolute highest arsenic concentrations in shore samples for each of the three sampling events with concentrations of 1,200 mg/Kg, 4,500 mg/Kg, and 3,400 mg/Kg in May 2006, November 2006, September 2007, respectively. The arsenic concentration at Station 2 (West of Rte. 55) was the highest recorded arsenic concentration (4,500 mg/Kg) measured for all sampling events (years 1992 through 2006).

May and November 2006 and September 2007 Arsenic Beach Soil (0-0.5ft) Data

Detected concentrations in May 2006, November 2006, and September 2007 were either comparable to or lower than those previously reported for all five stations and none of the 2006 or 2007 beach samples exceeded the Site Clean-up Level of 20 ppm for arsenic in soil.

### 5.3 Comparison of Arsenic Results to Historical Data

During 1992 and from 1994 through 1999, water, soil, and sediment samples were collected in the vicinity of and downstream of the Vineland site at beach stations for arsenic analyses (USEPA/ERTC 1999). These data were collected to evaluate the results against human health risk-base action levels and were part of an annual monitoring program performed at beaches along the Maurice River and Union Lake (USEPA/ERTC 1999). Data were collected from five beach locations including Alliance Beach (Station 4), Almond Beach (Station 5), "BareA" Beach (Station 6), Union Lake Beach (Station 9), and South End Union Lake Beach (Station 10). Each of the matrices (water, soil, and sediment) was not collected at each station every year. From 1992 and 1994 through 1999, no discernable trends in the historical arsenic data were evident; the concentration of arsenic in each matrix appeared to remain relatively constant over time (USEPA/ERTC 1999). The 1999 report that contains historical arsenic data for the 1992 and 1994 through 1999 is provided in Appendix E. These data were compared to data from the 2006 and 2007 sampling events.

The historical arsenic data for the five stations (listed above) were compared to the May 2006 Baseline Sampling, the November 2006 Periodic Sampling, and the September 2007 Post Remedial Action Sampling for surface water, beach soils, and surficial sediment arsenic concentrations (Tables 5-2 through 5-4 and Figures 5-1 through 5-3, respectively). The following paragraphs compare the historical arsenic data (1992 and 1994 through 1999) to the data collected in May 2006, November 2006, and September 2007 by matrix (water, soil, and sediment) and by station. Five of the ten total stations that were sampled in the May and November 2006 and the September 2007 sampling surveys were also sampled in 1992 and 1994 through 1999; the five stations included in the 2006 and 2007 surveys that were not previously sampled (Stations 1, 2, 3, 7, and 8) are not included in this discussion.

## Comparisons to Historical Arsenic Surface Water Data (1992, 1994 through 1999, 2006, and 2007)

Throughout the period of 1992 and 1994-1999, arsenic concentrations in surface waters at Station 4 (Alliance Beach), Station 5 (Almond Beach), and Station 6 ("BareA" Beach) were variable and substantially exceeded the current USEPA Drinking Water Criterion of 10 ppb. Arsenic concentrations in surface waters at Station 9 (Union Lake Beach) slightly declined from 1996 (above criterion) through 1999 (below criterion). The arsenic concentration in surface water at Station 10 (South End of Union Lake Beach) was above the criterion in both 1998 and 1999. Surface water data from samples collected in both May 2006 and November 2006 indicated that arsenic was not detected in the surface waters at each of these five previously sampled locations (Stations 4, 5, 6, 9, and 10), with the exception of Station 10 (South End of Union Lake Beach) in May 2006, when arsenic in the agitated water sample was equivalent to the USEPA Drinking Criterion. However, in September 2007, there were several exceedences of

the arsenic criterion in surface water. The majority of these exceedences corresponded to agitated water samples.

Comparisons to Historical Beach Soil Data (1992, 1994 through 1999, 2006, and 2007)

None of the beach soil samples collected in 1992, 1994 through 1999, and 2006 exceeded the Site Clean-up Level of 20 ppm for arsenic at Station 4 (Alliance Beach), Station 5 (Almond Beach), Station 6 ("BareA" Beach), Station 9 (Union Lake Beach), and Station 10 (South End of Union Lake Beach). Detected concentrations in May 2006, November 2006, and September 2007 were either comparable to or lower than those previously reported for each of the five stations and none of the 2006 or 2007 samples exceeded the Site Clean-up Level of 20 ppm.

Comparisons to Historical Surface Sediment Data (1992, 1994 through 1999, 2006, and 2007)

Throughout the period of 1992 and 1994-1999, arsenic concentrations were below the Site Clean-up Level of 20 ppm at each of the five stations, with the exception of Station 6 ("BareA" Beach) in 1998. Results from samples collected in May 2006 indicated that arsenic concentrations in surficial sediment (collected greater than 200 ft from the shoreline) at Station 9 (Union Lake Beach) and Station 10 (South End of Union Lake Beach) were substantially higher than concentrations previously reported in 1992 and 1994-1999. Results from samples collected in both November 2006 and September 2007 indicated that arsenic concentrations in surficial sediment (collected greater than 200 ft from the shoreline) at Station 9 (Union Lake Beach) were substantially higher than concentrations previously reported in 1992 and 1994-1999, but comparable to concentrations reported in May 2006 at this station. These changes could potentially be attributable to downstream transport of arsenic bound to fine grained materials (i.e., silts) and their subsequent accumulation in depositional areas of the lake.

### 5.4 PCB Results

None of the 20 surface water samples analyzed had detected concentrations of PCB aroclors. Two of the 36 sediment samples, boring-2 and boring-3 at Station 2 (West of Rte 55), had detected concentrations for Aroclor-1254 of 9.4 and 4.1 ppb ( $\mu g/Kg$ ), respectively. However, neither of these sediment concentrations exceeded the 200 ppb ( $\mu g/Kg$ ) Soil Remediation Standard for total PCBs.

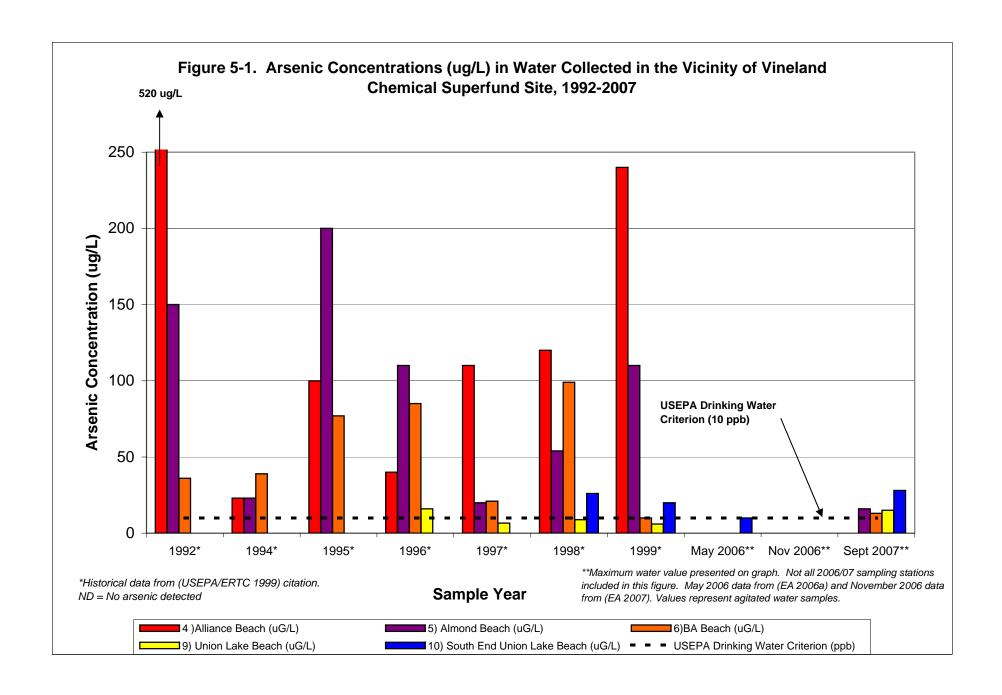


Figure 5-2. Arsenic Concentrations (mg/Kg) in Beach (Soil) Samples Collected in the Vicinity of Vineland Chemical Superfund Site, 1992-2007 25 Site Clean-up Level (20 ppm) 20 Arsenic Concentration (mg/kg) 5 ND 0 1992\* 1994\* 1995\* 1996\* 1997\* 1998\* 1999\* May 2006\*\* Nov 2006\*\* Sept 2007 \*Historical data from (USEPA/ERTC 1999) citation. Sample Year \*\*May 2006 data from (EA 2006a) and November 2006 data from ND = No arsenic detected (EA 2007). 4 )Alliance Beach (mg/Kg) ■5) Almond Beach (mg/Kg) ■6)BA Beach (mg/Kg) ■9) Union Lake Beach (mg/Kg) ■10) South End Union Lake Beach (mg/Kg) Site Clean-up Level (mg/Kg)

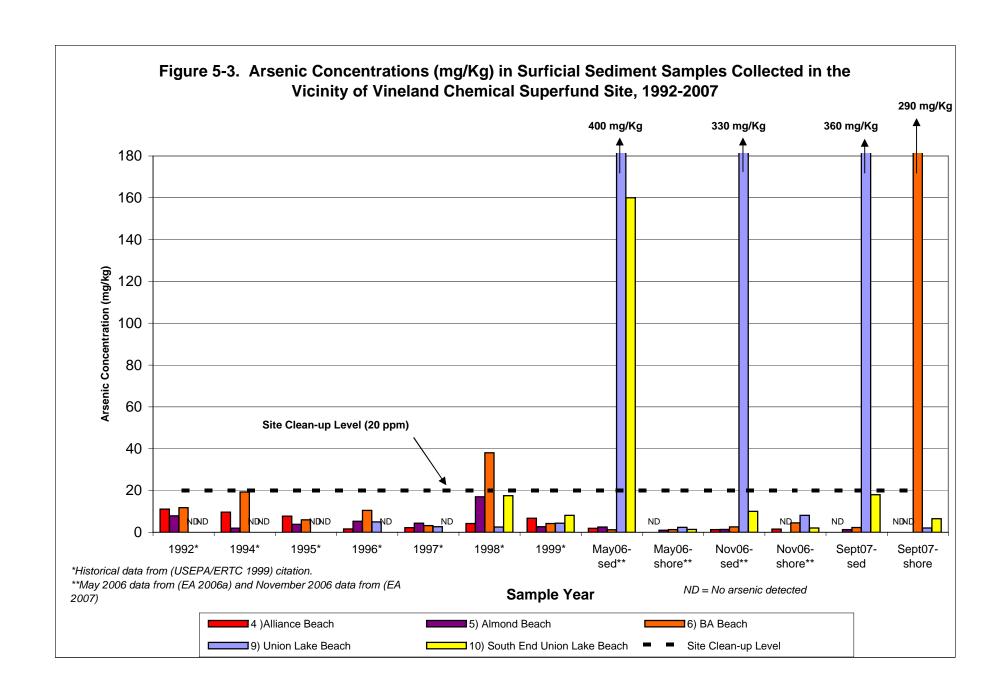


TABLE 5-1. COMPARISON OF ARSENIC RESULTS FOR SAMPLES COLLECTED IN VICINITY OF VINELAND CHEMICAL SUPERFUND SITE, MAY 2006 BASELINE SAMPLING, NOVEMBER 2006 PERIODIC SAMPLING, AND SEPTEMBER 2007 POST REMEDIAL ACTION SAMPLING VINELAND POST REMEDIAL ACTION SAMPLING AND MONITORING PROGRAM, OPERABLE UNITS #3 AND #4

Sampling Station	Units	May 2006 Baseline Sampling	November 2006 Periodic Sampling	September 2007 Post Remedial Action Sampling
Water Sample Results				
1) MILL-WAT1	ug/L	200	U	U
1) MILL-WAT2	ug/L	U	900	3,800
2) R55-WAT1	ug/L	14	8.8	14
2) R55-WAT2	ug/L	1,900	660	700
3) BWB-WAT1	ug/L	U	U	U
3) BWB-WAT2	ug/L	U	U	U
4) ALLIANCE-WAT1	ug/L	U	U	U
4) ALLIANCE-WAT2	ug/L	U	U	U
5) ALMOND-WAT1	ug/L	U	U	U
5) ALMOND-WAT2	ug/L	U	U	16
6) BA-WAT1	ug/L	U	U	13
6) BA-WAT2	ug/L	U	U	U
7) SHERMAN-WAT1	ug/L	U	U	12
7) SHERMAN-WAT2	ug/L	55	11	100
8) NUL-WAT1	ug/L	U	U	U
8) NUL-WAT2	ug/L	U	U	22
9) ULB-WAT1	ug/L	U	U	U
9) ULB-WAT2	ug/L	U	U	15
10) SUL-WAT1	ug/L	U	U	U
10) SUL-WAT2	ug/L	10	U	28
Solid Sediment Results				
1) MILL-SED	mg/Kg	14	160	8.2
2) R55-SED	mg/Kg	1,500	2,100	1,700
3) BWB-SED	mg/Kg	0.78	20	1.5
4) ALLIANCE-SED	mg/Kg	1.9	1.3	U
5) ALMOND-SED	mg/Kg	2.5	1.4	1.3
6) BA-SED	mg/Kg	1.2	2.6	2.3
7) SHERMAN-SED	mg/Kg	12	7.3	110
8) NUL-SED	mg/Kg	230	510	30
9) ULB-SED	mg/Kg	400	330	360
10) SUL-SED	mg/Kg	160	10	18
Solid Shore Results		•		
1) MILL-SHORE	mg/Kg	270	140	130
2) R55-SHORE	mg/Kg	1,200	4,500	3,400
3) BWB-SHORE	mg/Kg	U	19	2
4) ALLIANCE-SHORE	mg/Kg	U	1.5	U
5) ALMOND-SHORE	mg/Kg	1	U	U
6) BA-SHORE	mg/Kg	1.3	4.5	290
7) SHERMAN-SHORE	mg/Kg	6.3	U	5.5
8) NUL-SHORE	mg/Kg	88	340	320
9) ULB-SHORE	mg/Kg	2.4	8.1	2.1
10) SUL-SHORE	mg/Kg	1.4	2.1	6.5
Solid Beach Results		<u> </u>		
4) ALLIANCE-BEACH	mg/Kg	1	U	U
5) ALMOND-BEACH	mg/Kg	U	U	U
6) BA-BEACH	mg/Kg	Ü	U	U
9) ULB-BEACH	mg/Kg	0.9	1.1	1
10) SUL-BEACH	mg/Kg	U	U	U

TABLE 5-1. CONTINUED VINELAND POST REMEDIAL ACTION SAMPLING AND MONITORING PROGRAM, OPERABLE UNITS #3 AND #4

Sampling Station	Sampling Station Units		November 2006 Periodic Sampling	September 2007 Post Remedial Action Sampling
Sediment Core (Boring) Results				
1) MILL-BORING-1 (0-1 ft)	mg/Kg	120	NS	94
1) MILL-BORING-2 (1-2 ft)	mg/Kg	27	NS	460
1) MILL-BORING-3 (2-4 ft)	mg/Kg	4.7	NS	6.1
1) MILL-BORING-4 (4-5 ft)	mg/Kg	10	NS	NS
2) R55-BORING-1 (0-1 ft)	mg/Kg	160	NS	1,900
2) R55-BORING-2 (1-2 ft)	mg/Kg	82	NS	3,600
2) R55-BORING-3 (2-3 ft)	mg/Kg	30	NS	9.7
4) ALLIANCE-BORING-1 (0-1 ft)	mg/Kg	U	NS	U
4) ALLIANCE-BORING-2 (1-2 ft)	mg/Kg	U	NS	NS
4) ALLIANCE-BORING-3 (2-3.3 ft)	mg/Kg	U	NS	NS
5) ALMOND-BORING-1 (0-1 ft)	mg/Kg	U	NS	2
5) ALMOND-BORING-2 (1-2 ft)	mg/Kg	U	NS	NS
5) ALMOND-BORING-3 (2-2.4 ft)	mg/Kg	U	NS	NS
9) ULB-BORING-1 (0-1 ft)	mg/Kg	79	NS	1.4
9) ULB-BORING-2 (1-2 ft)	mg/Kg	31	NS	NS
10) SUL-BORING-1 (0-1 ft)	mg/Kg	390	NS	12
10) SUL-BORING-2 (1-2 ft)	mg/Kg	110	NS	NS
10) SUL-BORING-3 (2-3.4 ft)	mg/Kg	1.7	NS	NS

### NOTES:

U = undetected arsenic concentration

Bold values represent detected arsenic concentrations; shaded and bolded values indicate arsenic exceedences as noted below: Site Clean-up Level for Solids = 20 ppm (mg/kg)

USEPA Drinking Water Criterion for Water = 10 ppb (ug/L)

NS = no sample collected

TABLE 5-2. ARSENIC CONCENTRATIONS (uG/L) IN WATER SAMPLES COLLECTED IN VICINITY OF VINELAND CHEMICAL SUPERFUND SITE, 1992-2007

Sample Location		Sampling Year									
Sample Location	1992*	1994*	1995*	1996*	1997*	1998*	1999*	May 2006*+	Nov 2006+	Sept 2007+	
4 )Alliance Beach (uG/L)	520	23	100	40	110	120	240	ND/ND	ND/ND	ND/ND	
5) Almond Beach (uG/L)	150	23	200	110	20	54	110	ND/ND	ND/ND	ND/ <b>16</b>	
6)BA Beach (uG/L)	36	39	77	85	21	99	10	ND/ND	ND/ND	13/ND	
9) Union Lake Beach (uG/L)	NS	NS	NS	16	6.6	8.8	6	ND/ND	ND/ND	ND/15	
10) South End Union Lake Beach (uG/L)	NS	NS	NS	NS	NS	26	20	ND/10	ND/ND	ND/28	
USEPA Drinking Water Criterion (ppb)	10	10	10	10	10	10	10	10	10	10	

NOTE: Shaded and bold values represent detected arsenic concentrations equivalent to or above the USEPA Drinking Water Criterion of 10 ug/L (ppb) for arsenic

NS = no sample collected; ND = not detected; below analytical limit

<sup>\*</sup>Historical data from (USEPA/ERTC 1999) citation

<sup>\*\*</sup>May 2006 data from (EA 2006a) citation

<sup>+2006</sup> and 2007 data are presented as WAT1/WAT2

TABLE 5-3. ARSENIC CONCENTRATIONS (mg/Kg) IN BEACH (SOIL) SAMPLES COLLECTED IN VICINITY OF VINELAND CHEMICAL SUPERFUND SITE, 1992-2007

	Sampling Year									
Sample Location	1992*	1994*	1995*	1996*	1997*	1998*	1999*	May 2006**	Nov 2006**	Sept 2007
4 )Alliance Beach (mg/Kg)	NS	0.45 ND	2.2	0.89	0.49	1.1	0.59	1	ND	ND
5) Almond Beach (mg/Kg)	NS	0.92	0.86	0.76	0.46	1	0.43	ND	ND	ND
6)BA Beach (mg/Kg)	NS	0.44 ND	4	0.67	0.81	0.41 ND	0.47 ND	ND	ND	ND
9) Union Lake Beach (mg/Kg)	NS	NS	NS	1.3	3.1	1.2	2.6	0.9	1.1	1
10) South End Union Lake Beach (mg/Kg)	NS	NS	NS	NS	NS	2.3	0.48 ND	ND	ND	ND
Site Clean-up Level (mg/Kg)	20	20	20	20	20	20	20	20	20	20

NOTE: Italics = arsenic undetected at indicated concentration (detection limit)

NS = No sample collected; ND = not detected; below analytical detection limit

No beach (soil) samples exceeded the Site Clean-up Level of 20 mg/Kg for solids

\*Historical data from (USEPA/ERTC 1999) citation

### TABLE 5-4. ARSENIC CONENTRATIONS (mg/Kg) IN SURFICIAL SEDIMENT SAMPLES COLLECTED IN VICINITY OF VINELAND CHEMICAL SUPERFUND SITE, 1992-2007

### VINELAND POST REMEDIAL ACTION SAMPLING AND MONITORING PROGRAM, OPERABLE UNITS #3 AND #4

Sample Location	Sampling Year												
	1992*	1994*	1995*	1996*	1997*	1998*	1999*	May 2006-	May 2006-	Nov 2006-	Nov 2006-	Sept 2007-	Sept 2007-
								sed	shore	sed	shore	sed	shore
4 )Alliance Beach (mg/Kg)	11.1	9.65	7.75	1.6	2.2	4.2	6.7	1.9	ND	1.3	1.5	U	U
5) Almond Beach (mg/Kg)	7.9	2	3.85	5.3	4.35	17	2.7	2.5	1	1.4	ND	1.3	U
6)BA Beach (mg/Kg)	11.8	19.3	5.95	10.5	3.25	38	4.2	1.2	1.3	2.6	4.5	2.3	290
9) Union Lake Beach (mg/Kg)	NS	NS	NS	5	2.65	2.5	4.4	400	2.4	330	8.1	360	2.1
10) South End Union Lake Beach (	NS	NS	NS	NS	NS	17.5	8.1	160	1.4	10	2.1	18	6.5
Site Clean-up Level (mg/Kg)	20	20	20	20	20	20	20	20	20	20	20	20	20

NOTE: Shaded and bold values represent detected arsenic concentrations equivalent to or above the Site Clean-up Level of 20 mg/Kg (ppm) of arsenic for solids

NS = No sample collected; ND = not detected; below analytical detection limit

<sup>\*</sup>Historical data from (USEPA/ERTC 1999) citation

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### **APPENDIX A**

# ANALYTICAL RESULTS AND CHAIN-OF-CUSTODY (COC) FORMS FOR ARSENIC ANALYSES

### Case Narrative:

### Vineland Chemical #07090014

The National Environmental Laboratory Accreditation Conference (NELAC) is a voluntary environmental laboratory accreditation association of State and Federal agencies. NELAC established and promoted a national accreditation program that provides a uniform set of standards for the generation of environmental data that are of known and defensible quality. The EPA Region 2 Laboratory is NELAC accredited. The Laboratory tests that are accredited have met all the requirements established under the NELAC Standards.

### Comment(s):

- The contaminant concentrations for all sediment/soil samples are reported on a "dry-weight" basis.
- All field samples designated with a Matrix Spike, "MS" or Matrix Spike Duplicate, "MSD," extension were treated as environmental samples and reported as such. A separate matrix spike sample is prepared, analyzed and evaluated for each batch of 20 samples; however, the results for this sample, and other QC samples associated with each batch, are not included in the Laboratory report.
- The Laboratory did not receive the sample identified as "Almond Boring-2", collected on 9/13/07 at 11:56 a.m.

#### Data Qualifier(s):

- U- The analyte was not detected at or above the Reporting Limit.
- J- The identification of the analyte is acceptable; the reported value is an estimate.
- K- The identification of the analyte is acceptable; the reported value may be biased high.
- L- The identification of the analyte is acceptable; the reported value may be biased low.
- NJ-There is presumptive evidence that the analyte is present; the analyte is reported as a tentative identification. The reported value is an estimate.

### Reporting Limit(s):

The Laboratory was able to achieve the Contract Required Quantitation Limits (CRQLs), where applicable, for each analyte requested.

#### Method(s):

TAL Metals Analysis (As), EPA SOP C-109 (ICP/AES Method)

Approval:	0	R. Thea	Date:_	11/12/07	·
	. X/ ·				
	//				



### U.S. Environmental Protection Agency Region 2 Laboratory 2890 Woodbridge Avenue Edison, NJ 08837

Data Report: VINELAND CHEMICAL

Project Number: 07090014

Program: Y206

Project Leader: PEGGY DERRICK

Remark Codes	Explanation
U	THE ANALYTE WAS NOT DETECTED AT OR ABOVE THE REPORTING LIMIT.
J	THE IDENTIFICATION OF THE ANALYTE IS ACCEPTABLE; THE REPORTED VALUE IS AN ESTIMATE.
UJ	THE ANALYTE WAS NOT DETECTED AT OR ABOVE THE REPORTING LIMIT. THE REPORTING LIMIT IS AN ESTIMATE.
N	THERE IS PRESUMPTIVE EVIDENCE THAT THE ANALYTE IS PRESENT; THE ANALYTE IS REPORTED AS A TENTATIVE IDENTIFICATION.
NJ	THERE IS PRESUMPTIVE EVIDENCE THAT THE ANALYTE IS PRESENT; THE ANALYTE IS REPORTED AS A TENTATIVE IDENTIFICATION. THE REPORTED VALUE IS AN ESTIMATE.
R	THE PRESENCE OR ABSENCE OF THE ANALYTE CANNOT BE DETERMINED FROM THE DATA DUE TO SEVERE QUALITY CONTROL PROBLEMS. THE DATA ARE REJECTED AND CONSIDERED UNUSABLE.
K	THE IDENTIFICATION OF THE ANALYTE IS ACCEPTABLE; THE REPORTED VALUE MAY BE BIASED HIGH. THE ACTUAL VALUE IS EXPECTED TO BE LESS THAN THE REPORTED VALUE.
L	THE IDENTIFICATION OF THE ANALYTE IS ACCEPTABLE; THE REPORTED VALUE MAY BE BIASED LOW. THE ACTUAL VALUE IS EXPECTED TO BE GREATER THAN THE REPORTED VALUE.
NV	NOT VALIDATED
INC	RESULT NOT ENTERED

Project Number: 07090014

\*Sorted By Sample ID

AJ04600

Field/Station ID: MILL-SHORE-100907

Matrix: sediment

Date Received: 9/14/2007

Sample Description:

**Single Component Analyses** 

CAS Number Analyte Name

Remark\_

Result

<u>Resul</u>t

Codes

<u>Units</u>

•

Field/Station ID: MILL-SHORE-MS-100907

Date Received: 9/14/2007

Matrix: sediment

Sample Description:

Single Component Analyses

CAS Number Analyte Name

Remark\_

Codes

<u>Units</u>

3 mg/Kg

AJ04602

Field/Station ID: MILL-SHORE-MSD100907

Date Received: 9/14/2007

Matrix: sediment

Sample Description:

Single Component Analyses

CAS Number Analyte Name

Remark\_

Result

Codes

Units

AJ04603

Field/Station ID: MILL-SED-100907

Matrix: sediment

Date Received: 9/14/2007

Sample Description:

Single Component Analyses

CAS Number Analyte Name

Remark\_

Codes

**Units** 

/440-38-2

Result

Refer to Page 1 for an explanation of Remark Codes

Report Date: 11/5/2007 11:12AM

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Project Number: 07090014

\*Sorted By Sample ID

Field/Station ID: BA-SHORE-100907

Matrix: sediment

Sample Description:

Date Received: 9/14/2007

Single Component Analyses

CAS Number **Analyte Name**  Remark

Result

Codes

Units

AJ04605

Field/Station ID: BA-SED-100907

Matrix: sediment

Date Received: 9/14/2007

Sample Description:

Single Component Analyses

CAS Number

Analyte Name

Remark

Codes

Units

-- ARSENIC

Result

Result

Result

Field/Station ID: BA-SED-DUP-100907

Matrix: sediment

Date Received: 9/14/2007

Sample Description:

**Single Component Analyses** 

**CAS** Number

Analyte Name

Remark\_

Codes

**Units** 

AJ04607

Field/Station ID: BA-BEACH-100907

Date Received: 9/14/2007

Matrix: sediment

Sample Description:

Single Component Analyses

CAS Number Analyte Name Remark

Codes

Units

me/Ke

Refer to Page 1 for an explanation of Remark Codes Report Date: 11/5/2007 11:12AM

Page 3 of 22

Project Number: 07090014

\*Sorted By Sample ID

AJ04608

Field/Station ID: BA-BEACH-MS-100907

Analyte Name

Matrix: sediment

Date Received: 9/14/2007

Sample Description:

Single Component Analyses

CAS Number

Remark\_ Codes

Result

Units

Field/Station ID: BA-BEACH-MSD-100907

Matrix: sediment

Date Received: 9/14/2007

Sample Description:

**Single Component Analyses** 

Remark

CAS Number

Analyte Name

Result

Codes

**Units** 

Field/Station ID: SHERMAN-SHORE-100907

Matrix: sediment

Date Received: 9/14/2007

Sample Description:

**Single Component Analyses** 

Remark

CAS Number Analyte Name

Result

Codes

Units

Field/Station ID: SHERMAN-SED-100907

Date Received: 9/14/2007

Matrix: sediment

Sample Description:

**Single Component Analyses** 

CAS Number **Analyte Name**  Remark\_

Codes

Units

Refer to Page 1 for an explanation of Remark Codes

Report Date: 11/5/2007 11:12AM

Page 4 of 22

Project Number: 07090014

\*Sorted By Sample ID

AJ04612

Field/Station ID: SHERMAN-SEDDUP100907

Date Received: 9/14/2007

Matrix: sediment

Sample Description:

Single Component Analyses

CAS Number

Remark

Result

Codes

**Units** 

amg/Kg

AJ04613

Field/Station ID: R55-SHORE-110907

Analyte Name

Matrix: sediment

Date Received: 9/14/2007

Sample Description:

Single Component Analyses

CAS Number

Analyte Name

Remark

Codes

<u>Units</u>

.400

Result

Result

me/Ke

AJ04614

Field/Station ID: R55-SED-110907

Matrix: sediment

Date Received: 9/14/2007

Sample Description:

**Single Component Analyses** 

CAS Number Analyte Name

Remark\_

Codes

<u>Units</u>

Units

AJ04615

Field/Station ID: R55-SED-MS-110907

Date Received: 9/14/2007

Matrix: sediment

Sample Description:

Single Component Analyses

Remark CAS Number Analyte Name Result Codes

444  $38 \pm 2.5$  ARSENIC 1,200 1,200

Project Number: 07090014

\*Sorted By Sample ID

AJ04616

Field/Station ID: R55-SED-MSD-110907

Matrix: sediment

Date Received: 9/14/2007

Sample Description:

**Single Component Analyses** 

CAS Number Analyte Name

Remark\_

Codes

Units

AJ04617.

Field/Station ID: BWB-SHORE-110907

Matrix: sediment

Date Received: 9/14/2007

Sample Description:

Single Component Analyses

Remark

CAS Number

Analyte Name
ARSENIC

Result

Result

Codes

<u>Units</u>

Field/Station ID: BWB-SHORE-DUP-110907

Matrix: sediment

Date Received: 9/14/2007

Sample Description:

Single Component Analyses

CAS Number Analyte Name

Remark\_

Result

Result

Codes

Units

A.I04619

Field/Station ID: BWB-SED-110907

Date Received: 9/14/2007

Matrix: sediment

Sample Description:

**Single Component Analyses** 

CAS Number Analyte Name

Remark\_

Codes

<u>Units</u>

8-2.7 ARSENICE

Refer to Page 1 for an explanation of Remark Codes
Report Date: 11/5/2007 11:12AM

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Project Number: 07090014

\*Sorted By Sample ID

AJ04620

Field/Station ID: BWB-SED-DUP-110907

Matrix: sediment

Date Received: 9/14/2007

Sample Description:

Single Component Analyses

CAS Number Analyte Name

Remark\_

Codes

Result

Result

Result

<u>Units</u>

AJ04621/

Field/Station ID: ALLIANCE-SHORE110907

Matrix: sediment

Date Received: 9/14/2007

Sample Description:

Single Component Analyses

CAS Number Analyte Name

alyte Name

Remark\_

Codes

**Units** 

14 July 1

J04622

Field/Station ID: ALLIANCE-SED-110907

Matrix: sediment

Date Received: 9/14/2007

Sample Description:

**Single Component Analyses** 

CAS Number Analyte Name

Remark\_

Codes

<u>Units</u>

AJ04623

Field/Station ID: ALLIANCE-BEACH110907

Date Received: 9/14/2007

Matrix: sediment

Sample Description:

Single Component Analyses

CAS Number Analyte Name

Result

Remark\_

Codes

<u>Units</u>

Refer to Page 1 for an explanation of Remark Codes

Report Date: 11/5/2007 11:12AM

Page 7 of 22

Project Number: 07090014

\*Sorted By Sample ID

AJ04624

Field/Station ID: ALLIANCE-BEACH-DUP11

Date Received: 9/14/2007

Matrix: sediment

Sample Description:

CAS Number Analyte Name

**Single Component Analyses** 

Remark\_

Codes

Units

mo/Kb

Field/Station ID: ALMOND-SHORE-110907

Date Received: 9/14/2007

Matrix: sediment

Sample Description:

Single Component Analyses

Remark

0.74U

Codes **Units** 

CAS Number

Analyte Name

Field/Station ID: ALMOND-SHORE-DUP1109

Date Received: 9/14/2007

Matrix: sediment

Sample Description:

**Single Component Analyses** 

Remark\_

CAS Number Analyte Name Result

Result

Result

Codes **Units** 

AJ04627

Field/Station ID: ALMOND-SED-110907

ARSENIG

Date Received: 9/14/2007

Matrix: sediment

Sample Description:

Single Component Analyses

Remark

CAS Number Analyte Name Result

Codes

Units

Refer to Page 1 for an explanation of Remark Codes

Report Date: 11/5/2007 11:12AM

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Project Number: 07090014

\*Sorted By Sample ID

AJ04628

Field/Station ID: ALMOND-BEACH-110907

Date Received: 9/14/2007

Matrix: sediment

Sample Description:

Single Component Analyses

CAS Number Analyte Name

Remark\_

Result

Codes

<u>Units</u>

mg/Kg

AJ04629

Field/Station ID: NUL-SHORE-120907

Matrix: sediment

Date Received: 9/14/2007

Sample Description:

Single Component Analyses

CAS Number Analyte Name

Remark

Codes

Units

440538-2 - ARSENI(

Result

Result

AJ04630

Field/Station ID: NUL-SED-120907

Matrix: sediment

Date Received: 9/14/2007

Sample Description:

Single Component Analyses

CAS Number Analyte Name

Remark\_

Codes

<u>Units</u>

Units

AJ04631

Field/Station ID: ULB-SHORE-120907

Date Received: 9/14/2007

Matrix: sediment

Sample Description:

Single Component Analyses

Remark

<u>CAS Number Analyte Name</u> Result <u>Codes</u>

7440-38-2 - T ARSENIC 2.1 mg/Kg 38-

Refer to Page 1 for an explanation of Remark Codes

Report Date: 11/5/2007 11:12AM

Page 9 of 22

Project Number: 07090014

\*Sorted By Sample ID

**Units** 

AJ04632

Field/Station ID: ULB-SED-120907

Matrix: sediment

Sample Description:

Date Received: 9/14/2007

**Single Component Analyses** 

CAS Number Analyte Name Remark\_

Result

Codes

AJ04633

Field/Station ID: ULB-BRACH-120907

Matrix: sediment

Date Received: 9/14/2007

Sample Description:

**Single Component Analyses** 

Remark\_

CAS Number

Analyte Name

Result

Result

Codes

Units mg/Kg

Field/Station ID: ULB-BRACH-MS-120907

Matrix: sediment

Sample Description:

Date Received: 9/14/2007

**Single Component Analyses** 

**CAS Number** 

Analyte Name

Remark\_

Codes

**Units** 

Field/Station ID: ULB-BRACH-MSD-120907

Date Received: 9/14/2007

Matrix: sediment

Sample Description:

**Single Component Analyses** 

Remark

CAS Number Analyte Name Result

Codes

**Units** 

Refer to Page 1 for an explanation of Remark Codes

Report Date: 11/5/2007 11:12AM

Page 10 of 22

Project Number: 07090014

\*Sorted By Sample ID

AJ04636

Field/Station ID: SUL-SHORE-120907

Matrix: sediment

Date Received: 9/14/2007

Sample Description:

**Single Component Analyses** 

CAS Number Analyte Name

Remark\_ Result

Codes

**Units** 

Field/Station ID: SUL-SED-120907

Matrix: sediment

Date Received: 9/14/2007

Sample Description:

Single Component Analyses

CAS Number Analyte Name Remark\_

Codes

Result

Result

Result

**Units** 

Field/Station ID: SUL-BEACH-120907

Matrix: sediment

Date Received: 9/14/2007

Sample Description:

Single Component Analyses

CAS Number

Analyte Name

Remark

Codes

**Units** 

Field/Station ID: SUL-BEACH-DUP-120907

Date Received: 9/14/2007

Matrix: sediment

Sample Description:

**Single Component Analyses** 

Refer to Page 1 for an explanation of Remark Codes

**CAS Number** Analyte Name Remark\_

Codes

**Units** 

Report Date: 11/5/2007 11:12AM

Project Number: 07090014

\*Sorted By Sample ID

AJ04640

Field/Station ID: SUL-BORING-1

Matrix: sediment

Sample Description:

Date Received: 9/14/2007

**Single Component Analyses** 

CAS Number

Analyte Name

Remark Codes Result

**Units** 

AJ04641

Field/Station ID: ALMOND-BORING-1

Matrix: sediment

Date Received: 9/14/2007

Sample Description:

Single Component Analyses

CAS Number Analyte Name Remark Codes

Result

**Units** 

Field/Station ID: RSS-BORING-1

Matrix: sediment

Sample Description:

Date Received: 9/14/2007

Single Component Analyses

CAS Number Analyte Name Remark

Result

Codes

**Units** 

AJ04643

Field/Station ID: RSS-BORING-2

Date Received: 9/14/2007

Matrix: sediment

Sample Description:

**Single Component Analyses** 

CAS Number **Analyte Name**  Remark\_

Codes

**Units** 

Refer to Page 1 for an explanation of Remark Codes

Report Date: 11/5/2007 11:12AM

Page 12 of 22

Project Number: 07090014

\*Sorted By Sample ID

AJ04644

Field/Station ID: RSS-BORING-3

ARSENIC

Date Received: 9/14/2007

Matrix: sediment

Sample Description:

Single Component Analyses

Remark\_

CAS Number Analyte Name

Result

Codes

Units

AJ04645

Field/Station ID: ALLIANCE-BORING -1

Date Received: 9/14/2007

Matrix: sediment

Sample Description:

Single Component Analyses

Remark

CAS Number 7440-38-2

Analyte Name

Result

Codes

**Units** 

AJ04646

Field/Station ID: ULB-BORING-1

Matrix: sediment

Date Received: 9/14/2007

Sample Description:

Single Component Analyses

Remark

CAS Number

Analyte Name

Result

Codes

Units

Field/Station ID: MILL-BORING-1

Date Received: 9/14/2007

Matrix: sediment

Sample Description:

**Single Component Analyses** 

Remark\_

CAS Number Analyte Name Result

Codes

**Units** 

Refer to Page 1 for an explanation of Remark Codes

Report Date: 11/5/2007 11:12AM

Page 13 of 22

Project Number: 07090014

\*Sorted By Sample ID

AJ04648

Field/Station ID: MILL-BORING-2

Matrix: sediment

Sample Description:

Date Received: 9/14/2007

Single Component Analyses

CAS Number Analyte Name

Remark\_

Codes

**Units** me/Ke

AJ04649

Field/Station ID: MILL-BORING-3

Matrix: sediment

Date Received: 9/14/2007

Sample Description:

Single Component Analyses

CAS Number

Analyte Name

Remark\_

Result

Codes

**Units** 

Field/Station ID: MILL-WAT1-100907

Matrix: Aqueous

Sample Description:

Date Received: 9/14/2007

Single Component Analyses

CAS Number

Analyte Name

Result

Remark\_

Codes

**Units** 

Field/Station ID: MILL-WAT2-100907

Matrix: Aqueous

Sample Description:

Date Received: 9/14/2007

**Single Component Analyses** 

CAS Number Analyte Name Result

Remark\_ Codes

**Units** 

Refer to Page 1 for an explanation of Remark Codes

Report Date: 11/5/2007 11:12AM

Page 14 of 22

Project Number: 07090014

\*Sorted By Sample ID

Field/Station ID: BA-WAT1-100907

Date Received: 9/14/2007

Matrix: Aqueous

Sample Description:

Single Component Analyses

CAS Number Analyte Name

Remark\_ Result

Codes

Units

AJ04653

Field/Station ID: BA-WAT2-100907

AIRSIÐNIE!

Matrix: Aqueous

Date Received: 9/14/2007

Sample Description:

Single Component Analyses

CAS Number Analyte Name Remark\_

Codes

**Units** 

Field/Station ID: SHERMAN-WAT1-100907

Matrix: Aqueous

Date Received: 9/14/2007

Sample Description:

**Single Component Analyses** 

**CAS Number** Analyte Name

Result

Result

Remark

Codes

**Units** 

 $\Delta J04655$ 

Field/Station ID: SHERMAN-WAT2-100907

Matrix: Aqueous

Date Received: 9/14/2007

Sample Description:

**Single Component Analyses** 

Remark\_ CAS Number Analyte Name Codes Result

Units

Refer to Page 1 for an explanation of Remark Codes

Report Date: 11/5/2007 11:12AM

Page 15 of 22

Project Number: 07090014

\*Sorted By Sample ID

AJ04656

Field/Station ID: R55-WAT1-110907

Matrix: Aqueous

Sample Description:

Date Received: 9/14/2007

**Single Component Analyses** 

CAS Number Analyte Name Remark\_ Codes

Result

Result

Codes

Units

**Units** 

Field/Station ID: R55-WAT2-110907

Matrix: Aqueous

Date Received: 9/14/2007

Sample Description:

**Single Component Analyses** 

Remark\_ Codes

Result **Units** CAS Number Analyte Name

Field/Station ID: BWB-WAT1-110907

Matrix: Aqueous

Date Received: 9/14/2007

Sample Description:

Single Component Analyses

Remark\_

CAS Number Analyte Name

Field/Station ID: BWB-WAT2-110907

Date Received: 9/14/2007

Matrix: Aqueous

Sample Description:

**Single Component Analyses** 

Remark\_ Codes Analyte Name

CAS Number **Units** 

Refer to Page 1 for an explanation of Remark Codes

Report Date: 11/5/2007 11:12AM

Page 16 of 22

Project Number: 07090014

\*Sorted By Sample ID

AJ04660

Field/Station ID: BWB-WAT2-MS-110907

Matrix: Aqueous

Sample Description:

Date Received: 9/14/2007

Single Component Analyses

CAS Number

Analyte Name ARSENIC

Remark\_

Result

Result

Codes

**Units** 

AJ04661

Field/Station ID: BWB-WAT2-MSD-110907

Matrix: Aqueous

Date Received: 9/14/2007

Sample Description:

**Single Component Analyses** 

CAS Number Analyte Name Remark\_

Codes

8.0U

**Units** 

Units

Field/Station ID: ALLIANCE-WAT1-100907

Matrix: Aqueous

Date Received: 9/14/2007

Sample Description:

Single Component Analyses

CAS Number Analyte Name

Remark Result

Codes

Field/Station ID: ALLIANCE-WAT2-100907

Date Received: 9/14/2007

Matrix: Aqueous

Sample Description:

**Single Component Analyses** 

**CAS Number Analyte Name**  Remark\_

Codes

Result

Units

Refer to Page 1 for an explanation of Remark Codes Report Date: 11/5/2007 11:12AM

Page 17 of 22

Project Number: 07090014

\*Sorted By Sample ID

Units

AJ04664

Field/Station ID: ALMOND-WAT1-110907

Matrix: Aqueous

Date Received: 9/14/2007

Sample Description:

**Single Component Analyses** 

CAS Number Analyte Name

Remark\_

Codes

Field/Station ID: ALMOND-WAT2-110907

Matrix: Aqueous

Date Received: 9/14/2007

Sample Description:

**Single Component Analyses** 

Remark

CAS Number

Analyte Name

Result

Result

Codes

**Units** 

AJ04666

Field/Station ID: NUL-WAT1-120907

Matrix: Aqueous

Sample Description:

Date Received: 9/14/2007

Single Component Analyses

CAS Number Analyte Name Remark

Result

Codes

**Units** 

Field/Station ID: NUL-WAT1-DUP-120907

Date Received: 9/14/2007

Matrix: Aqueous

Sample Description:

**Single Component Analyses** 

Remark\_ CAS Number Analyte Name Codes

**Units** ug/L

Refer to Page 1 for an explanation of Remark Codes

Report Date: 11/5/2007 11:12AM

Page 18 of 22

Project Number: 07090014

\*Sorted By Sample ID

AJ04668

Field/Station ID: NUL-WAT2-120907

Date Received: 9/14/2007

Matrix: Aqueous

Sample Description:

Single Component Analyses

CAS Number Analyte Name

Remark\_

Result

Result

Result

Codes

<u>Units</u>

AJ04669

Field/Station ID: ULB-WAT1-120907

Matrix: Aqueous

Date Received: 9/14/2007

Sample Description:

Single Component Analyses

CAS Number Analyte Name

Remark\_

Codes

<u>Units</u>

- 12

AJ04670

Field/Station ID: ULB-WAT2-120907

Matrix: Aqueous

Date Received: 9/14/2007

Sample Description:

Single Component Analyses

CAS Number Analyte Name

Remark

C-1--

Codes

<u>Units</u>

AJ04671

Field/Station ID: ULB-WAT2-DUP-120907

Date Received: 9/14/2007

Matrix: Aqueous

Sample Description:

**Single Component Analyses** 

CAS Number Analyte Name

Remark\_

Codes

<u>Units</u>

/44U-30-

ARSENI

270

Result

49 H

Refer to Page 1 for an explanation of Remark Codes

Report Date: 11/5/2007 11:12AM

Page 19 of 22

Project Number: 07090014

\*Sorted By Sample ID

Field/Station ID: SUL-WAT1-120907

Date Received: 9/14/2007

Matrix: Aqueous

Sample Description:

**Single Component Analyses** 

CAS Number Analyte Name

Remark\_

Codes

**Units** 

Field/Station ID: SUL-WAT2-120907

Date Received: 9/14/2007

Matrix: Aqueous

Sample Description:

Single Component Analyses

Remark\_

CAS Number

Analyte Name

Result

Result

Codes **Units** 

Field/Station ID: PBlank-01-100907

Matrix: Aqueous

Sample Description:

Date Received: 9/14/2007

Single Component Analyses

Remark

CAS Number

Analyte Name

Result

Codes

**Units** 

Field/Station ID: PBlank-02-110907

Date Received: 9/14/2007

Matrix: Aqueous

Sample Description:

**Single Component Analyses** 

Remark\_

CAS Number

Analyte Name

Result

Codes

**Units** 

Refer to Page 1 for an explanation of Remark Codes

Report Date: 11/5/2007 11:12AM

Page 20 of 22

Project Number: 07090014

\*Sorted By Sample ID

AJ04676

Field/Station ID: PBlank-03-120907

Date Received: 9/14/2007

Matrix: Aqueous

Sample Description:

**Single Component Analyses** 

CAS Number Analyte Name

Result

Result

Result

Remark\_ Codes

<u>Units</u>

AJ04677

Field/Station ID: BSBlank-01-100907

Matrix: Aqueous

Date Received: 9/14/2007

Sample Description:

**Single Component Analyses** 

CAS Number

Analyte Name

Remark\_

Codes

Units ug/ls

AJ04678

Field/Station ID: BSBlank-02-110907

Matrix: Aqueous

Date Received: 9/14/2007

Sample Description:

**Single Component Analyses** 

CAS Number Analyte Name

Remark

(

Remark

Codes Units

**Units** 

A.104679

Field/Station ID: BSBlank-03-120907

Date Received: 9/14/2007

Matrix: Aqueous

Sample Description:

**Single Component Analyses** 

<u>CAS Number</u> <u>Analyte Name</u> <u>Result</u> <u>Codes</u>

Refer to Page 1 for an explanation of Remark Codes

Report Date: 11/5/2007 11:12AM Page 21 of 22

Project Number: 07090014

\*Sorted By Sample ID

AJ04680

Field/Station ID: BrlBlank-120907

Date Received: 9/14/2007

Matrix: Aqueous

Sample Description:

Single Component Analyses

CAS Number Analyte Name Remark\_

Codes

**Units** 

ug/L

**Units** 

Field/Station ID: TTBlank-120907

Matrix: Aqueous

Date Received: 9/14/2007

Sample Description:

Single Component Analyses

Remark\_

Codes Result

Result

CAS Number Analyte Name

**Project Approval:** 

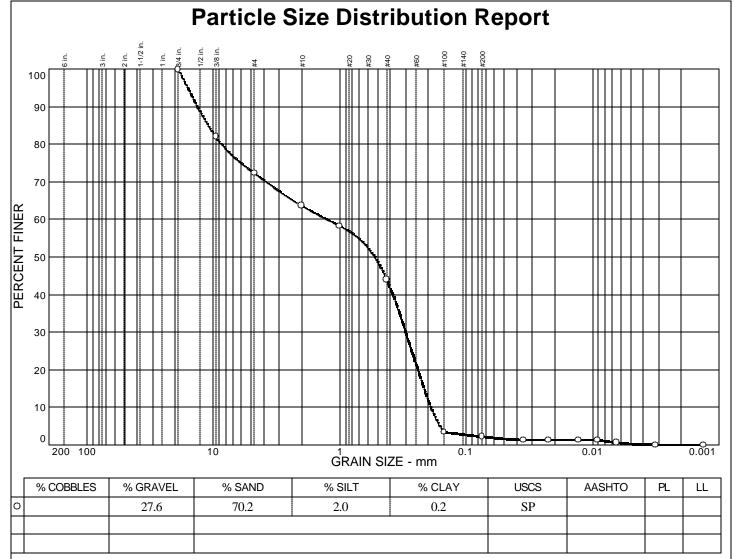
Date: //-12-07 Refer to Page 1 for an explanation of Remark Codes

Report Date: 11/5/2007-11:12AM

Page 22 of 22

### **APPENDIX B**

# ANALYTICAL RESULTS AND CHAIN-OF-CUSTODY (COC) FORMS FOR GRAIN SIZE ANALYSES



SIEVE	Pl	ERCENT FINE	ER .	
inches size	0			
3/4 3/8	100.0 82.0			
$>\!\!<$		GRAIN SIZE		
D <sub>60</sub>	1.25			
D <sub>30</sub>	0.302			
D <sub>10</sub>	0.190			
><	COEFFICIENTS			
C <sub>c</sub>	0.38			
C <sub>u</sub>	6.59			
o Cource: Alliance Poring 1				

SIEVE	PE	RCENT FINE	ĒR
number size	0		
#4 #10 #18 #40 #100 #200	72.4 63.7 58.3 44.0 3.4 2.2		

SOIL	DES	CRIP	HON

O Grayish Tan, SAND, Some Gravel

### REMARKS:

O Natural Moisture: 11.9%

F.M.=1.42

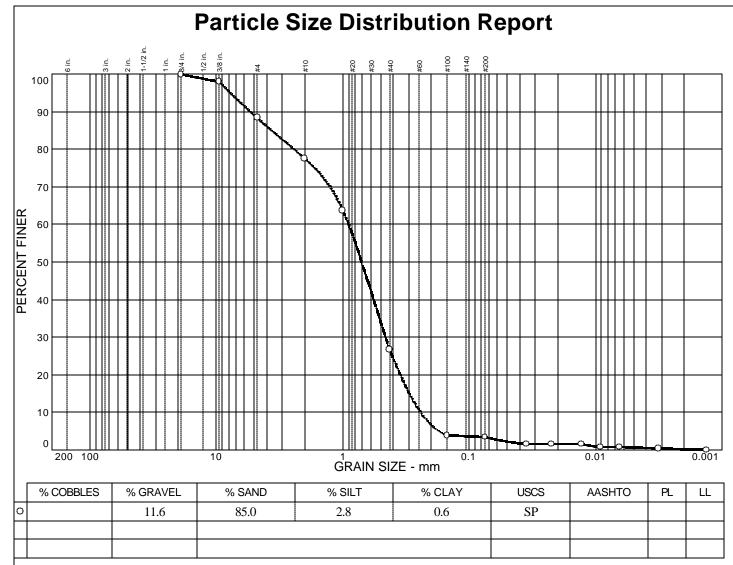
o Source: Alliance-Boring-1

E2CR, Inc.

Client: EA Engineering

Project: Vineland

Project No.: 07519-04



SIEVE	PE	ERCENT FINE	ER
inches size	0		
3/4 3/8	100.0 98.0		
		GRAIN SIZE	
D <sub>60</sub>	0.901		
D <sub>30</sub>	0.459		
D <sub>10</sub>	0.242		
>	COEFFICIENTS		
C <sub>c</sub>	0.97		
C <sub>u</sub>	3.72		

SIEVE	PE	RCENT FINE	ΞR
number size	0		
#4 #10 #18 #40 #100 #200	88.4 77.6 63.7 26.8 3.9 3.4		

O Tan, SAND, little Gravel

SOIL DESCRIPTION

REMARKS:

O	Natural Moisture : 13.7%	
	F.M.=1.10	

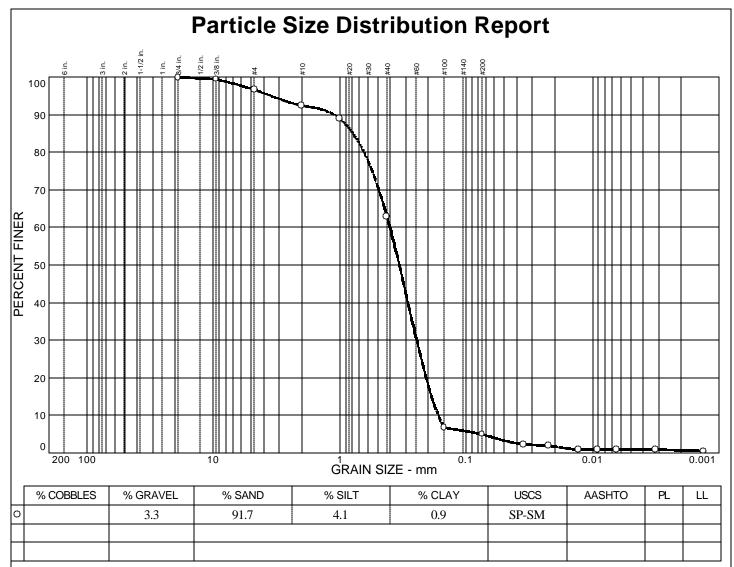
o Source: Almond-Boring-1

E2CR, Inc.

Client: EA Engineering

Project: Vineland

Project No.: 07519-04



0 100.0 99.5		
77.0		
	GRAIN SIZE	
0.403		
0.248		
0.167		
О	OEFFICIENT	S
0.91		
2.41		
1	0.403 0.248 0.167 0.91 2.41	0.248 0.167 COEFFICIENT 0.91

SIEVE	PE	RCENT FINE	ĒR
number size	0		
#4 #10 #18 #40 #100 #200	96.7 92.5 89.0 62.9 6.7 5.0		

SOIL DESCRIPTION	
O Tan and Dark Brown	S /

REMARKS:

O Natural Moisture : 48.8%

F.M.=0.97

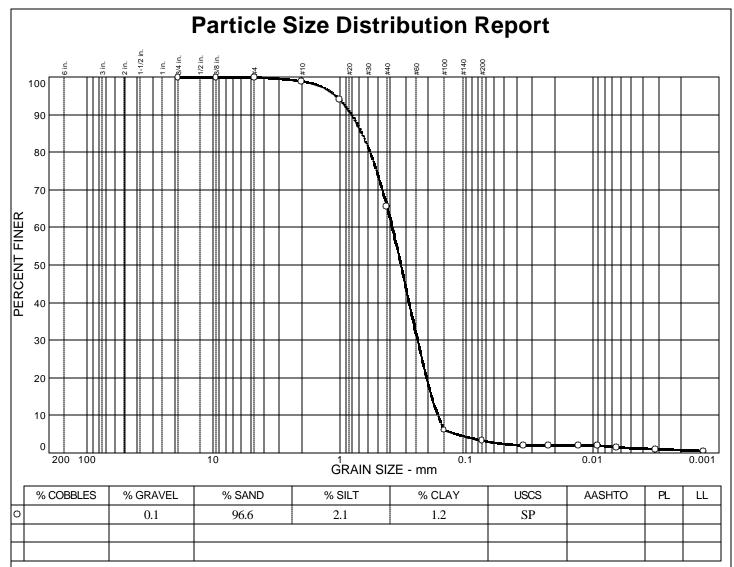
o Source: Mill-Boring-1

E2CR, Inc.

Client: EA Engineering

Project: Vineland

Project No.: 07519-04



SIEVE	PERCENT FINER			
inches size	0			
3/4	100.0			
3/8	100.0			
><		GRAIN SIZE		
D <sub>60</sub>	0.386			
D <sub>30</sub>	0.244			
D <sub>10</sub>	0.168			
><	COEFFICIENTS			
C <sub>C</sub>	0.92			
C <sub>u</sub>	2.30			
o Source: N	1.11 D .	2	•	

SIEVE	PE	RCENT FINE	ER
number size	0		
#44 #10 #18 #40 #100 #200	99.9 98.9 94.1 65.6 6.1 3.3		

SOIL DESCRIPTION

O Tannish Brown, SAND

# REMARKS: O natural Moisture : 17.3% F.M.=0.94

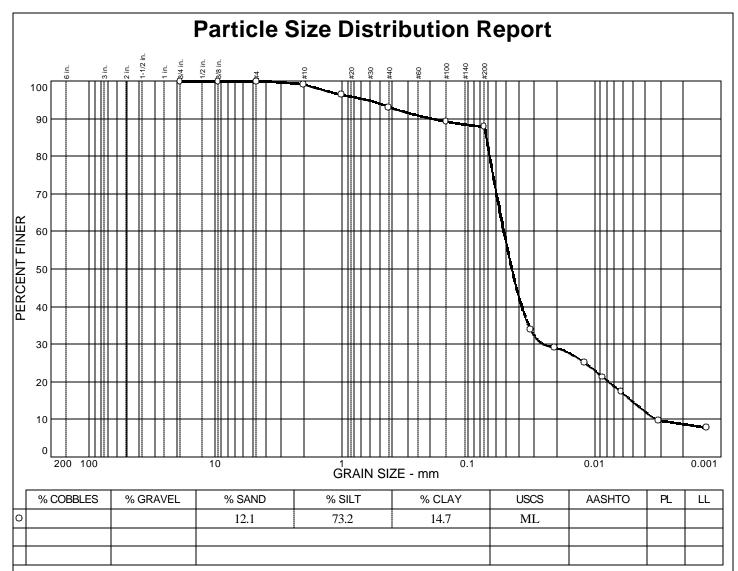
o Source: Mill-Boring-3

E2CR, Inc.

Client: EA Engineering

Project: Vineland

Project No.: 07519-04



SIEVE	PERCENT FINER			
inches size	0			
3/4 3/8	100.0 100.0			
5/0	100.0			
><		GRAIN SIZE	Ē	
D <sub>60</sub>	0.0526			
D <sub>30</sub>	0.0254			
D <sub>10</sub>	0.0032			
$>\!<$	COEFFICIENTS			
C <sub>C</sub>	3.78			
C <sub>u</sub>	16.28			
o Source: I	) 55 Di	_ 1		

SIEVE	PERCENT FINER			
number size	0			
#4 #10 #18 #40 #100 #200	100.0 99.2 96.4 93.1 89.3 87.9			

SOIL	DESCRIP	TION

O Dark Brown, Clayey SILT, little Sand, Wood, trace to little unknown silvery metal

### REMARKS:

O Natural Moisture : 642.9% F.M.=0.11

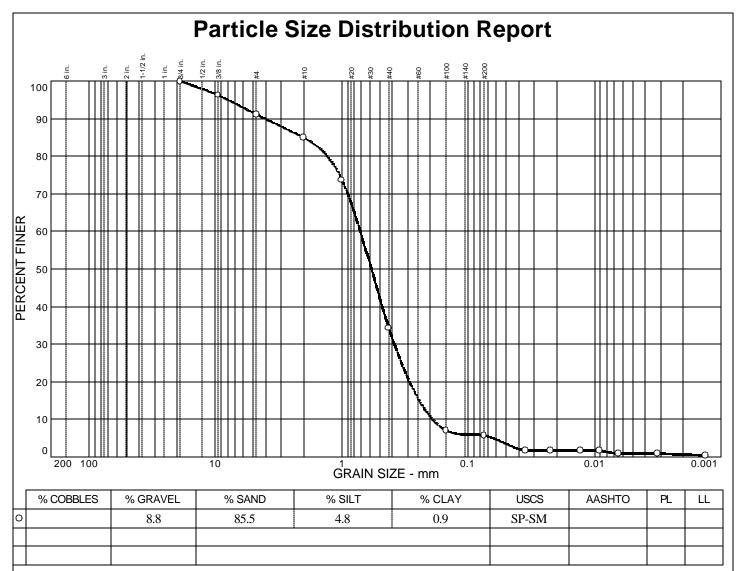
○ Source: R-55-Boring-1

E2CR, Inc.

Client: EA Engineering

Project: Vineland

Project No.: 07519-04



PERCENT FINER			
0			
100.0 96.3			
	GRAIN SIZE		
0.717			
0.385			
0.191			
COEFFICIENTS			
1.08			
3.74			
	0.717 0.385 0.191	GRAIN SIZE  0.717  0.385  0.191  COEFFICIENT  1.08	

SIEVE	PE	RCENT FINE	ĒR
number size	0		
#4 #10 #18 #40 #100 #200	91.2 85.0 73.7 34.3 7.1 5.7		

	SOIL DESCRIPTION
	O Gray and Dark Brown, SAND, little Gravel,
ı	W/

### REMARKS:

O Natural Moisture : 58.2% F.M.=1.05

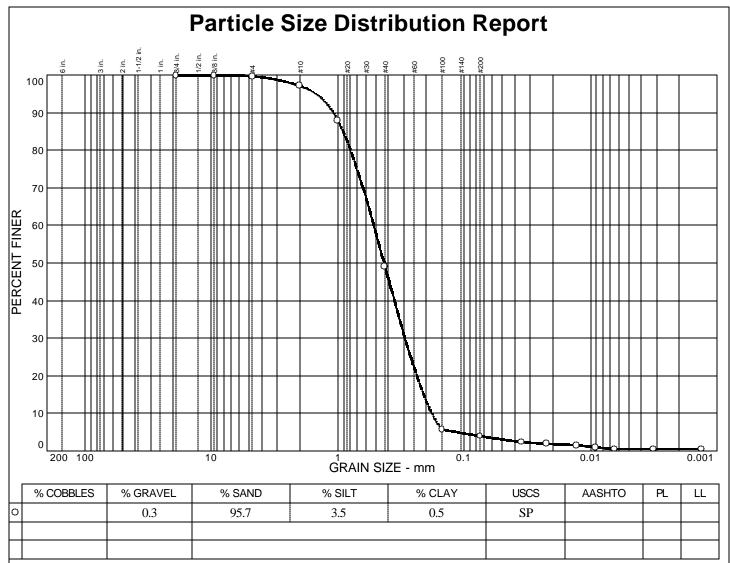
○ Source: R-55-Boring-3

E2CR, Inc.

Client: EA Engineering

Project: Vineland

Project No.: 07519-04



SIEVE	Pl	ERCENT FINE	-R	
inches size	0			
3/4	100.0			
3/8	100.0			
><		GRAIN SIZE		
D <sub>60</sub>	0.521			
D <sub>30</sub>	0.295			
D <sub>10</sub>	0.181			
$>\!<$	COEFFICIENTS			
C <sub>c</sub>	0.93			
C <sub>u</sub>	2.88			
- C	Sul-Boring	1		

SIEVE	PE	RCENT FINE	ER
number size	0		
#4 #10 #18 #40 #100 #200	99.7 97.2 87.9 49.1 5.7 4.0		

SOIL	DESC	<u>,KII</u>	<u> </u>	<u> </u>	

O Gray and Yellowish Brown, SAND

### REMARKS:

O Natural Moisture : 17.4% F.M.=0.95

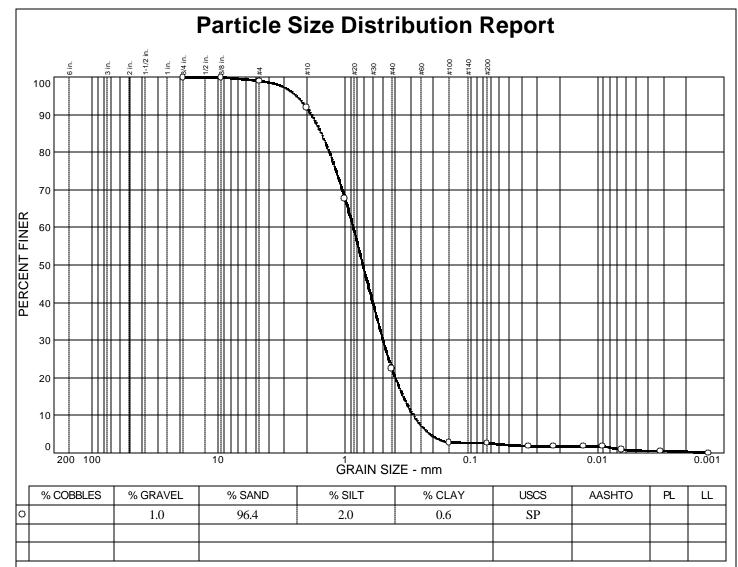
o Source: Sul-Boring-1

E2CR, Inc.

Client: EA Engineering

Project: Vineland

Project No.: 07519-04



PERCENT FINER		
0		
100.0		
100.0		
GRAIN SIZE		
0.862		
0.500		
0.286		
COEFFICIENTS		
1.01		
3.01		
	0.862 0.500 0.286	100.0 100.0 100.0 GRAIN SIZE 0.862 0.500 0.286 COEFFICIENT

SIEVE	PERCENT FINER		
number size	0		
#4 #10 #18 #40 #100 #200	99.0 91.9 67.7 22.5 2.7 2.6		

	<u> </u>
	O Tan, SAND
	REMARKS:
	O Natural Moisture: 15.2%
	F.M.=0.98
ı	

SOIL DESCRIPTION

○ Source: ULB-Boring-1

E2CR, Inc.

Client: EA Engineering

Project: Vineland

Project No.: 07519-04

### **APPENDIX C**

# ANALYTICAL RESULTS AND CHAIN-OF-CUSTODY (COC) FORMS FOR PCBs ANALYSES

ATTACHMENT 1 SOM01.2/Aroclors SOP NO. HW-37

Page 1 of 5

### **Functional Guidelines for Evaluating Organic Analysis**

CASE No.:

36799

SDG No.: B4FB6 & B4F84

LABORATORY:

**DATACHEM** 

SITE: Vineland Chemicals

SAMPLER: USACE

**ANALYSIS: PCB** 

### **DATA ASSESSMENT**

The current SOP HW-37 (Revision 1) August 2007, USEPA Region II Data Validation SOP for Statement of Work SOM01.2 for evaluating organic data have been applied.

All data are valid and acceptable except those analytes rejected "R"(unusable). Due to the detection of QC problems, some analytes may have the "J" (estimated), "N"(presumptive evidence for the presence of the material), "U" (non-detect) or "JN" (presumptive evidence for the presence of the material at an estimated value) flag. All action is detailed on the attached sheets.

The "R" flag means that the associated value is unusable. In other words, significant data bias is evident and the reported analyte concentration is unreliable.

Reviewer's

Signature:

Peer Reviewer's

Signature:

Shobitha/Amin

Date: November / 05 /2007

Verified By:

#### SDG#B4FB6

#### 1. HOLDING TIME:

The amount of an analyte in a sample can change with time due to chemical instability, degradation, volatilization, etc. If the specified holding time is exceeded, the data may not be valid. Those analytes detected in the samples whose holding time has been exceeded will be qualified as estimated, "J". The non-detects (sample quantitation limits) will be flagged as estimated, "J", or unusable, "R", if the holding times are grossly exceeded.

The following action was taken in the samples and analytes shown due to excessive holding time.

No problems found for this qualification.

### 2. SURROGATES

All samples are spiked with surrogate compounds prior to sample preparation to evaluate overall laboratory performance and efficiency of the analytical technique. If the measured surrogate concentrations were outside contract specifications, qualifications were applied to the samples and analytes as shown below.

No problems found for this qualification.

### 3. MATRIX SPIKE/SPIKE DUPLICATE, MS/MSD:

No problems found for this qualification.

Laboratory Control Samples (LCS):

The LCSs data provides information on the accuracy of the analytical method and laboratory performance. If LCS recoveries fell outside of the acceptable limits, qualifications were applied to the associated samples and compounds as shown below.

No problems found for this qualification.

#### 4. BLANK CONTAMINATION:

Quality assurance (QA) blanks, i.e., method, field, or rinse blanks are prepared to identify any contamination, which may have been introduced into the samples during sample preparation or field activity. Method blanks measure laboratory contamination. Field and rinse blanks measure cross-contamination of samples during field operations. Depending on the concentration of the analyte in the blank, the analytes are qualified as non-detects U.

The following analytes in the sample shown were qualified with "U" for these reasons:

#### A) Method blank contamination:

No problems found for this qualification.

### B) Field or rinse blank contamination:

#### 5. CALIBRATION:

Satisfactory instrument calibration is established to ensure that the instrument is capable of producing acceptable quantitative data. An initial calibration demonstrates that the instrument is capable of giving acceptable performance at the beginning of an experimental sequence. The continuing calibration checks document that the instrument is giving satisfactory daily performance.

A) Percent Relative Standard Deviation (%RSD) and Percent Difference (%D):

For the PCB fraction, if %RSD exceeds 20% for all analytes and the two surrogates, qualify all associated positive results "J" and non-detects "UJ".

For opening CCV, or closing CCV that is used as an opening CCV for the next 12-hour period, if %D exceeds 15% for analytes and the two surrogates, qualify all associated positive results "J" and non-detects "UJ".

For closing CCV, if %D exceeds 50% for all analytes and the two surrogates, qualify all associated positive results "J" and non-detects "UJ".

The following analytes in the sample shown were qualified for %RSD and %D:

No problems for this qualification.

- 6. COMPOUND IDENTIFICATION:
- A) PCB Fraction:

The retention times of reported compounds must fall within the calculated retention time windows for the two chromatographic columns and a GC/MS confirmation is required if the concentration exceeds 10ng/ml in the final sample extract.

- CONTRACT PROBLEMS NON-COMPLIANCE:
- 8. FIELD DOCUMENTATION:
- 9. OTHER PROBLEMS:
- 10. This package contains re-extractions, re-analyses or dilutions. Upon reviewing the QA results, the following Form 1(s) are identified NOT to be used.

#### SDG#B4F84

#### 1. HOLDING TIME:

The amount of an analyte in a sample can change with time due to chemical instability, degradation, volatilization, etc. If the specified holding time is exceeded, the data may not be valid. Those analytes detected in the samples whose holding time has been exceeded will be qualified as estimated, "J". The non-detects (sample quantitation limits) will be flagged as estimated, "J", or unusable, "R", if the holding times are grossly exceeded.

The following action was taken in the samples and analytes shown due to excessive holding time.

No problems found for this qualification.

#### 2. SURROGATES

All samples are spiked with surrogate compounds prior to sample preparation to evaluate overall laboratory performance and efficiency of the analytical technique. If the measured surrogate concentrations were outside contract specifications, qualifications were applied to the samples and analytes as shown below.

No problems found for this qualification.

### 3. MATRIX SPIKE/SPIKE DUPLICATE, MS/MSD:

No problems found for this qualification.

### Laboratory Control Samples (LCS):

The LCSs data provides information on the accuracy of the analytical method and laboratory performance. If LCS recoveries fell outside of the acceptable limits, qualifications were applied to the associated samples and compounds as shown below.

No problems found for this qualification.

### 4. BLANK CONTAMINATION:

Quality assurance (QA) blanks, i.e., method, field, or rinse blanks are prepared to identify any contamination, which may have been introduced into the samples during sample preparation or field activity. Method blanks measure laboratory contamination. Field and rinse blanks measure cross-contamination of samples during field operations. Depending on the concentration of the analyte in the blank, the analytes are qualified as non-detects U.

The following analytes in the sample shown were qualified with "U" for these reasons:

### A) Method blank contamination:

No problems found for this qualification.

#### B) Field or rinse blank contamination:

#### 5. CALIBRATION:

Satisfactory instrument calibration is established to ensure that the instrument is capable of producing acceptable quantitative data. An initial calibration demonstrates that the instrument is capable of giving acceptable performance at the beginning of an experimental sequence. The continuing calibration checks document that the instrument is giving satisfactory daily performance.

A) Percent Relative Standard Deviation (%RSD) and Percent Difference (%D):

For the PCB fraction, if %RSD exceeds 20% for all analytes and the two surrogates, qualify all associated positive results "J" and non-detects "UJ".

For opening CCV, or closing CCV that is used as an opening CCV for the next 12-hour period, if %D exceeds 15% for analytes and the two surrogates, qualify all associated positive results "J" and non-detects "UJ".

For closing CCV, if %D exceeds 50% for all analytes and the two surrogates, qualify all associated positive results "J" and non-detects "UJ".

The following analytes in the sample shown were qualified for %RSD and %D:

No problems for this qualification.

- 6. COMPOUND IDENTIFICATION:
- A) PCB Fraction:

The retention times of reported compounds must fall within the calculated retention time windows for the two chromatographic columns and a GC/MS confirmation is required if the concentration exceeds 10ng/ml in the final sample extract.

- 7. CONTRACT PROBLEMS NON-COMPLIANCE:
- 8. FIELD DOCUMENTATION:
- 9. OTHER PROBLEMS:
- 10. This package contains re-extractions, re-analyses or dilutions. Upon reviewing the QA results, the following Form 1(s) are identified NOT to be used.

B4F85	

Lab Name: Da	taChem Laboratories, Inc.	Contract:	EP-W-05-026	
	TAC Case No.: 36799 Mod. Ref			
Matrix: (SO	L/SED/WATER) WATER	Lab Sample	■ ID: 7256022002	
	ol: 1000 (g/mL) mL		ID: 20070917A069,200709	7B069
% Moisture:	Decanted: (Y/N)	-	ived: 09/12/2007	
Extraction:	/m		acted: 09/14/2007	
Concentrated	Extract Volume: 5000 (uL)			
	lume: 2.0 (uL) GPC Factor:			
	4 4		eanup: (Y/N) N	
	: (Y/N) <u>Y</u>	· · · · · · · · · · · · · · · · · · ·		
CAS NO.	COMPOUND	,	CONCENTRATION UNITS: (ug/L or ug/kg) ug/L	Q
12674-11-2	Aroclor-1016		0.50	IJ
11104-28-2	Aroclor-1221		0.50	U
11141-16-5	Aroclor-1232		0.50	
53469-21-9	Aroclor-1242			U.
12672-29-6	Aroclor-1248		0.50	U
11097-69-1	Aroclor-1254		0.50	Ü
11096-82-5	Aroclor-1260		0.50	U
37324-23-5	Aroclor-1262		0.50	U
	Arodlor-1269			Ü
	FH OCTO1 - 1208		0.50	U

B4F84	
D#1 0 #	

	· · ·			
Lab Name: Da	taChem Laboratories, Inc.	Contract	EP-W-05-026	
Lab Code: DA	TAC Case No.: 36799 Mod. Ref	No.:	SDG No.: B4F84	
the second secon			le ID: <u>7256022001</u>	•
	l: <u>1000</u> (g/mL) <u>mL</u>			17B068
	Decanted: (Y/N)		eived: 09/12/2007	
	(Type) SEPF			
Excraction.	(Type) SEFF	Date Exti	cacted: 09/14/2007	
Concentrated	Extract Volume: 5000 (uL)	Date Anal	yzed: <u>09/19/2007</u>	
Injection Vo	lume: 2.0 (uL) GPC Factor:	Dilut	ion Factor: 1.0	
	(Y/N) <u>N</u> pH:	•	eanup: (Y/N) N	
	: (Y/N) <u>Y</u>			
CAS NO.	COMPOUND		CONCENTRATION UNITS: (ug/L or ug/kg) ug/L	Q.
12674-11-2	Aroclor-1016		0.50	Ū
11104-28-2	Aroclor-1221		0.50	Ū
11141-16-5	Aroclor-1232		0.50	U
53469-21-9	Aroclor-1242		0.50	U
12672-29-6	Aroclor-1248		0.50	U
11097-69-1	Aroclor-1254	· .	0.50	Ŭ -
11096-82-5	Aroclor-1260		0.50	U
37324-23-5	Aroclor-1262		0.50	U
11100-14-4	Aroclor-1268		0.50	U.

EPA SAMPLE NO.

B4F89	

Lab Name: Da	taChem Laboratories, Inc.	Contract	: EP-W-05-026	
	TAC Case No.: 36799 Mod. Ref			
	L/SED/WATER) <u>WATER</u>		le ID: 7256022003	
Sample wt/vo	ol: 1000 (g/mL) <u>mL</u>		ID: 20070917A070,200709	17B070
% Moisture:	Decanted: (Y/N)		eived: <u>09/12/2007</u>	
Extraction:	(Type) SEPF			
	Extract Volume: 5000 (uL)			
	lume: 2.0 (uL) GPC Factor:			
	(Y/N) <u>N</u> pH:		leanup: (Y/N) N	
Acid Cleanup	: (Y/N) <u>Y</u>			
CAS NO.	COMPOUND		CONCENTRATION UNITS: (ug/L or ug/kg) ug/L	Q
7,	Aroclor-1016		0.50	ΤΪ
11104-28-2	Aroclor-1221		0.50	Ω.
11141-16-5	Aroclor-1232		0.50	IJ
3469-21-9	Aroclor-1242		0.50	Ū
2672-29-6	Aroclor-1248		0.50	U.
1097-69-1	Aroclor-1254		0.50	n n
1096-82-5	Aroclor-1260		0.50	U
7324-23-5	Aradlar 1262		0.30	<u> </u>

11100-14-4

Aroclor-1268

B4F90	
Darlo	

	•			
Lab Name: DataChem Laboratories,	Inc.	Contract:	EP-W-05-026	
Lab Code: DATAC Case No.: 30	5799 Mod. Re	f No.:	SDG No.: <u>B4F84</u>	
Matrix: (SOIL/SED/WATER) WATER		Lab Sampl	Le ID: 7256022004	
Sample wt/vol: 1000 (g/mL)	mL.	Lab File	ID: 20070917A071,200709	17B071
% Moisture: Decanted:	(Y/N)	Date Rece	eived: 09/12/2007	•
Extraction: (Type) SEPF		Date Extr	racted: 09/14/2007	
Concentrated Extract Volume: 5000	(uL)	Date Anal	yzed: 09/19/2007	
Injection Volume: 2.0 (uL) GPC	Factor:	Dilut:	ion Factor: 1.0	
GPC Cleanup: (Y/N) N pH:	Marie Committee		eanup: (Y/N) N	
Acid Cleanup: (Y/N) Y				
CAS NO. COMPOUND			CONCENTRATION UNITS: (ug/L or ug/kg) ug/L	Q
12674-11-2 Aroclor-1016			0.50	U
11104-28-2 Aroclor-1221			0.50	Ū
11141-16-5 Aroclor-1232			0.50	U
53469-21-9 Aroclor-1242			0.50	Ū
12672-29-6 Aroclor-1248			0.50	U
11097-69-1 Aroclor-1254			0.50	ט
11096-82-5 Aroclor-1260			0.50	U
37324-23-5 Aroclor-1262			0.50	U
11100-14-4 Aroclor-1268			0.50	Ū

EPA SAMPLE NO.

B4F93

,				. 174	F 33
Lab Name: Da	taChem Laboratories, Inc.	Contract	: EP-W-05-026		
Lab Code: DA	TAC Case No.: 36799 Mod. F	Ref No.:	SDG No.	: B4F84	
Matrix: (SO)	IL/SED/WATER) <u>WATER</u>		le ID: <u>7256022</u> (		
Sample wt/vo	ol: <u>1000</u> (g/mL) <u>mL</u>		ID: 20070917A0		917B072
% Moisture:	Decanted: (Y/N)	• •	eived: <u>09/12/2</u> 0		71.0012
Extraction:	(Type) SEPF		racted: <u>09/14/2</u>		
Concentrated	Extract Volume: 5000 (uL)				
	lume: 2.0 (uL) GPC Factor:				
GPC Cleanup:	(Y/N) <u>N</u> pH:	Sulfur Cl	eanup: (Y/N) N	1	
Acid Cleanup	: (Y/N) <u>Y</u>		· ·	-	
CAS NO.	COMPOUND		CONCENTRATION (ug/L or ug/k	UNITS: g) ug/L	Q
12674-11-2	Aroclor-1016			0.50	II.
11104-28-2	Aroclor-1221			0.50	U
11141-16-5	Aroclor-1232			0.50	U
3469-21-9	Aroclor-1242				· · · · · · · · · · · · · · · · · · ·
L2672-29-6	Aroclor-1248			0.50	Ü
	Aroclor-1254			0.50	U .
	Aroclor-1260			0.50	U U

37324-23-5

11100-14-4

Aroclor-1262

Aroclor-1268

0.50

B4F94	
D4F 94	

		La company de	
Lab Name: Da	ataChem Laboratories, Inc. Contract	: EP-W-05-026	
Lab Code: DA	ATAC Case No.: 36799	SDG No.: <u>B4</u> F84	
the state of the s		le ID: <u>7256022006</u>	
Sample wt/vo	7 4000	ID: 20070917A073,2007091	7B073
	Decanted: (Y/N) Date Rec		
		racted: 09/14/2007	
	Extract Volume: 5000 (uL) Date Ana		
	plume: 2.0 (uL) GPC Factor: Dilut		
•		leanup: (Y/N) N	
Acid Cleanup	): (Y/N) <u>Y</u>		
CAS NO.	COMPOUND	CONCENTRATION UNITS: (ug/L or ug/kg) ug/L	Q
12674-11-2	Aroclor-1016	0.50	U
11104-28-2	Aroclor-1221	0.50	Ū.
11141-16-5	Aroclor-1232	0.50	U
53469-21-9	Aroclor-1242	0.50	U
12672-29-6	Aroclor-1248	0.50	Ū
11097-69-1	Aroclor-1254	0.50	U
11096-82-5	Aroclor-1260	0.50	U
37324-23-5	Aroclor-1262	0.50	Ü
11100-14-4	Aroclor-1268	0.50	U

B4F97		

Lab Name: Da	taChem Laboratories, Inc.	Contract	: <u>EP-W-05-026</u>	
Lab Code: DA	TAC Case No.: 36799 Mod. Re	E No.:	SDG No.: B4F84	
A CANADA CONTRACTOR OF THE CON	IL/SED/WATER) WATER		ole ID: 7256022007	
Sample wt/vo	ol: 1000 (g/mL) mL		ID: 20070917A074,200709	17B074
% Moisture:	Decanted: (Y/N)		eived: 09/12/2007	
	(Type) SEPF	. •	racted: 09/14/2007	
Concentrated	Extract Volume: 5000 (uL)			
•	lume: 2.0 (uL) GPC Factor:			
	(Y/N) <u>N</u> pH:		leanup: (Y/N) N	<del></del>
	: (Y/N) <u>Y</u>			
CAS NO.	COMPOUND		CONCENTRATION UNITS: (ug/L or ug/kg) ug/L	Q
12674-11-2	Aroclor-1016		0.50	U
11104-28-2	Aroclor-1221	:	0.50	Ū
11141-16-5	Aroclor-1232		0.50	U
53469-21-9	Aroclor-1242		0.50	U
12672-29-6	Aroclor-1248		0.50	U
11097-69-1	Aroclor-1254		0.50	U
11096-82-5	Aroclor-1260		0.50	Ū
37324-23-5	Aroclor-1262		0.50	U
1100-14-4	Araclar-1268		0.50	

В	4F98	

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Lab Name: Da	taChem Laboratories, Inc.	Contract	: EP-W-05-026
Lab Code: DA	TAC Case No.: 36799 Mod. Re	f No.:	SDG No.: B4F84
	L/SED/WATER) WATER		le ID: <u>7256022008</u>
Sample wt/vo	1: 1000 (g/mL) <u>mL</u>		ID: 20070917A075,20070917B075
% Moisture:	Decanted: (Y/N)		eived: 09/12/2007
	(Type) SEPF	•	racted: 09/14/2007
Concentrated	Extract Volume: 5000 (uL)		yzed: 09/19/2007
Injection Vo	lume: 2.0 (uL) GPC Factor:		ion Factor: 1.0
GPC Cleanup:	(Y/N) <u>N</u> pH:		eanup: (Y/N) N
Acid Cleanup			
CAS NO.	COMPOUND		CONCENTRATION UNITS: (ug/L or ug/kg) ug/L Q
12674-11-2	Aroclor-1016		0.50 U
11104-28-2	Aroclor-1221		0.50 U
11141-16-5	Aroclor-1232		0.50 U
53469-21-9	Aroclor-1242		0.50 U
12672-29-6	Aroclor-1248		0.50 U
11097-69-1	Aroclor-1254		0.50 U
11096-82-5	Aroclor-1260		0.50 U
37324-23-5	Aroclor-1262		0.50 U
11100-14-4	Aroclor-1268		0.50 U

EPA SAMPLE NO.

B4FA1	

Lab Name: <u>Da</u>	taChem Laboratories, Inc. Cont	ract: <u>EP-W-05-026</u>
Lab Code: DA	TAC Case No.: 36799 Mod. Ref No.:	SDG No.: B4F84
	L/SED/WATER) WATER Lab	
	1: 1000 (g/mL) mL Lab I	
	Decanted: (Y/N) Date	
	(Type) <u>SEPF</u> Date	
2	Extract Volume: 5000 (uL) Date	
	lume: 2.0 (uL) GPC Factor: D	
		ur Cleanup: (Y/N) N
	: (Y/N) <u>Y</u>	
CAS NO.	COMPOUND	CONCENTRATION UNITS: (ug/L or ug/kg) ug/L Q
12674-11-2	Aroclor-1016	0.50 U
11104-28-2	Aroclor-1221	0.50 บ
11141-16-5	Aroclor-1232	0.50 Ū
53469-21-9	Aroclor-1242	0.50 U
12672-29-6	Aroclor-1248	0.50 U
11097-69-1	Aroclor-1254	0.50 U

11096-82-5

37324-23-5

11100-14-4

Aroclor-1260

Aroclor-1262

Aroclor-1268

0.50

0.50

0.50

U

U

EPA SAMPLE NO.

B4FA2	

Lab Name: Da	taChem Laboratories, Inc.	Contract: EP-W-05-026
Lab Code: DA	TAC Case No.: 36799 Mod. Re	f No.: SDG No.: B4F84
	L/SED/WATER) <u>WATER</u>	
		Lab File ID: 20070917A077,20070917B077
% Moisture:	and the second of the second o	Date Received: 09/12/2007
Extraction:	(Type) SEPF	Date Extracted: 09/14/2007
Concentrated	Extract Volume: 5000 (uL)	Date Analyzed: 09/19/2007
Injection Vo	lume: 2.0 (uL) GPC Factor:	Dilution Factor: 1.0
GPC Cleanup:	(Y/N) <u>N</u> pH:	Sulfur Cleanup: (Y/N) N
	: (Y/N) <u>Y</u>	
CAS NO.	COMPOUND	CONCENTRATION UNITS: (ug/L or ug/kg) ug/L Q
12674-11-2	Aroclor-1016	0.50 บ
11104-28-2	Aroclor-1221	0.50 U
11141-16-5	Aroclor-1232	0.50 U
53469-21-9	Aroclor-1242	0.50 U
12672-29-6	Aroclor-1248	0.50 U
11097-69-1	Aroclor-1254	0.50 U
11096-82-5	Aroclor-1260	0.50 U
37324-23-5	Aroclor-1262	0.50 U

11100-14-4 Aroclor-1268

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	B4FA	6	

Lab Name: DataChem Laboratories, Inc. Contract: EP-W-05-026			
Lab Code: DATAC Case No.: 36799 Mod.	Ref No.: SDG No.: <u>B4F84</u>		
Matrix: (SOIL/SED/WATER) WATER	Lab Sample ID: 7256022011		
Sample wt/vol: 1000 (g/mL) mL	Lab File ID: 20070917A078,20070917B078		
% Moisture: Decanted: (Y/N)			
Extraction: (Type) SEPF	Date Extracted: 09/14/2007		
Concentrated Extract Volume: 5000 (uL)			
Injection Volume: 2.0 (uL) GPC Factor:			
GPC Cleanup: (Y/N) N pH:			
Acid Cleanup: (Y/N) Y			
CAS NO. COMPOUND	CONCENTRATION UNITS: (ug/L or ug/kg) ug/L Q		
12674-11-2 Aroclor-1016	0.50 U		
11104-28-2 Aroclor-1221	0.50 U		
	0.50 U		
53469-21-9 Aroclor-1242	0.50 U		
12672-29-6 Aroclor-1248	0.50 U		
11097-69-1 Aroclor-1254	0.50 U		
11096-82-5 Aroclor-1260	0.50 U		
37324-23-5 Aroclor-1262	0.50 U		
11100-14-4 Aroclor-1268	0.50 U		

EPA SAMPLE NO.

0.50

0.50

0.50

0.50

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	B4F	17		

Lab Name: DataChem Laboratories, Inc.	Contract: EP-W-05-026
Lab Code: DATAC Case No.: 36799 Mod. Re	ef No.: SDG No.: <u>B4F84</u>
Matrix: (SOIL/SED/WATER) WATER	Lab Sample ID: 7256022012
Sample wt/vol: 1000 (g/mL) mL	
% Moisture: Decanted: (Y/N)	Date Received: <u>09/12/2007</u>
Extraction: (Type) SEPF	Date Extracted: 09/14/2007
Concentrated Extract Volume: 5000 (uL)	Date Analyzed: 09/19/2007
Injection Volume: 2.0 (uL) GPC Factor:	Dilution Factor: 1.0
GPC Cleanup: (Y/N) N pH:	Sulfur Cleanup: (Y/N) N
Acid Cleanup: (Y/N) Y	
CAS NO. COMPOUND	CONCENTRATION UNITS: (ug/L or ug/kg) ug/L Q
12674-11-2 Aroclor-1016	0.50 U
11104-28-2 Aroclor-1221	0.50 U
11141-16-5 Aroclor-1232	
	0.50 U
53469-21-9 Aroclor-1242	0.50 U 0.50 U

11097-69-1

11096-82-5

37324-23-5

11100-14-4

Aroclor-1254

Aroclor-1260

Aroclor-1262

Aroclor-1268

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B4F1	D 2	
1341.1	52	

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Lab Name: Da	taChem Laboratories, Inc.	Contract	: EP-W-05-026	
Lab Code: DA	TAC Case No.: 36799 Mod. Re	f No.:	SDG No.: <u>B4F84</u>	
	L/SED/WATER) WATER		le ID: <u>7256022015</u>	
Sample wt/vo	1: 1000 (g/mL) mL		ID: 20070917A082,200709	917B082
	Decanted: (Y/N)		eived: <u>09/12/2007</u>	
Extraction:	· · · · · · · · · · · · · · · · · · ·		racted: 09/14/2007	
	Extract Volume: 5000 (uL)			
		*	yzed: <u>09/19/2007</u>	
injection Vo	lume: 2.0 (uL) GPC Factor:	Dilut	ion Factor: 1.0	
GPC Cleanup:	(Y/N) <u>N</u> pH:	Sulfur Cl	eanup: (Y/N) N	
Acid Cleanup				•
CAS NO.			CONCENTRATION UNITS: (ug/L or ug/kg) ug/L	Q
12674-11-2	Aroclor-1016		0.50	Ü
11104-28-2	Aroclor-1221		0.50	Ū
11141-16-5	Aroclor-1232	A second	0.50	ט
53469-21-9	Aroclor-1242		0.50	U
12672-29-6	Aroclor-1248		0.50	U
11097-69-1	Aroclor-1254		0.50	Ū
11096-82-5			0.50	Ü
37324-23-5	Aroclor-1262		0.50	U
L1100-14-4	Aroclor-1268		0.50	ט

EPA SAMPLE NO.

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B4FB3	

Lab Name: DataChem Laboratories, Inc.	Contract: EP-W-05-026
Lab Code: DATAC Case No.: 36799 Moo	d. Ref No.: SDG No.: B4F84
Matrix: (SOIL/SED/WATER) WATER	Lab Sample ID: 7256022016
Sample wt/vol: 1000 (g/mL) mL	Lab File ID: 20070917A083,20070917B083
% Moisture: Decanted: (Y/N)	
Extraction: (Type) SEPF	Date Extracted: 09/14/2007
Concentrated Extract Volume: 5000 (	uL) Date Analyzed: 09/19/2007
Injection Volume: 2.0 (uL) GPC Factor:	Dilution Factor: 1.0
GPC Cleanup: (Y/N) N pH:	Sulfur Cleanup: (Y/N) N
Acid Cleanup: (Y/N) Y	
CAS NO. COMPOUND	CONCENTRATION UNITS: (ug/L or ug/kg) ug/L Q
12674-11-2 Aroclor-1016	0.50 U
11104-28-2 Aroclor-1221	0.50 U
11141-16-5 Aroclor-1232	0.50 U
53469-21-9 Aroclor-1242	0.50 U
12672-29-6 Aroclor-1248	0.50 U
11097-69-1 Aroclor-1254	0.50 U
11096-82-5 Aroclor-1260	0.50 U
37324-23-5 Aroclor-1262	0.50 U

11100-14-4

Aroclor-1268

EPA SAMPLE NO.

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B4FB6			

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Lab Name: Da	taChem Laboratories, Inc.	Contract	: EP-W-05-026	
	TAC Case No.: 36799 Mod. Ref			
·	L/SED/WATER) <u>WATER</u>			
	l: <u>1000</u> (g/mL) <u>mL</u>		-	28B053
•	Decanted: (Y/N)			
	(Type) SEPF			
	Extract Volume: 5000 (uL)			
	lume: 2.0 (uL) GPC Factor:			
			eanup: (Y/N) N	
Acid Cleanup:		•		
CAS NO.	COMPOUND		CONCENTRATION UNITS: (ug/L or ug/kg) ug/L	Q
12674-11-2	Aroclor-1016		0.50	U
11104-28-2	Aroclor-1221		0.50	n n
11141-16-5	Aroclor-1232		0.50	- []
53469-21-9	Aroclor-1242		0.50	U
12672-29-6	Aroclor-1248		0.50	U
11097-69-1	Aroclor-1254	-	0.50	U
11096-82-5	Aroclor-1260		0.50	<u>ט</u>
37324-23-5	Aroclor-1262	<del></del>	0.50	

11100-14-4

Aroclor-1268

B4FB7	

Lab Name: DataChem Laboratories, Inc.	Contract: EP-W-05-026
Lab Code: DATAC Case No.: 36799 Mod. Ref	E No.: SDG No.: <u>B4FB6</u>
Matrix: (SOIL/SED/WATER) WATER	Lab Sample ID: 7258003002
Sample wt/vol: 1000 (g/mL) mL	Lab File ID: 20070928A054,20070928B054
% Moisture: Decanted: (Y/N)	Date Received: 09/14/2007
Extraction: (Type) SEPF	Date Extracted: 09/16/2007
Concentrated Extract Volume: 5000 (uL)	Date Analyzed: 09/28/2007
Injection Volume: 2.0 (uL) GPC Factor:	Dilution Factor: 1.0
GPC Cleanup: (Y/N) N pH:	Sulfur Cleanup: (Y/N) N
Acid Cleanup: (Y/N) Y	
	CONCENTRATION UNITES

CAS NO.	COMPOUND	CONCENTRATION UNITS: (ug/L or ug/kg) ug/L	Q
12674-11-2	Aroclor-1016	0.50	Ū
11104-28-2	Aroclor-1221	0.50	Ū
11141-16-5	Aroclor-1232	0.50	ט
53469-21-9	Aroclor-1242	0.50	Ü
12672-29-6	Aroclor-1248	0.50	U
11097-69-1	Aroclor-1254	0.50	. U .
11096-82-5	Aroclor-1260	0.50	U
37324-23-5	Aroclor-1262	0.50	Ū
11100-14-4	Aroclor-1268	0.50	Ū

EPA SAMPLE NO.

DADOI	•	
B4FC1		

Lab Name: DataChem Laboratories, Inc.		Contract: EP-W-05-026
Lab Code: DAT	TAC Case No.: 36799 Mod. Ref	5 No.: SDG No.: <u>B4FB6</u>
Matrix: (SOI	L/SED/WATER) WATER	Lab Sample ID: 7258003005
Sample wt/vo	l: 1000 (g/mL) mL	Lab File ID: 20070928A057,20070928B057
% Moisture: _	Decanted: (Y/N)	Date Received: 09/14/2007
	(Type) SEPF	Date Extracted: 09/16/2007
Concentrated	Extract Volume: 5000 (uL)	Date Analyzed: 09/28/2007
Injection Vo	lume: 2.0 (uL) GPC Factor:	Dilution Factor: 1.0
GPC Cleanup:	(Y/N) <u>N</u> pH:	Sulfur Cleanup: (Y/N) N
Acid Cleanup	: (Y/N) <u>Y</u>	
CAS NO.	COMPOUND	CONCENTRATION UNITS: Q Q
12674-11-2	Aroclor-1016	0.50 U
11104-28-2	Aroclor-1221	0.50 U
11141-16-5	Aroclor-1232	0.50 U
53469-21-9	Aroclor-1242	0.50 U
12672-29-6	Aroclor-1248	
11097-69-1	Aroclor-1254	0.50 U
11096-82-5	Aroclor-1260	0.50 U
37324-23-5	Aroclor-1262	0.50 U

11100-14-4

Aroclor-1268

B4FC2	

Lab Name: Dat	taChem Laboratories, Inc.	Contract: EP-W-05-026
Lab Code: DAT	TAC Case No.: 36799 _ Mod. Ref	f No.: SDG No.: <u>B4FB6</u>
Matrix: (SOI	L/SED/WATER) <u>WATER</u>	Lab Sample ID: 7258003006
Sample wt/vo	l: 1000 (g/mL) mL	Lab File ID: 20070928A058,20070928B058
% Moisture: _	Decanted: (Y/N)	Date Received: 09/14/2007
Extraction: (Type) SEPF Date Extracted: 09/16/2007		
Concentrated Extract Volume: 5000 (uL) Date Analyzed: 09/28/2007		
Injection Volume: 2.0 (uL) GPC Factor: Dilution Factor: 1.0		
GPC Cleanup: (Y/N) N pH: Sulfur Cleanup: (Y/N) N		
Acid Cleanup	: (Y/N) <u>Y</u>	
CAS NO.	COMPOUND	CONCENTRATION UNITS: Q Q
12674-11-2	Aroclor-1016	0.50 U
11101-20-2	7~0~10~-1221	0 50 #

CAS NO.	COMPOUND	CONCENTRATION UNITS: (ug/L or ug/kg) ug/L	Q
12674-11-2	Aroclor-1016	0.50	Ü
11104-28-2	Aroclor-1221	0.50	Ū
11141-16-5	Aroclor-1232	0.50	U:
53469-21-9	Aroclor-1242	0.50	U
12672-29-6	Aroclor-1248	0.50	Ü
11097-69-1	Aroclor-1254	0.50	Ū
11096-82-5	Aroclor-1260	0.50	Ū
37324-23-5	Aroclor-1262	0.50	Ū
11100-14-4	Aroclor-1268	0.50	Ü

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	В	4 FC	6.	

Lab Name: DataChem Laboratories, Inc.		Contract: EP-W-05-026		
Lab Code: DAT	TAC Case No.: 36799 Mod.	Ref No.: SDG No.: <u>B4FB6</u>		
Matrix: (SOI	L/SED/WATER) WATER	Lab Sample ID: 7258003007		
Sample wt/vo.	l: 1000 (g/mL) mL	Lab File ID: 20070928A059,20070928B059		
% Moisture: _	Decanted: (Y/N)	Date Received: 09/14/2007		
Extraction:	(Type) SEPF	Date Extracted: 09/16/2007		
Concentrated	Concentrated Extract Volume: 5000 (uL) Date Analyzed: 09/28/2007			
Injection Vol	lume: 2.0 (uL) GPC Factor:	Dilution Factor: 1.0		
GPC Cleanup:	(Y/N) N pH:	Sulfur Cleanup: (Y/N) N		
	: (Y/N) <u>Y</u>			
CAS NO.	COMPOUND	CONCENTRATION UNITS: (ug/L or ug/kg) ug/L Q		
12674-11-2	Aroclor-1016	0.50 U		
11104-28-2	Aroclor-1221	0.50 U		

CAS NO.	COMPOUND	CONCENTRATION UNITS: (ug/L or ug/kg) ug/L	Q
12674-11-2	Aroclor-1016	0.50	Ū
11104-28-2	Aroclor-1221	0.50	Ū
11141-16-5	Aroclor-1232	0.50	Ū
53469-21-9	Aroclor-1242	0.50	U
12672-29-6	Aroclor-1248	0.50	Ū
11097-69-1	Aroclor-1254	0.50	Ū
11096-82-5	Aroclor-1260	0.50	Ū
37324-23-5	Aroclor-1262	0.50	Ü
11100-14-4	Aroclor-1268	0.50	Ū

EPA SAMPLE NO.

B4FC7	

Lab Name: Da	: EP-W-05-026	•		
	TAC Case No.: 36799 Mod. Ref			
•			le ID: 7258003008	
	1: 1000 (g/mL) mL		ID: 20070928A060,200709	928B060
÷"	Decanted: (Y/N)			
	(Type) SEPF		racted: 09/16/2007	
Concentrated	Extract Volume: 5000 (uL)		lyzed: 09/28/2007	
			ion Factor: 1.0	
	(Y/N) <u>N</u> pH:		leanup: (Y/N) N	
Acid Cleanup	: (Y/N) <u>Y</u>			
CAS NO.	COMPOUND		CONCENTRATION UNITS: (ug/L or ug/kg) ug/L	Q
12674-11-2	Aroclor-1016		0.50	Ū
11104-28-2	Aroclor-1221		0.50	Ü .
11141-16-5	Aroclor-1232		0.50	Ū
53469-21-9	Aroclor-1242			Ū
12672-29-6	Aroclor-1248		0.50	U
11097-69-1	Aroclor-1254		0.50	U
11096-82-5	Aroclor-1260		0.50	U
37324-23-5	Aroclor-1262		0.50	

11100-14-4

Aroclor-1268

	B4FJ8	
Lab Name: DataChem Laboratories, Inc.	Contract: EP-W-05-026	_
Lab Code: DATAC Case No.: 36799 Mod. Ref	f No.: SDG No.: <u>B4FJ1</u>	_
Matrix: (SOIL/SED/WATER) WATER	Lab Sample ID: <u>7306027008</u>	_
Sample wt/vol: 220 (g/mL) mL	Lab File ID: 20071107A020,20071107B020	_
% Moisture: Decanted: (Y/N)	Date Received: 11/01/2007	_
Extraction: (Type) SEPF	Date Extracted: 11/05/2007	
Concentrated Extract Volume: 10000 (uL)	Date Analyzed: 11/07/2007	_
Injection Volume: 2.0 (uL) GPC Factor:	Dilution Factor: 1.0	_
GPC Cleanup: (Y/N) N pH:	Sulfur Cleanup: (Y/N) N	_
Acid Cleanup: (Y/N) Y		
		_

CAS NO.	COMPOUND	. 1	i .	ONCENTRATION UNITS: ug/L or ug/kg) ug/L	Q
12674-11-2	Aroclor-1016		į	4.5	υJ
11104-28-2	Aroclor-1221		·	4.5	Ü
11141-16-5	Aroclor-1232			4.5	U
53469-21-9	Aroclor-1242			4.5	Ü
12672-29-6	Aroclor-1248			4.5	U
11097-69-1	Aroclor-1254			4.5	Ü
11096-82-5	Aroclor-1260			4.5	ט
37324-23-5	Aroclor-1262		•	4.5	U
11100-14-4	Aroclor-1268		Y	4.5	ū

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B4	FJ9	

Lab Name: DataChem Laboratories, Inc. C			EP-W-05-026
Lab Code: DAT	AC Case No.: 36799 Mod. Re	f No.:	SDG No.: B4FJ1
Matrix: (SOI)	L/SED/WATER) WATER	Lab Sampl	e ID: 7306027009
Sample wt/vol	L: 200 (g/mL) mL	Lab File	ID: 20071107A021,20071107B021
% Moisture: _	Decanted: (Y/N)	Date Rece	ived: 11/01/2007
*	(Type) SEPF	Date Extr	acted: 11/05/2007
	Extract Volume: 10000 (uL)	Date Anal	yzed: 11/07/2007
	Lume: 2.0 (uL) GPC Factor:		
	GPC Cleanup: (Y/N) N pH: Sulfur Cleanup: (Y/N) N		
	: (Y/N) <u>Y</u>		
CAS NO.	COMPOUND	.	CONCENTRATION UNITS: Q (ug/L or ug/kg) ug/L Q
12674-11-2	Aroclor-1016		5.0 U
11104-28-2	Aroclor-1221		5.0 U
11141-16-5	Aroclor-1232		5.0 U
53469-21-9	Aroclor-1242		5.0 U
12672-29-6	Aroclor-1248		5.0 U
11097-69-1	Aroclor-1254		5.0 U
11096-82-5	Aroclor-1260		5.0 U
37324-23-5	Aroclor-1262		5.0 U
11100-14-4	Aroclor-1268		5.0 U

EPA SAMPLE NO.

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B4FK	4

Lab Name: DataChem Laboratories, Inc. Contract: EP-W-05-026			
Lab Code: DATAC Case No.: 36799 Mod. Ref No.: SDG No.: B4FB6			
		ple ID: 7258003009	
Sample wt/vo	ol: 1000 (g/mL) mL Lab Fil	e ID: 20070928A061,20070928B061	
The second second		ceived: 09/14/2007	
Extraction:	(Type) SEPF Date Ex	tracted: 09/16/2007	
Concentrated	Extract Volume: 5000 (uL) Date An	alyzed: 09/29/2007	
Injection Vo	lume: 2.0 (uL) GPC Factor: Dilu	ition Factor: 1.0	
		Cleanup: (Y/N) N	
Acid Cleanup			
CAS NO.	COMPOUND	CONCENTRATION UNITS: (ug/L or ug/kg) ug/L Q	
12674-11-2	Aroclor-1016	0.50 U	
11104-28-2	Aroclor-1221	0.50 U	
11141-16-5	Aroclor-1232	0.50 U	
53469-21-9	Aroclor-1242	0.50 U	
12672-29-6	Aroclor-1248	0.50 U	
11097-69-1	Aroclor-1254	0.50 U	
11096-82-5	Aroclor-1260	0.50 U	
37324-23-5	Aroclor-1262	0.50 U	
11100-14-4	Aroclor-1268	0.50 п	

EPA SAMPLE NO.

B4FK5	

Lab Name: <u>Da</u>	taChem Laboratories, Inc.	Co	ontract	: EP-W-05-026	
Lab Code: DATAC Case No.: 36799 Mod. Ref No.: SDG No.: B4FB6					
Matrix: (SOI	IL/SED/WATER) <u>WATER</u>	Lá	ab Sampl	le ID: 7258003010	
Sample wt/vo	ol: 1000 (g/mL) mL	· . La	ab File	ID: 20070928A062,200709	28B062
% Moisture:	Decanted: (Y/N)	··· Da	ate Rece	eived: 09/14/2007	
Extraction:	(Type) SEPF	Da	ite Extr	racted: 09/16/2007	
Concentrated	Extract Volume: 5000	(uL) Da	ite Anal	Lyzed: <u>09/29/2007</u>	
Injection Vo	lume: 2.0 (uL) GPC Factor: _		Dilut	ion Factor: 1.0	
GPC Cleanup:	(Y/N) <u>N</u> pH:	Su	ılfur Cl	Leanup: (Y/N) N	
Acid Cleanup	: (Y/N) <u>Y</u>				
	COMPOUND			CONCENTRATION UNITS: (ug/L or ug/kg) ug/L	Q
12674-11-2	Aroclor-1016			0.50	U
11104-28-2	Aroclor-1221			0.50	Ū
11141-16-5	Aroclor-1232			0.50	Ū
53469-21-9	Aroclor-1242			0.50	Ū
12672-29-6	Aroclor-1248	1. 1.		0.50	Ü
11097-69-1	Aroclor-1254			0.50	Ü
11096-82-5	Aroclor-1260			0.50	Ü
37324-23-5	Aroclor-1262			0.50	U
11100-14-4	Aroclor-1268			0.50	П

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	B4FK6	
	DALKO	

Lab Name: Da	taChem Laboratories, Inc.	Contract: EP-W-05-026		
	TAC Case No.: 36799 Mod. Ref			
		Lab Sample ID: 7258003011		
Sample wt/vo	1: <u>1000</u> (g/mL) <u>mL</u>	Lab File ID: 20070928A063,20070928B063		
* .	Decanted: (Y/N)			
Extraction:	(Type) SEPF	Date Extracted: 09/16/2007		
Concentrated	Extract Volume: 5000 (uL)	Date Analyzed: 09/29/2007		
Injection Vo	lume: 2.0 (uL) GPC Factor:	Dilution Factor: 1.0		
GPC Cleanup:	(Y/N) <u>N</u> pH:	Sulfur Cleanup: (Y/N) N		
Acid Cleanup	: (Y/N) <u>Y</u> -			
CAS NO.	COMPOUND	CONCENTRATION UNITS: (ug/L or ug/kg) ug/L /Q		
12674-11-2	Aroclor-1016	0.50 U		
11104-28-2	Aroclor-1221	0.50 17		

CAS NO.	COMPOUND		CONCENTRATION UNITS: (ug/L or ug/kg) ug/L	√Q
12674-11-2	Aroclor-1016		0.50	ט
11104-28-2	Aroclor-1221		0.50	ט
11141-16-5	Aroclor-1232	<del></del>	0.50	Ū
53469-21-9	Aroclor-1242		0.50	U
12672-29-6	Aroclor-1248		0.50	U
11097-69-1	Aroclor-1254		0.50	Ū
11096-82-5	Aroclor-1260		0.50	Ū
37324-23-5	Aroclor-1262		0.50	Ū
11100-14-4	Aroclor-1268	-	0.50	U

EPA SAMPLE NO.

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B4FK7	

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Lab Name: Da	taChem Laboratories, Inc.	Contract	: EP-W-05-026	-
Lab Code: DA	TAC Case No.: 36799 Mod. Ref	No.:	SDG No.: B4FB6	
Matrix: (SO	IL/SED/WATER) WATER	Lab Samp	le ID: <u>7258003012</u>	
Sample wt/vo	ol: 1000 (g/mL) <u>mL</u>	Lab File	ID: 20070928A064,20070928	3B064
% Moisture:	Decanted: (Y/N)	Date Rece	eived: 09/14/2007	
Extraction:	(Type) SEPF	Date Ext	racted: <u>09/16/2007</u>	
Concentrated	Extract Volume: 5000 (uL)	Date Anal	Lyzed: <u>09/29/2007</u>	
	lume: 2.0 (uL) GPC Factor:			
GPC Cleanup:	(Y/N) N pH:	Sulfur C	Leanup: (Y/N) N	
Acid Cleanup	: (Y/N) <u>Y</u>			
CAS NO.	COMPOUND		CONCENTRATION UNITS: (ug/L or ug/kg) ug/L	Q
12674-11-2	Aroclor-1016		0.50	U
11104-28-2	Aroclor-1221		0.50	Ū
11141-16-5	Aroclor-1232		0.50	U
53469-21-9	Aroclor-1242		0.50	Ū
12672-29-6	Aroclor-1248		0.50	U
11097-69-1	Aroclor-1254		0.50	U
11096-82-5	Aroclor-1260		0.50	U
37324-23-5	Aroclor-1262			U
11100-14-4	Aroclor-1268		0.50	11

EPA SAMPLE NO.

В	4 FK8	•	,

Lab Name: DataChem Laboratories, Inc. Contract: EP-W-05-026			
Lab Code: DAT	TAC Case No.: 36799 Mod. Ref	No.: SDG No.: <u>B4FB6</u>	
Matrix: (SOI	L/SED/WATER) WATER	Lab Sample ID: 7258003013	
Sample wt/vo	l: <u>1000                                 </u>	Lab File ID: 20070928A068,20070928B068	
% Moisture:	Decanted: (Y/N)	Date Received: 09/14/2007	
Extraction:	(Type) SEPF	Date Extracted: 09/16/2007	
Concentrated	Extract Volume: 5000 (uL)	Date Analyzed: 09/29/2007	
		Dilution Factor: 1.0	
GPC Cleanup:	(Y/N) N pH:	Sulfur Cleanup: (Y/N) N	
	: (Y/N) <u>Y</u>		
CAS NO.	COMPOUND	CONCENTRATION UNITS: (ug/L or ug/kg) ug/L Q	
12674-11-2	Aroclor-1016	0.50 U	
11104-28-2	Aroclor-1221	0.50 U	
11141-16-5	Aroclor-1232	0.50 U	
53469-21-9	Aroclor-1242	0.50 U	
12672-29-6	Aroclor-1248	0.50 U	
11097-69-1	Aroclor-1254	7	
11096-82-5	Aroclor-1260	0.50 U	
37324-23-5	Aroclor-1262	0.50 U	

11100-14-4

Aroclor-1268

EPA SAMPLE NO.

B4FK9	
DALKO	

Lab Name: DataChem Laboratories, Inc. Contract: EP-W-05-026			
Lab Code: DA	TAC Case No.: 36799 Mod. Ref	No.:	SDG No.: B4FB6
	L/SED/WATER) WATER		le ID: 7258003014
Sample wt/vo	ol: 1000 (g/mL) mL		ID: 20070928A069,20070928B069
% Moisture:	Decanted: (Y/N)	•	eived: <u>09/14/2007</u>
Extraction:	(Type) SEPF		racted: 09/16/2007
Concentrated	Extract Volume: 5000 (uL)		
	lume: 2.0 (uL) GPC Factor:		
	GPC Cleanup: (Y/N) N pH: Sulfur Cleanup: (Y/N) N		
Acid Cleanup	: (Y/N) <u>Y</u>	. :	
CAS NO.	COMPOUND		CONCENTRATION UNITS: (ug/L or ug/kg) ug/L Q
12674-11-2	Aroclor-1016		0.50 U
11104-28-2	Aroclor-1221		0.50 U
11141-16-5	Aroclor-1232		0.50 U
53469-21-9	Aroclor-1242		0.50 U
12672-29-6	Aroclor-1248		0.50 U
11097-69-1	Aroclor-1254		0.50 U
11096-82-5	Aroclor-1260		0.50 U
37324-23-5	Aroclor-1262		0.50 U
11100-14-4	Aroclor-1268		0.50

EPA SAMPLE NO.

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	B4FL0	

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Lab Name: Da	taChem Laboratories, Inc.	Contract: EP-W-05-026
	TAC Case No.: 36799 Mod. Ref	
	L/SED/WATER) WATER	Lab Sample ID: 7258003015
Sample wt/vo	ol: 1000 (g/mL) mL	Lab File ID: 20070928A070,20070928B070
% Moisture:	Decanted: (Y/N)	Date Received: 09/14/2007
Extraction:	(Type) SEPF	Date Extracted: 09/16/2007
Concentrated	Extract Volume: 5000 (uL)	
	lume: 2.0 (uL) GPC Factor:	
	(Y/N) <u>N</u> pH:	
	: (Y/N) <u>Y</u>	
CAS NO.	COMPOUND	CONCENTRATION UNITS: (ug/L or ug/kg) ug/L Q
12674-11-2	Aroclor-1016	0.50 U
11104-28-2	Aroclor-1221	0.50 U
11141-16-5	Aroclor-1232	0.50 U
53469-21-9	Aroclor-1242	0.50 U
12672-29-6	Aroclor-1248	0.50 U
11097-69-1	Aroclor-1254	0.50 U
11096-82-5	Aroclor-1260	0.50 U
37324-23-5	Aroclor-1262	0.50 U
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11100-14-4

Aroclor-1268

B4FL1	

Lab Name: DataChem Laboratories, Inc.	Contract: EP-W-05-026
Lab Code: DATAC Case No.: 36799 Mod. Ref	No.: SDG No.: <u>B4FB6</u>
Matrix: (SOIL/SED/WATER) WATER	Lab Sample ID: 7258003016
Sample wt/vol: 1000 (g/mL) mL	Lab File ID: 20070928A071,20070928B071
% Moisture: Decanted: (Y/N)	Date Received: 09/14/2007
Extraction: (Type) SEPF	Date Extracted: 09/16/2007
Concentrated Extract Volume: 5000 (uL)	Date Analyzed: 09/29/2007
Injection Volume: 2.0 (uL) GPC Factor:	Dilution Factor: 1.0
GPC Cleanup: (Y/N) N pH:	Sulfur Cleanup: (Y/N) N
Acid Cleanup: (Y/N) Y	

CAS NO.	COMPOUND	CONCENTRATION UNITS: (ug/L or ug/kg) ug/L	Q
12674-11-2	Aroclor-1016	0.50	Ū
11104-28-2	Aroclor-1221	0.50	U
11141-16-5	Aroclor-1232	0.50	IJ
53469-21-9	Aroclor-1242	0.50	П
12672-29-6	Aroclor-1248	0.50	Ü.
11097-69-1	Aroclor-1254	0.50	
11096-82-5	Aroclor-1260	0.50	IJ
37324-23-5	Aroclor-1262	0.50	U
11100-14-4	Aroclor-1268	0.50	

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B4FA0	

Lab Name: DataChem Laboratories, Inc.	Contract: EP-W-05-026
Lab Code: DATAC Case No.: 36799 Mod. Ref	No.: SDG No.: B4F82
Matrix: (SOIL/SED/WATER) SOIL	Lab Sample ID: 7256028011
Sample wt/vol: 30.0 (g/mL) g	Lab File ID: 20070928A085,20070928B085
% Moisture: 16 Decanted: (Y/N) N	Date Received: 09/12/2007
Extraction: (Type) SONC	Date Extracted: 09/16/2007
Concentrated Extract Volume: 5000 (uL)	Date Analyzed: 09/29/2007
Injection Volume: 2.0 (uL) GPC Factor: 2.0	Dilution Factor: 1.0
GPC Cleanup: (Y/N) Y pH: 7.2	Sulfur Cleanup: (Y/N) N
Acid Cleanup: (Y/N) Y	

CAS NO.	COMPOUND	CONCENTRATION UNITS: (ug/L or ug/kg) ug/kg	Q
12674-11-2	Aroclor-1016	20	U
11104-28-2	Aroclor-1221	20	Ū
11141-16-5	Aroclor-1232	20	U
53469-21-9	Aroclor-1242	20	TI
12672-29-6	Aroclor-1248	20	Ū
11097-69-1	Aroclor-1254	20	Ū
11096-82-5	Aroclor-1260	20	11
37324-23-5	Aroclor-1262	20	
11100-14-4	Aroclor-1268	20	

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	B4	FA3	
	B4	FA3	

Lab Name: DataChem Laboratories, Inc.	Contract: EP-W-05-026
Lab Code: DATAC Case No.: 36799 Mod. Ref	No.: SDG No.: B4F82
Matrix: (SOIL/SED/WATER) SOIL	Lab Sample ID: <u>7256028014</u>
Sample wt/vol: $30.0$ (g/mL) g	Lab File ID: 20070928A091,20070928B091
% Moisture: 20 / Decanted: (Y/N) N	Date Received: 09/12/2007
Extraction: (Type) SONC	Date Extracted: 09/16/2007
Concentrated Extract Volume: 5000 (uL)	Date Analyzed: 09/29/2007
Injection Volume: 2.0 (uL) GPC Factor: 2.0	Dilution Factor: 1.0
GPC Cleanup: (Y/N) Y pH: 7.4	Sulfur Cleanup: (Y/N) N
Acid Cleanup: (Y/N) Y	

CAS NO.	COMPOUND	CONCENTRATION UNITS: (ug/L or ug/kg) ug/kg	Q
12674-11-2	Aroclor-1016	21	IJ
11104-28-2	Aroclor-1221	21	Ü
11141-16-5	Aroclor-1232	21	TI.
53469-21-9	Aroclor-1242	21	TT.
12672-29-6	Aroclor-1248	21	Ū
11097-69-1	Aroclor-1254	21	TI U
11096-82-5	Aroclor-1260		
37324-23-5	Aroclor-1262	21	U
11100-14-4	Aroclor-1268	21	U U

B4FA4	

Lab Name: DataChem Laboratories, Inc.	Contract: EP-W-05-026
Lab Code: DATAC Case No.: 36799 Mod. Ret	f No.: SDG No.: B4F82
Matrix: (SOIL/SED/WATER) SOIL	Lab Sample ID: 7256028015
Sample wt/vol: 30.0 (g/mL) g	Lab File ID: 20070928A092,20070928B092
% Moisture: 15 Decanted: (Y/N) N	Date Received: 09/12/2007
Extraction: (Type) SONC	Date Extracted: 09/16/2007
Concentrated Extract Volume: 5000 (uL)	Date Analyzed: 09/29/2007
Injection Volume: 2.0 (uL) GPC Factor: 2.0	Dilution Factor: 1.0
GPC Cleanup: (Y/N) Y pH: 7.1	Sulfur Cleanup: (Y/N) N
Acid Cleanup: (Y/N) Y	
Acid Cleanup: (Y/N) Y	

CAS NO.	COMPOUND	CONCENTRATION UNITS: (ug/L or ug/kg) ug/kg	Q
12674-11-2	Aroclor-1016	19	Ü
11104-28-2	Aroclor-1221	19	Ü
11141-16-5	Aroclor-1232	19	Ū
53469-21-9	Aroclor-1242	19	Ū
12672-29-6	Aroclor-1248	19	Ū
11097-69-1	Aroclor-1254	19	Ū
11096-82-5	Aroclor-1260	19	ט
37324-23-5	Aroclor-1262	19	Ū
11100-14-4	Aroclor-1268	19	U

B4FA5	•	

Lab Name: DataChem Laboratories, Inc.	Contract: EP-W-05-026
Lab Code: DATAC Case No.: 36799 Mod. Re	f No.: SDG No.: B4F82
Matrix: (SOIL/SED/WATER) SOIL	Lab Sample ID: 7256028016
Sample wt/vol: $30.0/$ (g/mL) g	Lab File ID: 20070928A093,20070928B093
% Moisture: 16 / Decanted: (Y/N) N	Date Received: 09/12/2007
Extraction: (Type) SONC	Date Extracted: 09/16/2007
Concentrated Extract Volume: 5000 (uL)	Date Analyzed: 09/29/2007
Injection Volume: 2.0 (uL) GPC Factor: 2.0	Dilution Factor: 1.0
GPC Cleanup: (Y/N) Y pH: 6.3	Sulfur Cleanup: (Y/N) N
Acid Cleanup: (Y/N) Y	

CAS NO.	COMPOUND	CONCENTRATION UNIT		Q
12674-11-2	Aroclor-1016		20	U
11104-28-2	Aroclor-1221		20	Ū
11141-16-5	Aroclor-1232		20	[]
53469-21-9	Aroclor-1242		2.0	U
12672-29-6	Aroclor-1248		20	
11097-69-1	Aroclor-1254		20	
11096-82-5	Aroclor-1260		20	ū
37324-23-5	Aroclor-1262		20	Ü
11100-14-4	Aroclor-1268		20	Ū

B4FB0	

Lab Name: DataChem Laboratories, Inc.	Contract: EP-W-05-026
Lab Code: DATAC Case No.: 36799 Mod. Ref	No.: SDG No.: <u>B4F82</u>
Matrix: (SOIL/SED/WATER) SOIL	Lab Sample ID: 7256028017
Sample wt/vol: 30.0 (g/mL) g	Lab File ID: 20070928A094,20070928B094
% Moisture: 21 / Decanted: (Y/N) N	Date Received: 09/12/2007
Extraction: (Type) SONC	Date Extracted: 09/16/2007
Concentrated Extract Volume: 5000 (uL)	Date Analyzed: 09/29/2007
Injection Volume: 2.0 (uL) GPC Factor: 2.0	Dilution Factor: 1.0
GPC Cleanup: (Y/N) Y pH: 7.4	Sulfur Cleanup: (Y/N) N
Acid Cleanup: (Y/N) Y	

CAS NO.	COMPOUND	CONCENTRATION UNITS: (ug/L or ug/kg) ug/kg	Q
12674-11-2	Aroclor-1016	21	Ū
11104-28-2	Aroclor-1221	21	U
11141-16-5	Aroclor-1232	21	U
53469-21-9	Aroclor-1242	21	Ū
12672-29-6	Aroclor-1248	21	Ü .
11097-69-1	Aroclor-1254	21	ט
11096-82-5	Aroclor-1260	21	Ū
37324-23-5	Aroclor-1262	21	Ū
11100-14-4	Aroclor-1268	21	U

I	34FB1	

Lab Name: DataChem Laboratories, Inc.	Contract: EP-W-05-026
Lab Code: DATAC Case No.: 36799 Mod. Ref	No.: SDG No.: B4F82
Matrix: (SOIL/SED/WATER) SOIL	Lab Sample ID: 7256028018
Sample wt/vol: $30.0$ (g/mL) g	Lab File ID: 20070928A095,20070928B095
% Moisture: 15 Decanted: (Y/N) N	Date Received: 09/12/2007
Extraction: (Type) SONC	Date Extracted: 09/16/2007
Concentrated Extract Volume: 5000 (uL)	Date Analyzed: 09/29/2007
Injection Volume: 2.0 (uL) GPC Factor: 2.0	Dilution Factor: 1.0
GPC Cleanup: (Y/N) Y pH: 7.0	Sulfur Cleanup: (Y/N) N
Acid Cleanup: (Y/N) Y	

CAS NO.	COMPOUND	CONCENTRATION UNITS: (ug/L or ug/kg) ug/kg	Q
12674-11-2	Aroclor-1016	19	Ü
11104-28-2	Aroclor-1221	19	U
11141-16-5	Aroclor-1232	1.9	Ū
53469-21-9	Aroclor-1242	19	Ū
12672-29-6	Aroclor-1248	19	Ū
11097-69-1	Aroclor-1254	19	<b>ט</b>
11096-82-5	Aroclor-1260	19	ט
37324-23-5	Aroclor-1262	19	Ū
11100-14-4	Aroclor-1268	19	υ

B4FB4	

Lab Name: Dat	taChem Laboratories, Inc.	Contract: E	EP-W-05-026
Lab Code: DAT	TAC Case No.: 36799 Mod. Ref	No.:	SDG No.: <u>B4F82</u>
Matrix: (SOI	L/SED/WATER) SOIL	Lab Sample	ID: 7256028019
Sample wt/vo	1: 30.0 (g/mL) g	Lab File II	D: 20070928A096,20070928B096
% Moisture: 4	1.1 / Decanted: (Y/N) N	Date Receiv	ved: 09/12/2007
Extraction:	(Type) SONC	Date Extrac	cted: <u>09/16/2007</u>
Concentrated	Extract Volume: 5000 (uL)	Date Analy:	zed: 09/29/2007
Injection Vol	lume: 2.0 (uL) GPC Factor: 2.0	Dilutio	n Factor: 1.0
GPC Cleanup:	(Y/N) Y pH: 5.2	Sulfur Clea	anup: (Y/N)'N
Acid Cleanup:	: (Y/N) <u>Y</u>		
CAS NO.	COMPOUND		ONCENTRATION UNITS: ug/L or ug/kg) ug/kg Q
12674-11-2	Aroclor-1016		17 ט
77704 00 0	7 1001		

CAS NO.	COMPOUND		CONCENTRATION UNITS: Q Q
12674-11-2	Aroclor-1016	***	17 0: U
11104-28-2	Aroclor-1221		17 · · · · · · · · · · · · · · · · · · ·
11141-16-5	Aroclor-1232		ט 17
53469-21-9	Aroclor-1242		17 U
12672-29-6	Aroclor-1248		ט 17
11097-69-1	Aroclor-1254		17 U
11096-82-5	Aroclor-1260		17 U
37324-23-5	Aroclor-1262		ָט עָל 17
11100-14-4	Aroclor-1268		17 U

B4	FB8	

Lab Name: DataChem Laboratories, Inc.	Contract: EP-W-05-026
Lab Code: DATAC Case No.: 36799 Mod. Ref	No.: SDG No.: B4FB5
Matrix: (SOIL/SED/WATER) SOIL	Lab Sample ID: 7258004002
Sample wt/vol: $30.0$ (g/mL) g	Lab File ID: 20070928A028,20070928B028
% Moisture: 87 Decanted: (Y/N) N	Date Received: 09/14/2007
Extraction: (Type) SONC	Date Extracted: 09/16/2007
Concentrated Extract Volume: 5000 (uL)	Date Analyzed: 09/28/2007
Injection Volume: 2.0 (uL) GPC Factor: 2.0	Dilution Factor: 1.0
GPC Cleanup: (Y/N) Y pH: 6.9	Sulfur Cleanup: (Y/N) N
Acid Cleanup: (Y/N) Y	

CAS NO.	COMPOUND	CONCENTRATION UNITS: (ug/L or ug/kg) ug/kg	Q
12674-11-2	Aroclor-1016	130	U
11104-28-2	Aroclor-1221	130	U
11141-16-5	Aroclor-1232	130	Ū
53469-21-9	Aroclor-1242	130	U
12672-29-6	Aroclor-1248	130	U
11097-69-1	Aroclor-1254	130	U
11096-82-5	Aroclor-1260	130	U
37324-23-5	Aroclor-1262	130	U <b>1</b> /
11100-14-4	Aroclor-1268	130	U • 1

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В4	FB9	

Lab Name: DataChem Laboratories, Inc.	Contract: EP-W-05-026
Lab Code: DATAC Case No.: 36799 Mod. Ref	No.: SDG No.: B4FB5
Matrix: (SOIL/SED/WATER) SOIL	Lab Sample ID: 7269022001
Sample wt/vol: 30.0 (g/mL) g	Lab File ID: 20070928A100,20070928B100
% Moisture: 20 Decanted: (Y/N) N	Date Received: 09/26/2007
Extraction: (Type) SONC	Date Extracted: 09/27/2007
Concentrated Extract Volume: 5000 (uL)	Date Analyzed: 09/29/2007
Injection Volume: 2.0 (uL) GPC Factor: 2.0	Dilution Factor: 1.0
GPC Cleanup: (Y/N) Y pH: 7.0	Sulfur Cleanup: (Y/N) N
Acid Cleanup: (Y/N) Y	

CAS NO.	COMPOUND	CONCENTRATION UNITS: (ug/L or ug/kg) ug/kg	Q
12674-11-2	Aroclor-1016	21	IJ
11104-28-2	Aroclor-1221	21	U U
11141-16-5	Aroclor-1232	21	U
53469-21-9	Aroclor-1242	21	. U
12672-29-6	Aroclor-1248	21	
11097-69-1	Aroclor-1254	21	<u>U</u>
11096-82-5	Aroclor-1260	21	· U
37324-23-5	Aroclor-1262	21	11
11100-14-4	Aroclor-1268	21	Ū

B4FC	0

Lab Name: DataChem Laboratories, Inc.	Contract: EP-W-05-026
Lab Code: DATAC Case No.: 36799 Mod. Ref	No.: SDG No.: B4FB5
Matrix: (SOIL/SED/WATER) SOIL	Lab Sample ID: 7258004003
Sample wt/vol: $30.0$ (g/mL) g	Lab File ID: 20070928A029,20070928B029
% Moisture: 78 Decanted: (Y/N) N	Date Received: 09/14/2007
Extraction: (Type) SONC	Date Extracted: 09/16/2007
Concentrated Extract Volume: 5000 (uL)	Date Analyzed: 09/28/2007
Injection Volume: 2.0 (uL) GPC Factor: 2.0	Dilution Factor: 1.0
GPC Cleanup: (Y/N) Y pH: 6.5	Sulfur Cleanup: (Y/N) N
Acid Cleanup: (Y/N) Y	
	CONCENTRATION INTEG

CAS NO.	COMPOUND		CONCENTRATION UNITS: (ug/L or ug/kg) ug/kg	Q
12674-11-2	Aroclor-1016		76	U T
11104-28-2	Aroclor-1221		76	UA
11141-16-5	Aroclor-1232	·	76	· U
53469-21-9	Aroclor-1242		76	U
12672-29-6	Aroclor-1248		76	II
11097-69-1	Aroclor-1254		76	U
11096-82-5	Aroclor-1260		76	TT .
37324-23-5	Aroclor-1262		76	11 A
11100-14-4	Aroclor-1268		76	17 1

EPA SAMPLE NO.

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Lab Name: Da	taChem Laboratories, Inc.	Contract: EP-W	V-05-026
Lab Code: DA'	TAC Case No.: 36799 Mod. Ref	No.:	SDG No.: B4FB5
Matrix: (SOI	L/SED/WATER) <u>SOIL</u>	Lab Sample ID:	: 7258004004
Sample wt/vo	1: <u>30.0</u> (g/mL) g	Lab File ID: 2	20070928A030,20070928B030
% Moisture:	Decanted: (Y/N) N	Date Received:	: 09/14/2007
Extraction:	(Type) SONC	Date Extracted	d: <u>09/16/2007</u>
Concentrated	Extract Volume: 5000 (uL)	Date Analyzed:	: 09/28/2007
Injection Vo.	lume: 2.0 (uL) GPC Factor: 2.0	Dilution F	actor: 1.0
GPC Cleanup:	(Y/N) Y pH: 5.9	Sulfur Cleanur	o: (Y/N) N
Acid Cleanup	: (Y/N) <u>Y</u>		
CAS NO.	COMPOUND		ENTRATION UNITS: L or ug/kg) ug/kg  Q
12674-11-2	Aroclor-1016		17 ט
11104-28-2	Aroclor-1221		ט 17
11141-16-5	Aroclor-1232		17 U

53469-21-9

12672-29-6

11097-69-1

11096-82-5

37324-23-5

11100-14-4

Aroclor-1242

Aroclor-1248

Aroclor-1254

Aroclor-1260

Aroclor-1262

Aroclor-1268

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Lab Name: DataChem Laboratories, Inc.	Contract: EP-W-05-026
Lab Code: DATAC Case No.: 36799 Mod. Ref	No.: SDG No.: B4FB5
Matrice (COTT (CDD (TTD DD))	Lab Sample ID: 7258004005
Sample wt/vol: 30.0 (g/mL) g	Lab File ID: 20070928A031,20070928B031
% Moisture: 16 Decanted: (Y/N) N	Date Received: 09/14/2007
Part mant de la company de la	Date Extracted: 09/16/2007
Concentrated Extract Volume: 5000 (uL)	Date Analyzed: 09/28/2007
Injection Volume: 2.0 (uL) GPC Factor: 2.0	Dilution Factor: 1.0
CPC Classical (Trians)	Sulfur Cleanup: (Y/N) N
Acid Cleanup: (Y/N) Y	

CAS NO.	COMPOUND	CONCENTRATION UNITS: (ug/L or ug/kg) ug/kg Q
12674-11-2	Aroclor-1016	
11104-28-2	Aroclor-1221	20 U
11141-16-5	Aroclor-1232	20 U
53469-21-9	Aroclor-1242	20 U
12672-29-6	Aroclor-1248	20 0
11097-69-1	Aroclor-1254	20 U
11096-82-5	Aroclor-1260	20 U
37324-23-5	Aroclor-1262	
11100-14-4	<u> </u>	20 U
1110-14-4	Aroclor-1268	20 U

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Lab Name: DataChem Laboratories, Inc.	Contract: EP-W-05-026
Lab Code: DATAC Case No.: 36799 Mod. Ref	No.: SDG No.: <u>B4FB5</u>
Matrix: (SOIL/SED/WATER) SOIL	Lab Sample ID: 7258004006
Sample wt/vol: 30.0 (g/mL) g	Lab File ID: 20070928A032,20070928B032
% Moisture: 24 Decanted: (Y/N) N	Date Received: 09/14/2007
Extraction: (Type) SONC	Date Extracted: 09/16/2007
Concentrated Extract Volume: 5000 (uL)	Date Analyzed: 09/28/2007
Injection Volume: 2.0 (uL) GPC Factor: 2.0	Dilution Factor: 1.0
GPC Cleanup: (Y/N) Y pH: 7.2	Sulfur Cleanup: (Y/N) N
Acid Cleanup: (Y/N) Y	

CAS NO.	COMPOUND	CONCENTRATION UNITS: (ug/L or ug/kg) ug/kg	Q
12674-11-2	Aroclor-1016	22	Ü
11104-28-2	Aroclor-1221	22	Ū ···
11141-16-5	Aroclor-1232	22	. []
53469-21-9	Aroclor-1242	22	Ü
12672-29-6	Aroclor-1248	22	U
11097-69-1	Aroclor-1254	22	U U
11096-82-5	Aroclor-1260	22	Ū.
37324-23-5	Aroclor-1262	22	U.
11100-14-4	Aroclor-1268	22	Ū

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Lab Name: DataChem Laboratories, Inc.	Contract: EP-W-05-026
Lab Code: DATAC Case No.: 36799 Mod. Ret	f No.: SDG No.: B4FB5
Matrix: (SOIL/SED/WATER) SOIL	Lab Sample ID: 7258004007
Sample wt/vol: 30.0 (g/mL) g	Lab File ID: 20070928A033,20070928B033
% Moisture: 3.7 Decanted: (Y/N) N	Date Received: 09/14/2007
Extraction: (Type) SONC	Date Extracted: 09/16/2007
Concentrated Extract Volume: 5000 (uL)	Date Analyzed: 09/28/2007
Injection Volume: 2.0 (uL) GPC Factor: 2.0	Dilution Factor: 1.0
GPC Cleanup: (Y/N) Y pH: 5.9	Sulfur Cleanup: (Y/N) N
Acid Cleanup: (Y/N) Y	

CAS NO.	COMPOUND	CONCENTRATION UNITS: (ug/L or ug/kg) ug/kg	Q
12674-11-2	Aroclor-1016	17	U
11104-28-2	Aroclor-1221	17	U
11141-16-5	Aroclor-1232	17	1 77
53469-21-9	Aroclor-1242	17	- 11
12672-29-6	Aroclor-1248	17	U
11097-69-1	Aroclor-1254	17	Ω
11096-82-5	Aroclor-1260	17	11
37324-23-5	Aroclor-1262	17	TI.
11100-14-4	Aroclor-1268	17	Ü

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Lab Name: Da	taChem Laboratories, Inc.	Contract: EF	Y-W-05-026	
Lab Code: DA	TAC Case No.: 36799 Mod. Ref	No.:	SDG No.: B4FB5	
Matrix: (SOI	L/SED/WATER) SOIL	Lab Sample	ID: 7258004008	
Sample wt/vo	l: <u>30.0</u> (g/mL) g	Lab File ID:	20070928A034,200709	28B034
% Moisture:	Decanted: (Y/N) N			
			ed: 09/16/2007	
			d: <u>09/28/2007</u>	
Injection Vo	lume: 2.0 (uL) GPC Factor: 2.0			
			up: (Y/N) N	*
Acid Cleanup				
CAS NO.	COMPOUND		NCENTRATION UNITS:	Q
12674-11-2	Aroclor-1016		19	U
11104-28-2	Aroclor-1221		19	Ü
11141-16-5	Aroclor-1232		19	U
53469-21-9	Aroclor-1242		19	U
12672-29-6	Aroclor-1248		19	U
11097-69-1	Aroclor-1254			U

Aroclor-1260

Aroclor-1262

Aroclor-1268

11096-82-5

37324-23-5

11100-14-4

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Lab Name: DataChem Laboratories, Inc.	Contract: EP-W-05-026
Lab Code: DATAC Case No.: 36799 Mod. Ref	5 No.: SDG No.: B4FB5
Matrix: (SOIL/SED/WATER) SOIL	Lab Sample ID: 7258004009
Sample wt/vol: $30.0$ (g/mL) g	Lab File ID: 20070928A035,20070928B035
% Moisture: 10 Decanted: (Y/N) N	Date Received: 09/14/2007
Extraction: (Type) SONC	Date Extracted: 09/16/2007
Concentrated Extract Volume: 5000 (uL)	Date Analyzed: 09/28/2007
Injection Volume: 2.0 (uL) GPC Factor: 2.0	Dilution Factor: 1.0
GPC Cleanup: (Y/N) Y pH: 7.3	Sulfur Cleanup: (Y/N) N
Acid Cleanup: (Y/N) Y	

CAS NO.	COMPOUND	CONCENTRATION UNITS: (ug/L or ug/kg) ug/kg	Q
12674-11-2	Aroclor-1016	18	Ū
11104-28-2	Aroclor-1221	18	71
11141-16-5	Aroclor-1232	18	T7
53469-21-9	Aroclor-1242	18	Ū
12672-29-6	Aroclor-1248	18	U U
11097-69-1	Aroclor-1254	18	Ū
11096-82-5	Aroclor-1260	18	Ū
37324-23-5	Aroclor-1262	18	Ū
11100-14-4	Aroclor-1268	18	<u> </u>

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Lab Name: Da	taChem Laboratories, Inc.	Contract: EP-W-05-026	
Lab Code: DA	TAC Case No.: 36799 Mod. Ref	5 No.: SDG No.: B4FB5	
Matrix: (SOI	L/SED/WATER) SOIL	Lab Sample ID: 7258004010	
Sample wt/vo	ol: 30.0 (g/mL) g	Lab File ID: 20070928A036,20070928B03	— 6
% Moisture:	Decanted: (Y/N) N	Date Received: 09/14/2007	
Extraction:	(Type) SONC	Date Extracted: 09/16/2007	
Concentrated	Extract Volume: 5000 (uL)	Date Analyzed: 09/28/2007	
	lume: 2.0 (uL) GPC Factor: 2.0	· · · · · · · · · · · · · · · · · · ·	
GPC Cleanup:	(Y/N) <u>Y</u> pH: 7.5	Sulfur Cleanup: (Y/N) N	<del></del>
Acid Cleanup	: (Y/N) <u>Y</u>		
CAS NO.	COMPOUND	CONCENTRATION UNITS: (ug/L or ug/kg) ug/kg Q	
12674-11-2	Aroclor-1016	17 U	ᅥ
11104-28-2	Aroclor-1221	17 0	┪
11141-16-5	Aroclor-1232	17 U.	┪
53469-21-9	Aroclor-1242	17 U	$\dashv$
12672-29-6	Aroclor-1248	17 U	$\dashv$
11097-69-1	Aroclor-1254	17 U	$\dashv$
11096-82-5	Aroclor-1260	17 0	$\dashv$
37324-23-5	Aroclor-1262	17 0	+

11100-14-4

Aroclor-1268

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Lab Name: DataChem Laboratories, Inc.	Contract: EP-W-05-026
Lab Code: DATAC Case No.: 36799 Mod. Ref	No.: SDG No.: <u>B4FB5</u>
Matrix: (SOIL/SED/WATER) SOIL	Lab Sample ID: 7258004011
Sample wt/vol: 30.0 (g/mL) g	Lab File ID: 20070928A037,20070928B037
% Moisture: 87 Decanted: (Y/N) N	Date Received: 09/14/2007
Extraction: (Type) SONC	Date Extracted: 09/16/2007
Concentrated Extract Volume: 5000 (uL)	Date Analyzed: 09/28/2007
Injection Volume: 2.0 (uL) GPC Factor: 2.0	Dilution Factor: 1.0
GPC Cleanup: (Y/N) Y pH: 7.1	Sulfur Cleanup: (Y/N) N
Acid Cleanup: (Y/N) Y	

CAS NO.	COMPOUND	CONCENTRATION UNITS: (ug/L or ug/kg) ug/kg	Q
12674-11-2	Aroclor-1016	130	UJ
11104-28-2	Aroclor-1221	130	U
11141-16-5	Aroclor-1232	 130	Ü
53469-21-9	Aroclor-1242	130	Ū
12672-29-6	Aroclor-1248	130	Ū
11097-69-1	Aroclor-1254	130	U
11096-82-5	Aroclor-1260	130	Ü
37324-23-5	Aroclor-1262	130	עֿט
11100-14-4	Aroclor-1268	130	רַ ַ ט

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Lab Name: Da	taChem Laboratories, Inc.	Contract:	EP-W-05-026	
Lab Code: DA'	TAC Case No.: 36799 Mod. Ref	No.:	SDG No.: B4FB5	
Matrix: (SOI	L/SED/WATER) SOIL	Lab Sampl	e ID: 7258004012	-
Sample wt/vo	1: 30.0 (g/mL) g	Lab File	ID: 20070928A038,200709	28B038
% Moisture:	Decanted: (Y/N) N	Date Rece	ived: <u>09/14/2007</u>	
Extraction:	(Type) SONC	Date Extr	acted: 09/16/2007	
Concentrated	Extract Volume: 5000 (uL)	Date Anal	yzed: <u>09/28/2007</u>	
Injection Vo	lume: 2.0 (uL) GPC Factor: 2.0	Dilut:	ion Factor: 1.0	
GPC Cleanup:	(Y/N) <u>Y</u> pH: 6.3	Sulfur Cl	eanup: (Y/N) N	
Acid Cleanup	: (Y/N) <u>Y</u>			
CAS NO.	COMPOUND		CONCENTRATION UNITS: (ug/L or ug/kg) ug/kg	Q
12674-11-2	Aroclor-1016		45	Ū
11104-28-2	Aroclor-1221		45	U
11141-16-5	Aroclor-1232		45	. U
53469-21-9	Aroclor-1242		45	
12672-29-6	Aroclor-1248		45	U
11097-69-1	Aroclor-1254		9.4	JP 7N

11096-82-5

37324-23-5

11100-14-4

Aroclor-1260

Aroclor-1262

Aroclor-1268

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Lab Name: DataChem Laboratories, Inc.		Contract: EP-W-05-026		
Lab Code: DA'	TAC Case No.: 36799 Mod. Ref	E No.:	SDG No.: B4FB5	
Matrix: (SOI	L/SED/WATER) SOIL	Lab Samp]	le ID: 7258004013	
Sample wt/vo	1: 30.0 (g/mL) g		ID: 20070928A039,200709	928B039
	Decanted: (Y/N) N	Date Received: 09/14/2007		
Extraction:			racted: <u>09/16/2007</u>	
Concentrated	Extract Volume: 5000 (uL)	Date Anal	Lyzed: 09/28/2007	
Injection Vo	lume: 2.0 (uL) GPC Factor: 2.0		ion Factor: 1.0	•
GPC Cleanup:	(Y/N) Y pH: 6.2		leanup: (Y/N) N	
Acid Cleanup				
CAS NO.	COMPOUND	18	CONCENTRATION UNITS: (ug/L or ug/kg) ug/kg	Q
12674-11-2	Aroclor-1016		19	ט
11104-28-2	Aroclor-1221		19	. ט
11141-16-5	Aroclor-1232	2	19	Ū
53469-21-9	Aroclor-1242		19	Ū
12672-29-6	Aroclor-1248		19	Ü
11097-69-1	Aroclor-1254		4.1	J
11096-82-5	Aroclor-1260		10	TT

37324-23-5

11100-14-4

Aroclor-1262

Aroclor-1268

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Lab Name: DataChem Laboratories, Inc.	Contract: EP-W-05-026
Lab Code: DATAC Case No.: 36799 Mod. Ref	No.: SDG No.: B4FB5
Matrix: (SOIL/SED/WATER) SOIL	Lab Sample ID: 7258004014
Sample wt/vol: 30.0 (g/mL) g	Lab File ID: 20070928A040,20070928B040
% Moisture: 9.8 Decanted: (Y/N) N	Date Received: 09/14/2007
Extraction: (Type) SONC	Date Extracted: <u>09/16/2007</u>
Concentrated Extract Volume: 5000 (uL)	Date Analyzed: 09/28/2007
Injection Volume: 2.0 (uL) GPC Factor: 2.0	Dilution Factor: 1.0
GPC Cleanup: (Y/N) Y pH: 5.7	Sulfur Cleanup: (Y/N) N
Acid Cleanup: (Y/N) Y	

CAS NO.	COMPOUND	CONCENTRATION UNITS: (ug/L or ug/kg) ug/kg	Q
12674-11-2	Aroclor-1016	18	IJ.
11104-28-2	Aroclor-1221	18	IJ
11141-16-5	Aroclor-1232	18	· U
53469-21-9	Aroclor-1242	18	ū
12672-29-6	Aroclor-1248	18	IJ
11097-69-1	Aroclor-1254	18	IJ
11096-82-5	Aroclor-1260	18	<u> </u>
37324-23-5	Aroclor-1262	18	
11100-14-4	Aroclor-1268	18	

Lab Name: DataChem Laboratories, Inc.	Contract: EP-W-05-026
Lab Code: DATAC Case No.: 36799 Mod	d. Ref No.: SDG No.: <u>B4FB5</u>
Matrix: (SOIL/SED/WATER) SOIL	Lab Sample ID: 7258004015
Sample wt/vol: 30.0 (g/mL) g	Lab File ID: 20070928A041,20070928B041
% Moisture: 15 Decanted: (Y/N) N	Date Received: 09/14/2007
Extraction: (Type) SONC	Date Extracted: 09/16/2007
Concentrated Extract Volume: 5000 (	uL) Date Analyzed: 09/28/2007
Injection Volume: 2.0 (uL) GPC Factor: 2	.0 Dilution Factor: 1.0
GPC Cleanup: (Y/N) Y pH: 6.2	Sulfur Cleanup: (Y/N) N
Acid Cleanup: (Y/N) Y	
CAS NO. COMPOUND	CONCENTRATION UNITS: (ug/L or ug/kg) ug/kg

CAS NO.	COMPOUND	CONCENTRATION UNITS: (ug/L or ug/kg) ug/kg	Q
12674-11-2	Aroclor-1016	19	Ū
11104-28-2	Aroclor-1221	19	Ū
11141-16-5	Aroclor-1232	19	Ū
53469-21-9	Aroclor-1242	19	Ū
12672-29-6	Aroclor-1248	19	Ü
11097-69-1	Aroclor-1254	19	Ū
11096-82-5	Aroclor-1260	19	U.
37324-23-5	Aroclor-1262	19	ט
11100-14-4	Aroclor-1268	19	U

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Lab Name: Da	taChem Laboratories, Inc.	Contract:	EP-W-05-026	
Lab Code: DA	TAC Case No.: 36799 Mod. Ref	No.:	SDG No.: B4FB5	·.
Matrix: (SOI	L/SED/WATER) SOIL	Lab Sample	ID: 7258004016	
	ol: 30.0 (g/mL) g		ID: 20070928A046,20070	0928B046
	41 Decanted: (Y/N) N		ved: 09/14/2007	
Extraction:	(Type) SONC		acted: 09/16/2007	
Concentrated	Extract Volume: 5000 (uL)		zed: <u>09/28/2007</u>	
•	lume: 2.0 (uL) GPC Factor: 2.0			
•	(Y/N) Y pH: 5.9	•	eanup: (Y/N) N	
Acid Cleanup				
CAS NO.	COMPOUND		CONCENTRATION UNITS: (ug/L or ug/kg) ug/kg	Q
12674-11-2	Aroclor-1016		21	3 U
11104-28-2	Aroclor-1221		28	3 U
11141-16-5	Aroclor-1232		28	3 U
53469-21-9	Aroclor-1242		28	3 0
12672-29-6	Aroclor-1248		28	<del></del>
11097-69-1	Aroclor-1254		28	
11096-82-5	Aroclor-1260		28	
37324-23-5	Aroclor-1262		28	

11100-14-4

Aroclor-1268

EPA SAMPLE NO.

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Lab Name: Da	taChem Laboratories, Inc.	Contract: EP-W-05-026
Lab Code: DA	TAC Case No.: 36799 Mod. Ref	No.: SDG No.: B4FB5
Matrix: (SOI	L/SED/WATER) SOIL	Lab Sample ID: 7258004017
Sample wt/vo	1: 30.0 (g/mL) g	Lab File ID: 20070928A047,20070928B047
% Moisture:	Decanted: (Y/N) N	Date Received: 09/14/2007
Extraction:		Date Extracted: 09/16/2007
Concentrated	Extract Volume: 5000 (uL)	
	lume: 2.0 (uL) GPC Factor: 2.0	
	•	Sulfur Cleanup: (Y/N) N
Acid Cleanup	· · · · · · · · · · · · · · · · · · ·	
CAS NO.	COMPOUND	CONCENTRATION UNITS: (ug/L or ug/kg) ug/kg Q
12674-11-2	Aroclor-1016	47 U
11104-28-2	Aroclor-1221	47 U
11141-16-5	Aroclor-1232	47 U
53469-21-9	Aroclor-1242	47 U
12672-29-6	Aroclor-1248	47 U
11097-69-1	Aroclor-1254	47 U
11096-82-5	Aroclor-1260	47. U
37324-23-5	Aroclor-1262	47 U
11100-14-4	Aroclor-1268	477 17

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Lab Name: Dat	aChem Laboratories, Inc.	Contract: EP-W-05-026
Lab Code: DAT	AC Case No.: 36799 Mod. Re	f No.: SDG No.: <u>B4FB5</u>
Matrix: (SOII	JSED/WATER) SOIL	Lab Sample ID: 7258004018
Sample wt/vol	: 30.0 (g/mL) g	Lab File ID: 20070928A048,20070928B048
% Moisture: 1	6 Decanted: (Y/N) N	Date Received: 09/14/2007
Extraction: (	Type) SONC	Date Extracted: 09/16/2007
Concentrated	Extract Volume: 5000 (uL)	Date Analyzed: 09/28/2007
Injection Vol	ume: 2.0 (uL) GPC Factor: 2.0	Dilution Factor: 1.0
GPC Cleanup:	(Y/N) Y pH: 6.5	Sulfur Cleanup: (Y/N) N
Acid Cleanup:	(Y/N) <u>Y</u>	
CAS NO.	COMPOUND	CONCENTRATION UNITS: (ug/L or ug/kg) ug/kg Q
12674-11-2	Aroclor-1016	20 U
11104-28-2	Aroclor-1221	20 U

12674-11-2	Aroclor-1016		20 U
11104-28-2	Aroclor-1221		20 U
11141-16-5	Aroclor-1232		20 U
53469-21-9	Aroclor-1242		20 U
12672-29-6	Aroclor-1248		20 U
11097-69-1	Aroclor-1254		20 U
11096-82-5	Aroclor-1260		20 U
37324-23-5	Aroclor-1262		20 U
11100-14-4	Aroclor-1268		20 U

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Lab Name: DataChem La	aboratories, Inc.	Contract: EP-W-05-026
Lab Code: DATAC	Case No.: <u>36799</u> Mod. Re	ef No.: SDG No.: <u>B4F82</u>
Matrix: (SOIL/SED/WAT	rer) soil	Lab Sample ID: 7256028006
Sample wt/vol: 30.0	(g/mL) <u>g</u>	Lab File ID: 20070928A080,20070928B080
% Moisture: 49	Decanted: (Y/N) N	Date Received: 09/12/2007
Extraction: (Type) SO	ONC	Date Extracted: 09/16/2007
Concentrated Extract	Volume: 5000 (uL)	Date Analyzed: 09/29/2007
Injection Volume: 2.0	(uL) GPC Factor: 2.0	Dilution Factor: 1.0
GPC-Cleanup: $(Y/N)$ $\underline{Y}$	PH: 6.4	Sulfur Cleanup: (Y/N) N
Acid Cleanup: (Y/N) Y		
CAS NO. COMPOUND	)	CONCENTRATION UNITS:  (ug/L or ug/kg) ug/kg

CAS NO.	COMPOUND	CONCENTRATION UNITS: (ug/L or ug/kg) ug/kg	Q
12674-11-2	Aroclor-1016	32	· · ʊ
11104-28-2	Aroclor-1221	32	Ū
11141-16-5	Aroclor-1232	32	Ū
53469-21-9	Aroclor-1242	32	Ū
12672-29-6	Aroclor-1248	32	ט
11097-69-1	Aroclor-1254	32	Ü
11096-82-5	Aroclor-1260	32	U
37324-23-5	Aroclor-1262	32	Ü
11100-14-4	Aroclor-1268	32	Ü

		·	
Lab Name: Dat	aChem Laboratories, Inc.	Contract: EP-W-05-026	
Lab Code: DAT	TAC Case No.: 36799 Mc	Mod. Ref No.: SDG No.: B4F82	
Matrix: (SOI	L/SED/WATER) SOIL	Lab Sample ID: 7256028005	
Sample wt/vo.	1: <u>30.0</u> (g/mL) g	Lab File ID: 20070928A079,20070928B0	7.9
% Moisture: 8	Decanted: (Y/N) N	Date Received: 09/12/2007	
Extraction:	(Type) SONC	Date Extracted: 09/16/2007	
Concentrated	Extract Volume: 5000	(uL) Date Analyzed: 09/29/2007	
Injection Vol	lume: 2.0 (uL) GPC Factor: 2	2.0 Dilution Factor: 1.0	
GPC Cleanup:	(Y/N) Y pH: 6.5	Sulfur Cleanup: (Y/N) N'	
Acid Cleanup:	: (Y/N) <u>Y</u>		
CAS NO.	COMPOUND	CONCENTRATION UNITS: (ug/L or ug/kg) ug/kg	
12674-11-2	Aroclor-1016	120 п	

CAS NO.	COMPOUND		CONCENTRATION UNITS: (ug/L or ug/kg) ug/kg	Q
12674-11-2	Aroclor-1016		120	U
11104-28-2	Aroclor-1221		120	U
11141-16-5	Aroclor-1232		120	U
53469-21-9	Aroclor-1242		120	U
12672-29-6	Aroclor-1248		120	ָט
11097-69-1	Aroclor-1254		120	Ū
11096-82-5	Aroclor-1260		120	U
37324-23-5	Aroclor-1262		120	U
11100-14-4	Aroclor-1268		120	UJ

	*	
В4	F87	

Lab Name: DataChem Laboratories, Inc.	Contract: EP-W-05-026
Lab Code: DATAC Case No.: 36799 Mod. Ref	No.: SDG No.: <u>B4F82</u>
Matrix: (SOIL/SED/WATER) SOIL	Lab Sample ID: 7256028004
Sample wt/vol: 30.0 (g/mL) g	Lab File ID: 20070928A078,20070928B078
% Moisture: 18 / Decanted: (Y/N) N	Date Received: 09/12/2007
Extraction: (Type) SONC	Date Extracted: 09/16/2007
Concentrated Extract Volume: 5000 (uL)	Date Analyzed: 09/29/2007
Injection Volume: 2.0 (uL) GPC Factor: 2.0	Dilution Factor: 1.0
GPC Cleanup: (Y/N) Y pH: 6.5	Sulfur Cleanup: (Y/N) N
Acid Cleanup: (Y/N) Y	

CAS NO.	COMPOUND	CONCENTRATION UNITS: (ug/L or ug/kg) ug/kg	Q
12674-11-2	Aroclor-1016	20	Ū
11104-28-2	Aroclor-1221	20	Ü
11141-16-5	Aroclor-1232	20	Ū
53469-21-9	Aroclor-1242	20	U
12672-29-6	Aroclor-1248	20	Ū
11097-69-1	Aroclor-1254	20	U
11096-82-5	Aroclor-1260	20	ט
37324-23-5	Aroclor-1262	20	U
11100-14-4	Aroclor-1268	20	Ū

B4F86	

Lab Name: DataChem Laboratories, Inc.	Contract: EP-W-05-026		
Lab Code: DATAC Case No.: 36799 Mod. Ref	No.: SDG No.: <u>B4F82</u>		
Matrix: (SOIL/SED/WATER) SOIL	Lab Sample ID: 7256028003		
Sample wt/vol: 30.0 (g/mL) g	Lab File ID: 20070928A077,20070928B077	7	
% Moisture: 0.54 / Decanted: (Y/N) N	Date Received: 09/12/2007		
Extraction: (Type) SONC	Date Extracted: 09/16/2007		
Concentrated Extract Volume: 5000 (uL)	Date Analyzed: 09/29/2007		
Injection Volume: 2.0 (uL) GPC Factor: 2.0	Dilution Factor: 1.0		
GPC Cleanup: (Y/N)'Y pH: 4.6	Sulfur Cleanup: '(Y/N) N		
Acid Cleanup: (Y/N) Y			

CAS NO.	COMPOUND				CONCENTRATION UNITS: (ug/L or ug/kg) ug/kg	Q
12674-11-2	Aroclor-1016			*	17	Ü
11104-28-2	Aroclor-1221				17	Ü.
11141-16-5	Aroclor-1232	1.			17	Ü
53469-21-9	Aroclor-1242	.*			17.	יט
12672-29-6	Aroclor-1248				17	Ū
11097-69-1	Aroclor-1254	:			17	Ū
11096-82-5	Aroclor-1260				17	Ü
37324-23-5	Aroclor-1262				17	U
11100-14-4	Aroclor-1268				17	Ü

D 4 DO 0		
B4F83		

Lab Name: DataChem Laboratories, Inc.	Contract: EP-W-05-026
Lab Code: DATAC Case No.: 36799 Mod. Ref	No.: SDG No.: B4F82
Matrix: (SOIL/SED/WATER) SOIL	Lab Sample ID: 7256028002
Sample wt/vol: 30.0 (g/mL) g	Lab File ID: 20070928A076,20070928B076
% Moisture: 11 / Decanted: (Y/N) N	Date Received: 09/12/2007
Extraction: (Type) SONC	Date Extracted: 09/16/2007
Concentrated Extract Volume: 5000 (uL)	Date Analyzed: 09/29/2007
Injection Volume: 2.0 (uL) GPC Factor: 2.0	Dilution Factor: 1.0
GPC Cleanup: (Y/N) Y pH: 6.9	Sulfur Cleanup: (Y/N) N
Acid Cleanup: (Y/N) Y	

CAS NO.	COMPOUND	CONCENTRATION UNITS: (ug/L or ug/kg) ug/kg	Q
12674-11-2	Aroclor-1016	18	U
11104-28-2	Aroclor-1221	18	Ū
11141-16-5	Aroclor-1232	18	Ū
53469-21-9	Aroclor-1242	18	Ū
12672-29-6	Aroclor-1248	18	U
11097-69-1	Aroclor-1254	18	U U
11096-82-5	Aroclor-1260	18	U
37324-23-5	Aroclor-1262	18	TT T
11100-14-4	Aroclor-1268	18	U

B4F82	

Lab Name: DataChem Laboratories, Inc.	Contract: EP-W-05-026
Lab Code: DATAC Case No.: 36799 Mod. Ref	No.: SDG No.: <u>B4F82</u>
Matrix: (SOIL/SED/WATER) SOIL	Lab Sample ID: 7256028001
Sample wt/vol: 30.0 (g/mL) g	Lab File ID: 20070928A075,20070928B075
% Moisture: 87 Decanted: (Y/N) N	Date Received: 09/12/2007
Extraction: (Type) SONC	Date Extracted: 09/16/2007
Concentrated Extract Volume: 5000 (uL)	Date Analyzed: 09/29/2007
Injection Volume: 2.0 (uL) GPC Factor: 2.0	Dilution Factor: 1.0
GPC Cleanup: (Y/N) Y pH: 5.3	Sulfur Cleanup: (Y/N) N
Acid Cleanup: (Y/N) Y	

CAS NO.	COMPOUND			CONCENTRATION UNITS (ug/L or ug/kg) ug/l	-	Q	
12674-11-2	Aroclor-1016				120	Ü	Ī
11104-28-2	Aroclor-1221	•			120	U.	1
11141-16-5	Aroclor-1232				120	Ū	Γ
53469-21-9	Aroclor-1242				120	Ū	Γ
12672-29-6	Aroclor-1248				120	Ū	
11097-69-1	Aroclor-1254			1	120	u U	Ī
11096-82-5	Aroclor-1260			]	L20	U	
37324-23-5	Aroclor-1262			 1	20	Ū ·	V
11100-14-4	Aroclor-1268		•,	1	20	ָ ט	Ī

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B41	:92		
	-		

Lab Name: DataChem Laboratories, Inc.	Contract: EP-W-05-026
Lab Code: DATAC Case No.: 36799 Mod. Ref	No.: SDG No.: <u>B4F82</u>
Matrix: (SOIL/SED/WATER) SOIL	Lab Sample ID: 7256028007
Sample wt/vol: $30.0$ (g/mL) g	Lab File ID: 20070928A081,20070928B081
% Moisture: 21 Decanted: (Y/N) N	Date Received: 09/12/2007
Extraction: (Type) SONC	Date Extracted: 09/16/2007
Concentrated Extract Volume: 5000 (uL)	Date Analyzed: 09/29/2007
Injection Volume: 2.0 (uL) GPC Factor: 2.0	Dilution Factor: 1.0
GPC Cleanup: (Y/N) Y pH: 6.8	Sulfur Cleanup: (Y/N) N
Acid Cleanup: (Y/N) Y	

CAS NO.	COMPOUND	CONCENTRATION UNITS: (ug/L or ug/kg) ug/kg	Q
12674-11-2	Aroclor-1016	21	Ū
11104-28-2	Aroclor-1221	21	Ū
11141-16-5	Aroclor-1232	21	Ü
53469-21-9	Aroclor-1242	21	Ū
12672-29-6	Aroclor-1248	21	מ
11097-69-1	Aroclor-1254	21	Ū
11096-82-5	Aroclor-1260	21	Ū
37324-23-5	Aroclor-1262	21	Ü
11100-14-4	Aroclor-1268	21	ט

B4	F95	

Lab Name: DataChem Laboratories, Inc.	Contract: EP-W-05-026
Lab Code: DATAC Case No.: 36799 Mod. Ref	5 No.: SDG No.: <u>B4F82</u>
Matrix: (SOIL/SED/WATER) SOIL	Lab Sample ID: 7256028008
Sample wt/vol: $30.0$ (g/mL) g	Lab File ID: 20070928A082,20070928B082
% Moisture: 82 Decanted: (Y/N) N	Date Received: 09/12/2007
Extraction: (Type) SONC	Date Extracted: 09/16/2007
Concentrated Extract Volume: 5000 (uL)	Date Analyzed: 09/29/2007
Injection Volume: 2.0 (uL) GPC Factor: 2.0	Dilution Factor: 1.0
GPC Cleanup: (Y/N) Y pH: 6.7	Sulfur Cleanup: (Y/N) N
Acid Cleanup: (Y/N) Y	

CAS NO.	COMPOUND	CONCENTRATION UNITS: (ug/L or ug/kg) ug/kg	Q
12674-11-2	Aroclor-1016	90	U
11104-28-2	Aroclor-1221	90	
11141-16-5	Aroclor-1232	90	U
53469-21-9	Aroclor-1242	90	U
12672-29-6	Aroclor-1248	90	U
11097-69-1	Aroclor-1254	90	U
11096-82-5	Aroclor-1260	90	U
37324-23-5	Aroclor-1262	90	, ט
11100-14-4	Aroclor-1268	90	UT

were the second man	B4F96	***********
	B4196	

Lab Name: DataChem Laboratories, Inc.	Contract: EP-W-05-026
Lab Code: DATAC Case No.: 36799 Mod. Ref	No.: SDG No.: <u>B4F82</u>
Matrix: (SOIL/SED/WATER) SOIL	Lab Sample ID: 7256028009
Sample wt/vol: $30.0$ (g/mL) g	Lab File ID: 20070928A083,20070928B083
% Moisture: 90 / Decanted: (Y/N) N	Date Received: 09/12/2007
Extraction: (Type) SONC	Date Extracted: 09/16/2007
Concentrated Extract Volume: 5000 (uL)	Date Analyzed: 09/29/2007
Injection Volume: 2.0 (uL) GPC Factor: 2.0	Dilution Factor: 1.0
GPC Cleanup: (Y/N) Y pH: 7.3	Sulfur Cleanup: (Y/N) N
Acid Cleanup: (Y/N) Y	

CAS NO.	COMPOUND	CONCENTRATION UNITS: (ug/L or ug/kg) ug/kg	Q
12674-11-2	Aroclor-1016	170	-IR
11104-28-2	Aroclor-1221	170	J/M
11141-16-5	Aroclor-1232	170	ш
53469-21-9	Aroclor-1242	170	JJ.
12672-29-6	Aroclor-1248	170	<b>8</b>
11097-69-1	Aroclor-1254	170	25
11096-82-5	Aroclor-1260	170	Ja I
37324-23-5	Aroclor-1262	170	وأصلا
11100-14-4	Aroclor-1268	170	UR

B4F99	

Lab Name: DataChem Laboratories, Inc.	Contract: EP-W-05-026
Lab Code: DATAC Case No.: 36799 Mod. Ref	No.: SDG No.: B4F82
Matrix: (SOIL/SED/WATER) SOIL	Lab Sample ID: 7256028010
Sample wt/vol: 30.0 (g/mL) g	Lab File ID: 20070928A084,20070928B084
% Moisture: 23 Decanted: (Y/N) N	Date Received: 09/12/2007
Extraction: (Type) SONC	Date Extracted: 09/16/2007
Concentrated Extract Volume: 5000 (uL)	Date Analyzed: 09/29/2007
Injection Volume: 2.0 (uL) GPC Factor: 2.0	Dilution Factor: 1.0
GPC Cleanup: (Y/N) Y pH: 6.5	Sulfur Cleanup: (Y/N) N
Acid Cleanup: (Y/N) Y	

CAS NO.	COMPOUND	CONCENTRATION UNITS: (ug/L or ug/kg) ug/kg	Q
12674-11-2	Aroclor-1016	21	Ü
11104-28-2	Aroclor-1221	21	ט
11141-16-5	Aroclor-1232	21	Ū
53469-21-9	Aroclor-1242	21	Ū.
12672-29-6	Aroclor-1248	21	U
11097-69-1	Aroclor-1254	21	U
11096-82-5	Aroclor-1260	21	IJ
37324-23-5	Aroclor-1262	21	Ü
11100-14-4	Aroclor-1268	21	Ü

EPA SAMPLE NO.

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В4	FJ1	L	

Lab Name: DataChem Laboratories, Inc. Contract: EP-W-05-026				
Lab Code: DAT	TAC Case No.: 36799 Mod. Ref	f No.: SDG No.: <u>B4FJ1</u>		
Matrix: (SOI	L/SED/WATER) SOIL	Lab Sample ID: 7306027001		
Sample wt/vo	1: <u>30.0</u> (g/mL) g	Lab File ID: 20071107A025,20071107B025		
% Moisture: 2	Decanted: (Y/N) N	Date Received: 11/01/2007		
•	(Type) SONC	Date Extracted: 11/04/2007		
	Extract Volume: 10000 (uL)	Date Analyzed: 11/07/2007		
	lume: 2.0 (uL) GPC Factor: 2.0			
	(Y/N) Y pH: 6.4			
Acid Cleanup	· · · · · · · · · · · · · · · · · · ·			
CAS NO.	COMPOUND	CONCENTRATION UNITS: (µg/L or ug/kg) ug/kg Q		
12674-11-2	Aroclor-1016	41 U		
11104-28-2	Aroclor-1221	41 U		
11141-16-5	Aroclor-1232	41 U		
53469-21-9	Aroclor-1242	41 U		
12672-29-6	Aroclor-1248	41 U		
11097-69-1	Aroclor-1254	41 U		
11096-82-5	Aroclor-1260	41 U		

37324-23-5

11100-14-4

Aroclor-1262 Aroclor-1268

B4FJ2	

Lab Name: Dat	aChem Laboratories, Inc.	Contract: EP-W	I-05-026.		
Lab Code: DAT	AC Case No.: 36799 Mod. Ref	No.:	SDG No.: <u>B4FJ1</u>		
Matrix: (SOI)	L/SED/WATER) SOIL	Lab Sample ID	7306027002		·
Sample wt/vo	l: 30.0 (g/mL) g	Lab File ID: 2	0071107A026,2007110	)7B026	<u>5</u>
	5 Decanted: (Y/N) N		•		
	(Type) SONC	Date Extracted			
	Extract Volume: 10000 (uL)	Date Analyzed	: 11/08/2007		
	lume: 2.0 (uL) GPC Factor: 2.0				
	(Y/N) Y pH: 7.0	Sulfur Cleanu		-	
	: (Y/N) <u>Y</u>		· · · · · · · · · · · · · · · · · · ·		
CAS NO.	COMPOUND	· · · · · · · · · · · · · · · · · · ·	ENTRATION UNITS: (L or ug/kg) ug/kg	Q	
12674-11-2	Aroclor-1016	ļ <u>;</u>	39	ָ ט .	7
11104-28-2	Aroclor-1221		39	Ū	
11141-16-5	Aroclor-1232		39	Ū	
53469-21-9	Aroclor-1242		39	Ū	
12672-29-6	Aroclor-1248	,	39	U	
11097-69-1	Aroclor-1254		39	Ū	
11096-82-5	Aroclor-1260		. 39	Ü	
37324-23-5	Aroclor-1262		39	U	
11100 14 4	71 1000		30	TT:	

EPA SAMPLE NO.

B4FJ3	

		•		
Lab Name: DataChem Laboratories, Inc.		Contract: EP-W-05-026		
Lab Code: DAT	CAC Case No.: 36799 Mod. Ref	No.: SDG No.: <u>B4FJ1</u>		
Matrix: (SOIL/SED/WATER) SOIL		Lab Sample ID: 7306027003		
Sample wt/vol: 30.0 (g/mL) g		Lab File ID: 20071107A027,20071107B027		
% Moisture: 16 Decanted: (Y/N) N		Date Received: 11/01/2007		
Extraction:	(Type) SONC	Date Extracted: 11/04/2007		
	Extract Volume: 10000 (uL)	Date Analyzed: 11/08/2007		
Injection Vo.	lume: 2.0 (uL) GPC Factor: 2.0	Dilution Factor: 1.0		
GPC Cleanup: (Y/N) Y pH: 5.2 Sulfur Cleanup: (Y/N) N			anup: (Y/N) N	
Acid Cleanup: (Y/N) Y				
CAS NO.	COMPOUND		CONCENTRATION UNITS: (µg/L or ug/kg) ug/kg	Q
12674-11-2	Aroclor-1016		39	リズ
11104-28-2	Aroclor-1221		39	U Ì
11141-16-5	Aroclor-1232		39	Ū
53469-21-9	Aroclor-1242		39	Ū´
12672-29-6	Aroclor-1248		39	Ü
11097-69-1	Aroclor-1254		<sup>.</sup> 39	Ū
11096-82-5	Aroclor-1260		39	Ū,
37324-23-5	Aroclor-1262		.39	U

11100-14-4

Aroclor-1268

EPA SAMPLE NO.

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													B4FJ4			
B4 FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4				
B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4				
B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4				
B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4				
B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4				
B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4				
B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4				
B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4				
B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4				
B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4	B4FJ4				
B4FJ4	B4FJ4	B4104	B4FJ4	B4FJ4	B4FJ4	B4104	B4F04	B4F04	B4FJ4	B4FJ4	B4FJ4	B4FJ4				
B4FJ4	B4104	B4104	B4104	B4104	B4104	B4104	B4104	B4104	B4FJ4	B4FJ4	B4FJ4	B4FJ4				
D4104	P01PG	P01PG	D4104	D4104	D4104	D4104	D4104	D4104	D4104	D4104	D4104	D4104				
D4104	P01PG	P01PG	D4104	D4104	D4104	D4104	D4104	D4104	D4104	D4104	D4104	D4104				
D41U4	P01PG	P01PG	D4104	D4104	DALUA	D4104	D4104	P01P0	D4104	D4104	DALUA	P014G				
	DALOA	DALOA	DALOA	DALUA	DALUA	DALOA	DALUA	DALUA	DALOA	DALOA	DALUA	DALOA				
		Daloa	Daroa	DATOA	DATOA	Daroa	Datioa	Datioa	Daroa	Daroa	Daroa	DALOA				
			Daroa	DATOA	DATOA	Daroa	Datioa	Datioa	Daroa	Daroa	Daroa	DALOA				
				ロオエロオ	ロオエロオ	DATOA	Daroa	Daroa	Daroa	Daroa	Daroa	Daroa				
					Daroa	D4104	777.07	777.07	Daroa	Daroa	Daroa	Daroa				
						22 2 2 2 2	22 2 2 2 2	22101	27101	27101	21101	21101				

Lab Name: Dat	aChem Laboratories, Inc.	Contract: EP-W-05-026
Lab Code: DAT	PAC Case No.: 36799 Mod. Ref	No.: SDG No.: <u>B4FJ1</u>
Matrix: (SOI)	L/SED/WATER) <u>SOIL</u>	Lab Sample ID: 7306027004
Sample wt/vo.	1: <u>30.0 (g/mL) g</u>	Lab File ID: 20071107A028,20071107B028
	5 Decanted: (Y/N) N	Date Received: 11/01/2007
Extraction:	(Type) SONC	Date Extracted: 11/04/2007
		Date Analyzed: 11/08/2007
	lume: 2.0 (uL) GPC Factor: 2.0	Dilution Factor: 1.0
	(Y/N) Y pH: 6.4	
	: (Y/N) <u>Y</u>	
CAS NO.	COMPOUND	CONCENTRATION UNITS: [ug/L or ug/kg) ug/kg Q
12674-11-2	Aroclor-1016	39 U 🕽
11104-28-2	Aroclor-1221	39 U. <b>/</b>
11141-16-5	Aroclor-1232	39 0
53469-21-9	7roglor-1242	20 17

12672-29-6

11097-69-1

11096-82-5

37324-23-5

11100-14-4

Aroclor-1248

Aroclor-1254

Aroclor-1260

Aroclor-1262

Aroclor-1268

EPA SAMPLE NO.

B4FJ5	

	•	
Lab Name: Dat	taChem Laboratories, Inc.	Contract: EP-W-05-026
Lab Code: DAT	FAC Case No.: 36799 Mod. Ref	No.: SDG No.: B4FJ1
Matrix: (SOI	L/SED/WATER) SOIL	Lab Sample ID: 7306027005
		Lab File ID: 20071107A029,20071107B029
	Decanted: (Y/N) N	
	(Type) SONC	Date Extracted: 11/04/2007
	Extract Volume: 10000 (uL)	
	lume: 2.0 (uL) GPC Factor: 2.0	
	(Y/N) Y pH: 7.1	Sulfur Cleanup: (Y/N) N
Acid Cleanup		-
CAS NO.	COMPOUND	CONCENTRATION UNITS:  (ug/L or ug/kg) ug/kg Q
12674-11-2	Aroclor-1016	39 0
11104-28-2	Aroclor-1221	39 U
11141-16-5	Aroclor-1232	39 U
53469-21-9	Aroclor-1242	39 U
12672-29-6	Aroclor-1248	39 U
11097-69-1	Aroclor-1254	39 U
11096-82-5	Aroclor-1260	39 U
37324-23-5	Aroclor-1262	39 U.

11100-14-4

Aroclor-1268

EPA SAMPLE NO.

B4FJ6	

Lab Name: Dat	aChem Lab	oratories, Inc.		Contract:	EP-W-05-026	
Lab Code: DAT	'AC	Case No.: 36799	_ Mod. Ref	E No.:	SDG No.: B4FJ1	
Matrix: (SOI)	L/SED/WAT	ER) SOIL		Lab Sampl	e ID: 7306027006	·
Sample wt/vo.	1: 15.0	(g/mL) <u>g</u>		Lab File	ID: 20071107A030,200711	07B030
% Moisture: 4	8	Decanted: (Y/N)	<u>N</u>	Date Rece	eived: 11/01/2007	·
Extraction:	(Type) SO	1C		Date Extr	acted: 11/04/2007	
Concentrated	Extract	Volume: 10000	(uL)	Date Anal	yzed: <u>11/08/2007</u>	- <u> </u>
Injection Vo	lume: 2.0	(uL) GPC Facto	or: <u>2.0</u>	Dilut	ion Factor: 1.0	
GPC Cleanup:	(Y/N) <u>Y</u>	pH: 6.2		Sulfur Cl	eanup: (Y/N) N	
Acid Cleanup	: (Y/N) <u>Y</u>		•			
CAS NO.	COMPOUND			! n	CONCENTRATION UNITS: (ug/L or ug/kg) ug/kg	Q

CAS NO.	COMPOUND	1	CONCENTRATION UNITS: (ug/L or ug/kg) ug/kg	Q
12674-11-2	Aroclor-1016		130	υJ
11104-28-2	Aroclor-1221		130	<del>                                     </del>
11141-16-5	Aroclor-1232		130	U
53469-21-9	Aroclor-1242	****	130	U
12672-29-6	Aroclor-1248	•	. 130	Ü
11097-69-1	Aroclor-1254		130	U.
11096-82-5	Aroclor-1260		130	Ū
37324-23-5	Aroclor-1262		130	Ū.
11100-14-4	Aroclor-1268	¥ .	130	ū

EPA SAMPLE NO.

_		_	
Ŀ	34 FJ	1	

		•
Lab Name: Dat	aChem Laboratories, Inc.	Contract: EP-W-05-026
Lab Code: DAT	AC Case No.: 36799 Mod. Ref	No.: SDG No.: <u>B4FJ1</u>
Matrix: (SOII	L/SED/WATER) SOIL	Lab Sample ID: 7306027007
Sample wt/vol	l: <u>30.0</u> (g/mL) g	Lab File ID: 20071107A031,20071107B031
% Moisture: 3	Decanted: (Y/N) N	Date Received: <u>11/01/2007</u>
Extraction:	(Type) SONC	Date Extracted: 11/04/2007
Concentrated	Extract Volume: 10000 (uL)	Date Analyzed: 11/08/2007
Injection Vol	lume: 2.0 (uL) GPC Factor: 2.0	Dilution Factor: 1.0
GPC Cleanup:	(Y/N) Y pH: 5.7	Sulfur Cleanup: (Y/N) N
Acid Cleanup:	·	
CAS NO.	COMPOUND	CONCENTRATION UNITS: [(ug/L or ug/kg) ug/kg Q
12674-11-2	Aroclor-1016	34 U J
11104-28-2	Aroclor-1221	34 U
11141-16-5	Aroclor-1232	34 U
53469-21-9	Aroclor-1242	34 Ü
12672-29-6	Aroclor-1248	34 U
11097-69-1	Aroclor-1254	34 U
11096-82-5	Aroclor-1260	34 U
37324-23-5	Aroclor-1262	34 U

Aroclor-1268

11100-14-4

EPA SAMPLE NO.

ALCSS1(1)

			•	
Lab Name: Dat	aChem Laboratories, Inc.	Contract:	EP-W-05-026	
Lab Code: DAT	AC Case No.: 36799 Mod. Ref	No.:	SDG No.: B4FJ1	
Matrix: (SOI)	L/SED/WATER) SOIL	Lab Sampl	e ID: <u>33834</u>	-
Sample wt/vo	:: 30.0 (g/mL) g	Lab File	ID: 20071107A024	`
	.0 Decanted: (Y/N) N	Date Rece	ived:	
Extraction:	(Type) SONC	Date Extr	acted: 11/04/2007	
Concentrated	Extract Volume: 10000 (uL)	Date Anal	yzed: <u>11/07/2007</u>	
Injection Vo.	Lume: 2.0 (uL) GPC Factor: 2.0	Diluti	ion Factor: 1.0	
GPC Cleanup:	(Y/N) Y pH:	Sulfur Cl	eanup: (Y/N) N	
	: (Y/N) <u>Y</u>			
CAS NO.	COMPOUND	1	CONCENTRATION UNITS: (ug/L or ug/kg) ug/kg	Q
12674-11-2	Aroclor-1016		37	١.
11104-28-2	Aroclor-1221		33	Ū
11141-16-5	Aroclor-1232		33	U
53469-21-9	Aroclor-1242		. 33	Ü
12672-29-6	Aroclor-1248		33	Ū
11097-69-1	Aroclor-1254		33	Ū
11096-82-5	Aroclor-1260		37	
37324-23-5	Aroclor-1262		33	Ū
11100-14-4	Aroclor-1268	v	33	Ū

## **APPENDIX D**

# FIELD DOCUMENTATION AND LOGBOOK

10 .	t 7007		
VINE	7. 2007 LAND CHEM (	00#3 + #4	T = T = T
ROUM		WATER	
"BA" B	EACH		
Sample	ID		
Arsenic	ļ	Time Jane	
	e-100907 B4F8		
	100907 BYF83		
	341-100907	Maa	
BA-WAT	1-100907 BYF	34 1410	
8A-WAT	2-100907 BYF.	35 1411	
BA-BEAC	4-100907 BYF	86 1340	
BA - BEAC	W-M5-100907	1340	
BA - BEA	CH-115D-100907	1340	
Coordin	ate NJ S	tate Plane NAD 83	
Location	North (FX)	Eart (A)	
BA-Shor			
8A - Sed	1	330352,4	
+ WAT 1/1	ا     ! !   !     ا		
BA-Beach	238011.2	330374.8	
			$\wedge$

	-	-	-	_	_
L	40.0	0			
₽ — <u> </u>	10 SEP.				
<u></u> -	Emp = 23	at qual			<u>-</u>
	al = 0.0	7 .		11 ga 80.	
	and in	5 ms/cm	pH- 7		
	<u></u>	J my /cm'	Jurb."	ONTU	-
	Checo	Avenue			
Acr	enic	Menus	-	les	
_	1	e-100907	PCB		Line
		d-100907	84F8		1515
She	man Sed	- DUP- 100	307 00		1510
Sher	man - lal	2+1-10090	97 B4F8	9	1510
She	man - W	at 2 - 1009	07 BYF9		1500
	7 7	1001	5 17-		1501
	30- unt	qual.@			<del> </del>
Te	m, 2 22,	67	Do = 7.		+
_ Sa	l. = 0.0	59	04:7	į	<del>                                     </del>
Con	d: 0,1	93	Turb =		<del> </del>
•			197.5,	677	<del> </del> -
	condina	rta	v_	E	
	rman- 1 tl Wat		4390,3	330557	
She	erman	Share 22	4384.3	330561	1
· · · · · · · · · · · · · · · · · · ·					
					(2)
	ļ	İ	1		6

10 SEP. 2007

10 321, 200 i	
West of Mill Rd.	
Arsenia PCB	Time
Mill-Share-10907 B4F9	1700
MIII - Shore - MS-100907	1700
Mill- Mare- HSD-100907	1700
Hill-Sed-100907 B4F	92 1710
Mill-Wat1-100707 BHF	
Mill- Wat 2 - 100907 BY	
1714 - Not core @ Mill (	+ Mill-Sed (2)
N 247692.5 F4	
F 384013.9 FA	
7 H core barrel	
Pen = 2.9 ft outside of 5	
Comp + 3,5 At in ride of &	
Cop 1 3,0 1 1 1 1 2 7	
173	,
1730- and qual @ Mill	
	21
Sal = 0.04 pH=	7.6
Cand : 0,121 Turk	5, 20
Coordinate for M.W. S.G.	ione
N 247687, P //	
E 334 324.5 A	
	(3)
• 1	(・ク)

/			!		
10 5	ep. 200	7			
- Equ	in blan	ka			
PBlan	k-01-	100907	1800	Arien	·.`c
11		100907			
	84FL1 84FL0	1800 -	PCB		
	<del>.</del>				
1	ep. 200	1 _	/		
ARSENIC	,	5 Samp	св	Time	
R55-Show			4F 95	0900	
R55-√ec				0910	
R55-Sec				0910	
R55-Sec	1-MSD-	110907		0910	
855-W	AT 1- 110	907 6	4F97	0850	
R55- W	7T1-11C	907 B	4F98	0851	
0943-	unt. 91	wal. @	Rt. 55		
Temp=			Do = 8.	5	
Conda		5	p H= 6	3,0	
_	-0.0		Turb	1	
0949	Core a	Rt. 55			
)		768.26			
	E 3313	58.3 H			
	barcel =				
			tride of	barel	
	Camp: 7	Off ou	7 Ft ins	ide	

2448499

2448654

BWB- Stad

(5)

329 105.7

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		ep. 20	07			
	ALLIA	NE 8	EACH			
	Arsenic			β		<u>ne</u> _
	Alliance-			4FA3	ļ	320
	Alliance-			YFAY.	-	25
	Alliance -			4FA5		30
	Alliana-					330
	Alliance-			B4FA6		15
	Alliance-	Wat 2-1	0907	84FA7		16
					(MS) 1	
			<u> </u>	BYFA9	(MSD) 1	3/6
	1404 - Tem Con	F rel = 6 rel = 6 rest = 4.0 rest = 22.0 d = 0.14	24392 32859 FH OUTU 9 PH 15.	8.2 ft 9.8A Vide of Wiance Do: 7.4	Barrel 34	
	Jal	. = 6.6	<b>f</b>	Turb. « c	0.0	0

11 Sep. 2007	··· <del>····</del> ····		<del></del>	ı— <del>I</del> ı <del></del>
Coord	W		E	
Alliance - Sed	2439	282/	4 3285	99,84
Wat Watz				
ALLIANTE - SHORE	243 94	14, 2	32856	7.61
Alliance Beach	243 96	66,7 M	32857	
ALMOND BEACH				
Arsenia		PCB		ine
Almond Shore-11090	37	BYFBO		1515
Almond Shore DUP-11	0907		1 - 1 - 1 - 1 - 1 - 1 - 1	1515
Almond - Sed - 11090	37	84 FB1		1525
Almond - Wat 1-11	0907	BYFB2		1500
Almand - Wat 2 - 11	0907	BYFB3		1501
Almond - Beach -	110907	84FB5	4	1530
1527- core @ A	and	(+ SED	and WA	1/6/11/2)
W 341	822.5	T (+)		
E 329		PH		
Core Garrel =	6 62			
1 1 1 1 1 1 1 1 1 1 1 1	outride of	1 1 1 1 1	<del>- -</del>  - - -	
Can = 3.6	At	ide of	Sarcel	
	4	<u> </u>		
	4/833.		329545	1 1 1.
Almand Barch	14/833	13 AH	329554	9 6
				(T)

,					
	11 Sep. 2	007			···
	1538-	val qual @	Almond	Beach	
	Tem:		Do. 7.		
•	Cond :	i	PHZ 7.	16	
	Sal = 0		Turb = c	9. <i>0</i>	
	4 cooler	or shipped to	Datach	om	
•		pries & PCI	1		
	Blanks		PCB	ź I	iox
	PBlank-	-02-110907	+ B4FK	171	<b>5</b>
	BS8 lank	(-02 110907	+ BYFK	172	5
	12 SE	p. 2007			
	Blanke	7 0	PC	86	
		-03-12090-		_	0805
		-03-12090			0810
	- "	4-120907			0815
		1-120907	+ 841	K4	0820
	Workh	End of Oni.	n Lake		
	Arrenia		er a	Time	-
	NUL-SED-1	20907 5	F 84F85	1100	
	NUL-WATI-		34FB6	1115	
	NUL-WATI	- DUP-120907	B4F57	1116	
	PUL WAT	RE-120907	34F88	1125	L STATIS

12 SEr. 2007	
Coard	N
NUL-SED + WAT 1/WAT 2	21975664 33123134
NOL-Shore	2197297 B 33/802,9A
1/21 - wat qual @ N (	).
	+1,21
Cond = 0.176 pH=	7.7
	6 - 2.5
UNION LAKE BEACH	
ARMENIA PC.	B TIME
	F 89 1230
	F40 1220
ULB-Wax1-120907 BY	
ULB-WAST-120907 B4	
ULB- W212-DUR-12096	
ULB-BEAKH-120907	
ULB-BEACH-HJ-12095	1   1   [   + 1   1,1
ULB-BEACH-HIB-123	
1214- wat gual a Union	Cake Beach
	= 6,99
	1: 7.43
	6 0.0
	(9)

·	12 Sep. 2007		
	Coordinator		
	ULB- Sed, WATI / WE	210474	84 335173.84
	ULB-Beach	210346.	1 FL 335387,4A
	ULB- Share	2/0356	,3Q 335358.2Q
	1340 - core a Un	ion Lake Bea	d.
	N 210358	,914	
	E 335 205,	-	
	SOUTH END UN	ION LAKE BE	ACH
	Arsenie	PCB	Time
	SUL-Shore-120907	84FC4	1450
	SUL- Sed-120907	84FC5	1440
	SUL-Wat 1-120907	BYFCL	1430
	SUL-Wat2-120907	B4FC7	1431
÷	JUL-Beach - 12090	7 B4FC8	1500
	SUL-Beach-DUP-12	0907	1500
	1447 - hat qual a	» SUL	
	Tem. = 25.3/	Do = 7.23	
	Cand = 0.160	pH= 7.58	
	Sal 0.07	Turb = 0.	0

12 SE1. 2007 336 375,6 A SUL-Shore SUL-Beach 208799,24 336532.14 336 5 55. 9 208812.20 1535 - core N 208746.2 A E 336485-3 R 2007 Union Cak SUL-Baring PCB = B4C9 B4FC9 for EZCR (14arge Almond Beach Rec 1155 Almond - Boring - 1 0-11 BYFDO PCB Almond-Baring-Z 1156 168 BYFDI Almond-Boring 1155

M

•	/3	Sep. 200	7			
		st of Rt		Rec. = 4.	ft.	
	Time	Sample		Analys	<u>'                                     </u>	Interva
	1215	R55- B	pring-1	arsen	je	0-1'
		BYFD	2	PCB		
		155- Bor	ing - 1	Grain	Size	
	1216	155-B	oring-2	arsen	/c	1-2'
2		BHFD	<b></b>	PCB	ļ	
	1217	R55- B	oring - 3	arsen	c	2-4'
		B4FD4		PCB	ļ	
		R55- Bo	ring - 3	Grain	Size	
	Allia	nce Beau	h R.	ec = / f/		
		1	Boring-1	!		0-11
	<del></del>	B4FD		PCB		
		Alliance	Bacing-1	Grain.	Size	
	UNION	LAKE B	EACH	Rec. = 1	PJ	
	1250	ULB-BO	ring-1	arrenic		0-1'
		84F D6	-	PCB		
·		ULB- Ba	ring-1	Grains	(Ze	
-				<del> </del>	<del></del>	
•	<del></del>			<del></del>		· ·
-		<del>-</del>	,			
	1					(2)

13 SEP. 2007 West of Mill Rd. Rec. 2,9 A Hoalysis Interval Granic 1300 Mill-Boring B4FD7 Grain Size Mill - Borns-1 1-21 Hill-Boxingaiseni 1301 PCB BYFD8 21-2.91 Mill-Boring-3 B4FD9 arsenic 1302 PCB

### **APPENDIX E**

USEPA REGION 2 DESA - STANDARD OPERATING PROCEDURES C-109 AND C-116

SOP Number : C-116 Effective Date: 06/30/05 Revision #: 1.0 Page 1 of 13



#### STANDARD OPERATING PROCEDURE

# PREPARATION OF AQUEOUS, TCLP EXTRACTS, SOIL/SEDIMENT/SLUDGE/SOLID, WASTE OIL/ORGANIC SOLVENTS, AND BIOLOGICAL TISSUE MATRICES BY BLOCK DIGESTION

	Signature and Title	
Prepared by:	Renee Lettieri, Chemist, OICS	Date
Peer Reviewed by:	Linda Boyer, Env. Protection Specialist, OICS	Date
QA Reviewed by:	Sumy P. Cherukara, Quality Assurance Officer	Date
Approved by:		
Approved by:	John R. Bourbon, Chief, OICS	
	Deborah A. Szaro, Chief, Laboratory Branch  Annual Review	Date
Reviewed by:		
Reviewed by:	Signature	Date
-	Signature	Date

U.S. ENVIRONMENTAL PROTECTION AGENCY REGION 2 DIVISION OF ENVIRONMENTAL SCIENCE & ASSESSMENT LABORATORY BRANCH

SOP Number: C-116 Effective Date: 06/30/05 Revision #: 1.0 Page 2 of 13

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- 2. Summary of SOP
- 3. Definitions
- 4. Interferences
- 5. Safety
- 6. Apparatus and Materials
- 7. Reagents and Solutions
- 8. Sample Collection, Preservation, Storage and Holding Time
- 9. Sample Preparation
- 10. Instrument Operating Conditions
- 11. Sample Analysis
- 12. Data Analysis and Calculations
- 13. Method Performance
- 14. Quality Control
- 15. Reporting and Validation
- 16. Pollution Prevention
- 17. Waste Management
- 18. References

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#### STANDARD OPERATING PROCEDURE

#### PREPARATION OF AQUEOUS, TCLP EXTRACTS, SOIL/SEDIMENT/SLUDGE, WASTE OIL/ORGANIC SOLVENTS, AND BIOLOGICAL TISSUE MATRICES BY BLOCK DIGESTION

#### 1. Scope and Application

- 1.1 This method is used to digest by DigiBLOC, all environmental samples, with the exception of drinking water. These include aqueous, TCLP extracts, soil/sediment/sludge/solid, waste oil/organic solvent, and biological tissue. Samples are then analyzed using ICP-AES, SOP #C109 or ICP-MS, SOP # C-112.
- 1.2 This SOP is based on EPA Methods 200.2, Revision 8.8 and EPA Method 200.8, Revision 5.4.

#### 2. Summary of Method

- 2.1 Aqueous or Aqueous TCLP: A suitable aliquot (usually 50 mL) of a well mixed, aqueous or homogeneous extract sample is accurately measured into a DigiTUBE and heated on the DigiBLOC at 85° C with HNO<sub>3</sub> and HCL until the volume is reduced to 20mL. A watch glass is then placed on the tube and the sample is gently refluxed for an additional 30 minutes. After cooling, the sample is brought up to a known volume, capped and mixed. If needed, the digestates may be filtered.
- 2.2 Soil/Sediment/Sludge/Solid: Samples may be dried for a minimum of 12 hours at 60°C, ground well and mixed thoroughly or the drying step may be eliminated by digesting the samples as they are received. A correction factor derived from a Percent Solids determination is applied to the final result for either method. An aliquot is accurately weighed into a DigiTUBE and digested with HNO<sub>3</sub> and HCL at 95° C for 30 minutes. After cooling, the sample is filtered and brought up to a known volume, capped and mixed.
- 2.3 For biological tissue digestion, the sample is accurately weighed into a DigiTUBE and digested with HNO<sub>3</sub> and 30% H<sub>2</sub>O<sub>2</sub>.
- 2.4 Samples are then analyzed using ICP-AES or ICP-MS. In all instances, great care must be exercised to avoid contamination.

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#### 3. Definitions

See SOP#G-15 for definitions.

#### 4. Interferences

Samples must be well mixed and as homogenous as possible. Soil/Sediments/Sludges/Solids must be reduced to as small a particle size as practicable.

#### 5. Safety

5.1 The toxicity or carcinogenicity of each reagent used in this method has not been fully established. Each chemical should be regarded as a potential health hazard and exposure should be kept to an absolute minimum by following the appropriate standard safety procedures, e.g. wear proper protective equipment, gloves, lab coat, and working inside hoods whenever possible. Refer to Edison Facility Safety Manual Region II, Part 2 - Laboratory Safety for specific guidelines.

#### 5.2 Safety guidelines for the DigiBLOC

- 5.2.1 The DigiBLOC must be grounded and have a clearance of 3 inches on all sides. It must be located in an operable fume hood if the DigiVAC is not available. Do not mount DigiBLOC on a surface of flammable material.
- 5.2.2 The DigiBLOC must be lifted only from the bottom, not by the top white trim. Acquire assistance to move the unit.
- 5.2.3 Use caution when working around the instrument during operation. The unit has exposed hot surfaces.

#### 6. Apparatus and Materials

- 6.1 DigiBLOC Digestion System consisting of the Hot Block, with two 24 Position Racks with front and back airfoils,
- 6.2 DigiPROBE Sample Temperature Controller and probe.
- 6.3 DigiSET Sample Volume Controller and volume probe.
- 6.4 DigiVAC Exhaust System

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- 6.5 50 mL DigiTUBEs, screw caps and disposable ribbed watch glasses.
- 6.6 Top loading balance capable of measuring 0.01 gram, disposable spatulas and small weighing dishes for soil/sediment/sludge/solid digestion.
- 6.7 Porcelain evaporating dishes (195mL), pestles and glass stirring rods for soil/sediment/sludge/solid digestion.
- 6.8 Two re-pipettes capable of dispensing 0.25-5.0mL.
- 6.9 Two automatic pipettes (1-250uL & 1-1000uL).
- 6.10 Whatman #41 filter paper 125mm.
- 6.11 Disposable polypropylene funnels (65mm) and 100mL disposable beakers.
- 6.12 Filtration rack

#### 7. Reagents and Solutions

All reagents must be of high purity and suitable for trace metal analysis.

- 7.1 Concentrated Nitric Acid (HN0<sub>3</sub>)
- 7.2 Ultrex Concentrated Nitric Acid (HN0<sub>3</sub>)
- 7.3 Concentrated Hydrochloric Acid (37%)
- 7.4 Hydrogen Peroxide, 30%
- 7.5 Reagent Grade Water
- 7.6 SPEX CertiPrep Custom Claritas Standard High Check containing 250ug/mL (250ppm) of each of the following: Ag, Al, As, B, Ba Be, Ca, Cd, Co, Cr, Cu, Fe, K, Mg, Mn, Mo, Na, Ni, Pb, Se, Sb, Si, Sr, Ti, Tl, V, Zn, Sn or equivalent.
- 7.7 SPEX CertiPrep Custom Multi-element Standard ICV II containing 250mg/l (250 ppm) of each of the following: Al, Ca, Fe, Mg, K, Na, Si or equivalent.
- 7.8 Soil LCS Environmental Resource Associates: Trace Metals in Soil or equivalent.

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7.9 Biological Tissue LCS - DOLT-3 National Research Council, Canada, Tort-2 National Research Council, Canada, NIST 15666 - Oyster Tissue or other suitable material.

#### 8. Sample Collection, Preservation, Storage and Holding Time

- 8.1 Aqueous: Samples may be collected in plastic or glass. Samples must be preserved to a pH<2 using HN0<sub>3</sub>, may be stored at room temperature and should be digested and analyzed within 6 months of collection.
- 8.2 Soil/Sediment/Sludge/Solid: Samples may be collected in plastic or glass. Samples must be stored at 4° C and should be digested and analyzed within 6 months of collection unless stored at -20° C after air-drying.
- 8.3 Biological Tissue: Samples may be collected in plastic or glass containers and must be stored at -20°C.
- 8.4 Waste Oil/Organic Solvents: Samples do not require any preservation and are stored at room temperature.
- 8.5 Drum sample usually have no temperature or holding time requirements.

#### 9. Sample Preparation

- 9.1 Aqueous Sample Preparation
  - 9.1.1 Verify that the pH of the sample is <2 using pH test paper. Record in the Metals Sample pH log book. If the pH is >2, add concentrated HNO<sub>3</sub> until the pH is <2, then wait at least 16 hours before rechecking the pH and proceeding with the sample prep.
  - 9.1.2 Transfer 50mL (or other suitable aliquot) from a well mixed, acid preserved sample to a 50mL DigiTUBE. In addition, prepare a Prep Blank, two LCS's and 1 matrix spike for each project/matrix with at least one matrix spike per batch of 20 or fewer samples. Also prepare extra blank sample tubes that will hold the DigiPROBE and, if used, the DigiSET volume control probe. Refer to Section 14.1 of this SOP for QC procedure.
  - 9.1.3 Add 0.5 mL concentrated nitric acid and 0.25mL of concentrated hydrochloric acid to each tube.

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- 9.1.4 Insert the tubes into the DigiBLOC for solution evaporation at a pre-tuned temperature of 85 °C. If space permits, leave the outer rows empty. Position the extra blank samples under the exhaust hole of the DigiVAC. Carefully position the DigiPROBE in one tube and the DigiSET volume control set to 20mL in the other tube. If the DigiVAC is unavailable, carefully place the instrument in a hood and proceed. If the DigiSET is not used, monitor volume visually.
- 9.1.5 Close the DigiVAC lid and turn power on to the DigiVAC, DigiSET and DigiBLOC.
- 9.1.6 Reduce volume to approximately 20 mL by gently heating at 85°C then cap each tube with a disposable ribbed watch glass and reflux for 30 minutes.
- 9.1.7 Remove from DigiBLOC. Allow to cool. Filter, if necessary. (See Section 9.2.7) Dilute to 50 mL with Reagent Grade water, cap and mix well.

#### 9.2 Soil/ Sediment/Sludge/Solid Preparation

#### 9.2.1 Sample Drying

#### 9.2.1.2 Pre-Drying Method

This method works best for samples that have a high water content. Evaporating dishes and pestles must be rinsed with 10% HNO<sub>3</sub>.

Transfer the sample to a 195mL porcelain evaporating dish using a glass stirring rod or disposable spatula and dry at 60°C for a minimum of 12 hours. Cool, then grind with a pestle in the porcelain evaporating dish. Mix well, transfer to a plastic or glass container and store at 4°C until ready to digest.

These sediment samples require a % Solids determination. See the METALS % SOLIDS LOGBOOK for procedure. % Solids results are reported under SOLA in LIMS. This does not preclude an analysis request for % Solids.

#### 9 2 1 2 "As Received" Method

This method works best for samples that have a low water content. Samples are digested as received. A % Solids determination is performed using the procedure described in the METALS % SOLIDS LOGBOOK. % Solids results are reported under SOLA in LIMS. This does not preclude an analysis request

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for % Solids.

- 9.2.2 Weigh 0.5g or other suitable aliquot of well mixed sample into a 50 mL DigiTUBE. Use approximately 5mL of Reagent Grade water to rinse down the sides of the DigiTUBE. In addition, prepare a Prep Blank, two LCS's, 1 matrix spike for each project/matrix with at least one matrix spike per batch of 20 or fewer samples and an extra sample that will hold the DigiPROBE. Refer to Section 14.2 for QC prep.
- 9.2.3 Under a fume hood, add 5 mL Reagent Grade water, 1.0mL conc. HNO<sub>3</sub> and 1.0mL of conc. HCL to the tubes. Keep samples under the hood until any reaction subsides.
- 9.2.4 Insert the tubes into the DigiBLOC for digestion at a pre-tuned temperature of 95°C. If space permits, leave the outer rows empty. In the extra sample tube, using a disposable watch glass with a hole, carefully position the DigiPROBE. This digestion may also be carried out using the DigiVAC.
- 9.2.5 Place a disposable watch glass on each tube and turn on the power to the DigiBLOC. If being used, close the DigiVAC lid and turn the power on to the DigiVAC
- 9.2.6 Heat samples at 95°C for 30 minutes. The DigiBLOC takes about 30 minutes to heat up to temperature.
- 9.2.7 Filtration is required for soil/ sediment/sludge/solid samples. Label a duplicate set of DigiTUBEs. Rinse Whatman #41 filter paper in disposable funnels with approximately 10mL Reagent Grade water. Place rinsed funnels into duplicate DigiTUBEs and transfer corresponding sample. Rinse original tube several times with Reagent Grade water. Dilute to 50mL with Reagent Grade water, cap and mix well.

#### 9.3 Biological Tissue Digestion

- 9.3.1 Homogenize the samples. Store samples in the freezer if digestion is delayed, then defrost prior to preparation for digestion. All determinations, including the LCS and matrix spike must be done in triplicate. Refer to Section 14.3 of this SOP for QC prep.
- 9.3.2 Weigh 2.0 g finely ground and well mixed sample or 1.0 g LCS (Dolt-3, Tort-2 or NIST 15666 Oyster Tissue) into a 50 mL DigiBLOC tube. Record actual weights

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in Prep Book. Be careful not to let tissue stick to sides of tube - aim for the bottom of tube. Prepare three tubes for the Prep Blanks and weigh an additional tissue sample for the DigiPROBE.

- 9.3.3 Add 5 mL concentrated Ultrex HNO<sub>3</sub> and swirl to mix. Heat gently in the DigiBLOC (tuned to 95° C) with continued swirling. If samples begin to foam, remove from heat until foam subsides. Continue to heat/cool and swirl until samples no longer foam, then digest at 95° C until sample appears clear. A 2 g sample should be clear after about 15 minutes of digestion.
- 9.3.4 Foaming is a more serious problem with the LCS. Dolt –3 will foam copiously when warmed with HNO<sub>3</sub>. Extreme care is needed in swirling and gently heating until the LCS/HNO<sub>3</sub> mixture appears clear. Then allow to digest at 95°C in the DigiBLOC for an additional 15 minutes.
- 9.4.5 After digestion with HNO<sub>3</sub> is completed, add 0.5 mL 30% H<sub>2</sub>O<sub>2</sub> in 0.1 mL portions to each of the tubes, swirling and heating with each addition until any effervescence subsides. It then should be safe to add 0.5 mL portions of the H<sub>2</sub>O<sub>2</sub>, heating in between additions until the samples become totally clear. After samples are totally clear, add 1 mL additional H<sub>2</sub>O<sub>2</sub>, cover the tubes with a plastic watch glass and digest for 30 minutes more. Remove from the DigiBLOC, cool and dilute to 20 mL with Reagent Grade water. Cap securely and mix well.

#### 10. Instrument Operating Conditions

- 10.1 DigiBLOC set-up
  - 10.1.1 Power ON Power switch.
  - 10.1.2 Check Temperature Set-Point by pressing the star button (\*). Temperature should be set at 85° C for Aqueous and TCLP extracts, 95° C for soil/sediment/sludge/solid, waste oil/organic solvent and biological tissue. If the temperature must be changed or the DigiPROBE is either connected or disconnected, the instrument must be tuned.
    - 10.1.2.1 Set temperature by pressing and holding the star button (\*) while simultaneously pressing the ▲ (arrow up) or the ▼ (arrow down) button to obtain the desired temperature.
    - 10.1.2.2 Tune DigiBLOC as follows:

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- Set desired temperature.
- Hold the ▲(arrow up) and ▼(arrow down) keys simultaneously for ≈ 3 seconds to enter program mode. The display will show *tunE*.
- -While holding the star button(\*), hit the  $\triangle$ (arrow up) to reach AESP (the E is actually an upside down F) and then release the star button (\*).
- Press and hold buttons simultaneously for 3 seconds until the temperature appears. The system will flash between *tunE*, *AESP* and the current temperature.
- -When tuning is complete, the system will automatically turn *tunE* off and display the current temperature only.

#### 10.2 DigiBLOC Shut-down

- 10.2.1 Power OFF DigiBLOC and DigiVAC if used
- 10.2.2 Rinse DigiPROBE with Reagent Grade water and place in a clean empty tube.

#### 11. Sample Analysis

Actual sample analysis is carried out using methods SOP #C-109 Trace Metals in Aqueous, Soil/Sediment/Sludge/Solid, Waste Oil, Organic Solvent, Biological Tissue- ICP-AES or SOP #C-112 Trace Elements in Aqueous, Soil/Sediment/Sludge/Solid, Waste Oil, Organic Solvent and Biological Tissue by ICP-MS.

#### 12. Data Analysis and Calculations

Calculations are not done as part of this method. All weights and dilutions are recorded in the Metals Sample Prep Log Book

#### 13. Method Performance

Method performance is evaluated as part of methods SOP #C-109 Trace Metals in Aqueous, Soil/Sediment/Sludge/Solid, Waste Oil,Organic Solvent, Tissue - ICP-AES or SOP #C-112 Trace Elements in Aqueous, Soil/Sediment/Sludge/Solid, Waste Oil, Organic Solvent and Biological Tissue by ICP-MS.

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#### 14. Quality Control

#### 14.1 Aqueous Quality Control

- 14.1.1 One Prep Blank (PB) should be prepared for every batch of 20 or fewer samples. The PB is prepared by transferring 50mL of Reagent Grade water into a 50mL DigiTUBE and adding 0.5mL HNO<sub>3</sub> and 0.25mL HCl.
- 14.1.2 Two LCS's (Laboratory Control Samples) are prepared for every batch of 20 or fewer samples. These LCS's are made by pipetting 20uL of (CAL I) SPEX CertiPrep Custom Claritas Standard (250PPM) and 400uL of SPEX CertiPrep Custom Multi-element Standard ICV II (250PPM) into 50mL DigiTUBEs containing 50 mL Reagent Grade water, 0.5mL of concentrated HNO<sub>3</sub> and 0.25mL concentrated HCl.
- 14.1.3 One Matrix Spike (MS) is prepared for each matrix per project with at least one MS per batch of 20 or fewer samples. The MS is prepared by adding 20uL of (CAL I) SPEX CertiPrep Custom Claritas Standard (250PPM) and 400uL of SPEX CERTIPREP Custom Multi-element Standard ICV II (250PPM) to a DigiTUBE containing 50 mL of a duplicate environmental sample, 0.5mL of concentrated HNO<sub>3</sub> and 0.25mL concentrated HCl.

#### 14.2 Sediment Quality Control

- 14.2.1 One Prep Blank should be prepared for every batch of 20 or fewer samples. The PB is prepared by transferring 5mL of Reagent Grade water into a 50mL DigiTUBE and adding 1.0mL HNO<sub>3</sub> and 1.0mL HCl.
- 14.2.2 Two LCS's are prepared for every batch of 20 or fewer samples. These LCS's are made by weighing 0.5g of ERA's Trace Metals in Soil into a 50mL DigiTUBEs and adding 5mL of Reagent Grade water to wash down the sides of the tube, 1.0mL HNO<sub>3</sub> and 1.0mL HCl.
- 14.2.3 One Matrix Spike is prepared for each matrix per project with at least one MS per batch of 20 or fewer samples. The MS is prepared by adding 20uL of (CAL I) SPEX CertiPrep Custom Claritas Standard (250PPM) and 400uL of SPEX CERTIPREP Custom Multi-element Standard ICV II (250PPM) to a 50mL DigiTUBE containing 0.5g of a duplicate environmental sample, 5mL of Reagent Grade water, 1.0mL HNO<sub>3</sub> and 1.0mL HCl.

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#### 14.3 Biological Tissue Quality Control

- 14.3.1 Three Prep Blanks are prepared for every batch of 20 or fewer samples. The PB is prepared by transferring 5mL of conc. HNO<sub>3</sub> and 5 mL 30% H<sub>2</sub>O<sub>2</sub> into a 50mL DigiTUBE and digesting at 95°C for about 45 minutes, then diluting to 20mL with Reagent Grade water.
- 14.3.2 The LCS is prepared in triplicate for every batch of 20 or fewer samples. DOLT-3 is presently being used for the LCS, but Tort-2, NIST 15666 Oyster Tissue or other suitable material are also acceptable. Digest as directed in 7.4.
- 14.3.3 The Matrix Spike (MS) is prepared in triplicate for each matrix per project for each batch of 20 or fewer samples. The MS is prepared by adding 20uL of (CAL I) SPEX CertiPrep Custom Claritas Standard (250PPM) and 400uL of SPEX CertiPrep Custom Multi-element Standard ICV II (250PPM) to a DigiTUBE containing 2.0g of a duplicate biological tissue sample and digesting as directed in 7.4.

#### 15. Reporting and Validation

Copies of all Log Book entries (pH, Sample Preparation, Percent Solids) are included in the final data packages.

#### **16. Pollution Prevention**

- 16.1 Pollution prevention encompasses any technique that reduces or eliminates the quantity or toxicity of waste at the point of generation. Numerous opportunities for pollution prevention exist in laboratory operation. The USEPA has established a preferred hierarchy of environmental management techniques that places pollution prevention as the management option of first choice. Whenever feasible, laboratory personnel should use pollution prevention techniques to address their waste generation. When wastes cannot be feasibly reduced at the source, the USEPA recommends recycling as the next best option.
- 16.2 The quantity of chemicals purchased should be based on expected usage during its shelf life and disposal cost of unused material. Actual reagent preparation volumes should reflect anticipated usage and reagent stability.
- 16.3 For information about pollution prevention that may be applicable to laboratories, consult "Less is Better: Laboratory Chemical Management for Waste Reduction", available from the American Chemical Society's Department of Government

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Regulations and Science Policy, 115 16<sup>th</sup> Street N.W., Washington D.C 20036, (202)872-4477.

#### 17. Waste Management

The USEPA requires that laboratory waste management practice be conducted consistent with all applicable rules and regulations. Excess reagents, samples and method process wastes should be characterized and disposed of in an acceptable manner. The agency urges laboratories to protect the air, water and land by minimizing and controlling all releases from hoods and bench operations, complying with the letter and spirit of any water discharge permit and regulations, and by complying with all solid and hazardous waste regulations, particularly the hazardous waste identification rules and land disposal restrictions. For further information on waste management consult the Region 2 SOP #G-6, "Disposal of Samples and Hazardous Wastes".

#### 18. References

- 1. EPA Method 200.2, Revision 2.8.
- 2. SW846 3010A
- 3. Operation Manual for DigiBLOC 3000 Digestion System
- 4. DigiVAC Operation Manual

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#### STANDARD OPERATING PROCEDURE

# DETERMINATION OF METALS IN AQUEOUS, TCLP EXTRACTS, SOIL/SEDIMENT, SLUDGE, WASTE OIL/ORGANIC SOLVENTS, AND BIOLOGICAL TISSUE SAMPLES BY TRACE (AXIAL CONFIGURATION) INDUCTIVELY COUPLED PLASMA-ATOMIC EMISSION SPECTROMETRY

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U.S. ENVIRONMENTAL PROTECTION AGENCY REGION 2
DIVISION OF ENVIRONMENTAL SCIENCE AND ASSESSMENT LABORATORY BRANCH

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#### Appendices:

Appendix A: Data Work-Up

Table(s):

Table 1: Standard Solutions Preparation

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# DETERMINATION OF METALS IN AQUEOUS, TCLP EXTRACT, SOIL/SEDIMENT, SLUDGE, WASTE OIL/ORGANIC SOLVENTS, AND BIOLOGICAL TISSUE MATRICES BY TRACE (AXIAL CONFIGURATION) INDUCTIVELY COUPLED PLASMA-ATOMIC EMISSION SPECTROMETRY

#### 1. Scope and Application

1.1 This SOP is applicable to the preparation of environmental samples, including aqueous, TCLP extract, soil/sediment, biological tissue, and waste oil/organic solvents, for the determination of the following metals:

Ag, Al, As, B, Ba, Be, Ca, Cd, Co, Cu, Cr, Fe, K, Mg, Mn, Mo, Na, Ni, Pb, Se, Sb, Si, Sn, Sr, Ti, Tl, V, and Zn

Note - This SOP is not applicable to the preparation and analysis of drinking water compliance monitoring samples. The procedure for the preparation and analysis of drinking water compliance monitoring samples using the Trace ICAP is detailed in Laboratory SOP DW-5.

- 1.2 All analysts must satisfactorily perform an initial demonstration of capability (DOC) by meeting the method performance criteria in Sec. 13.1 prior to performing sample analysis using this SOP.
- 1.3 The standard reporting limits for both aqueous and non-aqueous samples are listed in Table 2.
- 1.4 This SOP is based on EPA Method 200.7, Revision 4.4.

#### 2. Summary of SOP

2.1 Environmental samples, e.g., aqueous, TCLP extracts, soil/sediment, sludges, waste oil/organic solvent, and biological tissue, are digested in a mixture of acids, according to the procedures described in U. S. Environmental Protection Agency, Region 2, SOP C-116 "Preparation of Aqueous, TCLP Extracts, Soil/Sediment/Sludge, Waste Oil/Organic Solvents, and Biological Tissue Matrices by Block Digestion."

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2.2 The analysis described in this method involves multi-element determinations by ICAP-AES using a simultaneous Thermo-Jarrell Ash Trace Purge ICAP. The instrument measures characteristic atomic-line emission spectra by optical spectrometry. Samples are nebulized and the resulting aerosol is transported to the plasma torch. Element specific emission spectra are produced be a radio-frequency inductively coupled argon plasma. The spectra are dispersed by a grating spectrometer, and the intensities of the line spectra are monitored at specific wavelengths by a photomultplier tube (PMT). Photocurrents from the pmt are processed and controlled by a computer system. A background correction technique is required to compensate for background contribution to the determination of the analytes. Background must be measured adjacent to the analyte wavelength during analysis. Various interferences must be considered and addressed appropriately.

#### 3. Definitions

See **SOP G-15** for definitions.

#### 4. Interferences

- 4.1 Several types of interference effects may contribute to inaccuracies in the determination of trace elements. They can be summarized as follows:
  - 4.1.1 Spectral Interferences - these interferences can be categorized as 1) overlap of a spectral line from another element; 2) unresolved overlap of molecular band spectra; 3) background contribution from continuous or recombination phenomena; and 4) background contribution from stray light from the line emission of high concentration elements. The first of these effects can be compensated for by utilizing a computer correction of the raw data, requiring the monitoring and measurement of the interfering element. The second effect may require selection of an alternate wavelength. The third and fourth effects can usually be compensated for by a background correction adjacent to the analyte line. In addition, users of simultaneous multi element instrumentation must assume the responsibility of verifying the absence of spectral interference from an element that could occur in a sample but for which there is no channel in the instrument array. For this purpose, linear relations between concentration and intensity for the analytes and the interferences must be demonstrated over the range of interest.
  - 4.1.2 Physical Interferences these interferences are generally considered to be effects associated with the sample nebulization and transport processes. Such properties

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as change in viscosity and surface tension can cause significant inaccuracies especially in samples which may contain high dissolved solids and/or acid concentrations. The use of a peristaltic pump may lessen these interferences. If these types of interferences are operative, they must be reduced by dilution of the sample. Another problem which can occur from high dissolved solids is salt buildup at the tip of the nebulizer. This affects aerosol flow-rate causing instrumental drift. Wetting the argon prior to nebulization, the use of a tip washer, or sample dilution have been used to control this problem. This problem can also be alleviated by using a Bergener nebulizer instead of a Meinhardt nebulizer. Also, it has been reported that better control of the argon flow rate improves instrument performance. This is accomplished with the use of mass flow controllers.

- 4.1.3 Chemical Interferences these interferences are characterized by molecular compound formation, ionization effects and solute vaporization effects. Normally these effects are not pronounced with the ICP technique, however, if observed they can be minimized by careful selection of operating conditions (that is, incident power, observation position, and so forth), buffering of the sample and matrix matching. These types of interferences can be highly dependent on matrix type and specific analyte element.
- 4.2 Generally, whenever a new or unusual sample matrix is encountered, a series of tests on the matrix-type are performed, e.g., background check of the sample, sample overlay with standards, etc., prior to analyzing samples associated with that matrix. If the problems associated with the new matrix cannot be overcome, the sample must either be diluted appropriately (and the Reporting Limit raised accordingly) or analyzed by an acceptable different method.

#### 5. Safety

The toxicity and carcinogenicity of each reagent used in this method has not been fully established. Each chemical should be regarded as a potential health hazard and exposure to these compounds should be minimized by good laboratory practices. Normally accepted laboratory safety practices should be followed during reagent preparation and instrument operation. Always wear safety glasses or full-face shield for eye protection when working with these reagents. Each laboratory is responsible for maintaining a current safety plan, a current awareness file of OSHA regulations regarding the safe handling of the chemicals specified in this SOP.

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#### 6. Apparatus and Materials

- 6.1. Inductively coupled argon plasma:
  - 6.1.1 Thermo Jarrell Ash (TJA) ICAP 61E Trace Purge Analyzer (with an axial torch) controlled by a computer.
  - 6.1.2 TJA radio-frequency generator.
  - 6.1.3 High purity (99.99%) liquid argon.
  - 6.1.4 A variable speed peristaltic pump which is used to deliver both standards and samples to the nebulizer.
  - 6.1.5 Computer controlled mass flow controllers which regulate the argon flow rates.
- 6.2. A balance which has the capability to measure 0.1mg.
- 6.3. Labware (See Section 6.10 of EPA Method 200.7 Rev. 4.4).

#### 7. Reagents and Solutions

- 7.1 Reagents All reagents must be of high purity and suitable for trace metals analysis.
  - 7.1.1 Hydrochloric acid, concentrated HCl (GFS HCl, 37% Reagent ACS or equivalent)
  - 7.1.2 Nitric acid, concentrated HNO<sub>3</sub> (GFS HNO<sub>3</sub>, Redistilled or equivalent)
  - 7.1.3 Reagent grade water ASTM Type I Water
- 7.2 Solutions Refer to Table 1 for standard solutions preparation summary.
  - 7.2.1 Calibration Stock Standard Solutions Claritas Custom Standards manufactured by Spex CertiPrep under UL ISO 9001 Quality Assurance Program.
    - 7.2.1.1 Calibration Standard 1 SPEX CertiPrep Custom Claritas Standard (250 ppm of Ag, As, B, Ba, Be, Cd, Co, Cr, Cu, Mn, Mo, Ni, Pb, Se, Sb, Sn, Sr, Ti, Tl, V, and Zn) or equivalent.
    - 7.2.1.2 Calibration Standard 2 SPEX CertiPrep Custom Claritas Standard (250 ppm of Al, Ca, Fe, Mg, K, Na and Si) or equivalent.
  - 7.2.2 Working Calibration Solution The Working Calibration Solution is prepared from the Stock Calibration Standard Solutions (7.2.1) to a final concentration of 1,000 ug/L for all elements except for Al, Ca, Fe, K, Mg, Na and Si which are 10,000 ug/L. The solution is in 2% HNO<sub>3</sub> and 5% HCl.

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- 7.2.3 Blanks Four types of blanks are required for the analysis. The (1) calibration blank is used in establishing the analytical curve, the (2) initial calibration blank/continuing calibration blank (ICB/CCB) run after the calibration check standards to assess carryover, (3) a rinse blank is used to flush the instrument uptake system and nebulizer between standards, check solutions, and samples to reduce memory interferences and (4) a Laboratory Reagent Blank/Prep Blank (LRB/PB) is used to assess possible contamination from the sample preparation procedure and to assess spectral background.
  - 7.2.3.1 The calibration blank is prepared by adding HNO<sub>3</sub> and HCl to reagent grade water to the same concentrations used for the calibration standard solution.
  - 7.2.3.2 The rinse blank is prepared by acidifying reagent grade water to the same concentration of the acids as used in the calibration blank.
  - 7.2.3.3 The ICB and CCB are prepared by acidifying reagent grade water to the same concentration of acids as used in the calibration blank
  - 7.2.3.4 Laboratory reagent blank (LRB)/Prep blank (PB) must contain all the reagents in the same volumes as used in digesting the samples. The LRB/PB must be carried through the same preparation scheme as the samples including digestion, if applicable.
- 7.2.4 Initial Calibration Verification/Continuing Calibration Verification Solution (ICV/CCV) These verification standard solutions are used to initially and periodically verify instrument performance during analysis. The ICV/CCV stocks must be obtained from a source different from the calibration stock standard solutions and prepared in the same acid mixture as the calibration standards. The concentration of the analytes in the ICV/CCV solution is 200 ug/L for all elements except Al, Ca, Fe, K, Mg, Na and Si which is 5,000 ug/L.
  - 7.2.4.1 Claritas Custom Standard ICV1, 250ppm (Ag, As, B, Ba, Be, Cd, Co, Cr, Cu, Mn, Mo, Ni, Pb, Se, Sb, Sn, Sr, Ti, Tl, V, and Zn)
  - 7.2.4.2 Claritas Custom Standard ICV2, 250ppm (Al, Ca, Fe, K, Mg, Na, Si)
- 7.2.5 Low Level Check (ICV/50, ICV/20 and ICV/10) The low level checks are used to initially and periodically verify instrument performance at lower concentration levels. The concentration of the analytes should be at or above the analyte reporting limit. Al, Ca, Fe, K, Mg, and Na are not evaluated in these low level check standards. The instrument performance low level checks are at 2%, 5% and 10% dilution of the ICV. The concentration of analytes of concern in ICV/50 are 4 ug/L for ICV1 elements and 100 ug/L for ICV2 elements; ICV/20 are 10 ug/L for the ICV1 elements and 250ug/L for ICV2 elements. The concentration

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of the analytes of concern in the ICV/10 are 20 ug/L for ICV1 and 500 ug/L for ICV2.

- 7.2.6 Internal Standard Solution (5ppm Y; Li 400ppm) The normal calibration procedure for arc/spark involve the use of an internal standard. An element not found in the matrix being analyzed is added to each standard and each sample. Should the volume of aspirated sample change a corresponding intensity change will occur for all elements. Since the ratio remains constant, the possible error is eliminated
- 7.2.7 Profiling Solution (5ppm As) External problems like temperature and humidity changes can cause short and long term drift. This drift is due to the expansion or contraction of the focal curve with the result that the analytical line moves in relation to the measuring device. Periodic profiling corrects for this drift.
- 7.2.8 Inter-Element Correction (IEC) Solution (previously known as Interferents Only Solution IOS) When inter-element corrections are applied, a spectral interference check solution is needed containing concentrations of the interfering elements at levels that will provide an adequate test of the correction factors.

#### 8. Sample Collection, Preservation, Storage and Holding Time

- 8.1 Sample Collection Samples must be collected in plastic or glass containers.
- 8.2 Preservation and Storage
  - 8.2.1 Aqueous samples the samples are preserved using concentrated HNO<sub>3</sub>. The preservation is performed either a) in the field at the time of collection, or b) in the Laboratory upon receipt (within one business day). If the samples are preserved in the Laboratory, the samples must be held for sixteen hours after acidification and then verified to a pH<2 prior to sample processing. If the sample pH is verified to be pH>2 after the sixteen hours, additional HNO<sub>3</sub> must be added and the sample held for an additional sixteen hours until verified to a pH<2. The samples are stored at room temperature.
  - 8.2.2 Soil/Sediment/Sludge samples these samples are preserved in a refrigerator at  $\leq 4^{\circ}$ C. Alternatively, the samples maybe stored at  $\leq -20^{\circ}$ C in a freezer.
  - 8.2.3 Biological Tissue samples The samples are stored at ≤-20°C in a freezer.
  - 8.2.4 Waste Oil/Organic Solvents these samples do not require any preservation. The

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samples are stored at room temperature.

8.2.5 Drum Samples - no temperature requirement for these samples.

#### 8.3 Holding time

- 8.3.1 Aqueous samples must be prepared and analyzed within six months of collection.
- 8.3.2 Soil/Sediment/Sludge samples must be digested and analyzed within six months of collection.

Note: If soil/sediment samples are stored at  $\leq$ -20°C, the holding time is extended. The samples must be prepared within 12 months of collection and analyzed within 6 months of digestion.

- 8.3.3 Biological Tissue samples must be digested within 12 months of collection and analyzed within 6 months of digestion.
- 8.3.4 Waste Oil/Solvent samples a holding time is not established for the digestion or analysis of these samples.
- 8..3.5 Drum Samples do not require any holding time.

#### 9. Sample Preparation

All Environmental samples, e.g., aqueous, soil/sediment, waste oil/organic solvent, and biological tissue, including NPDES wastewater compliance monitoring samples, are digested in a mixture of acids using the procedures described in SOP Number C-116 "Digestion of Metals Aqueous, TCLP Extracts, Soil/Sediment, Sludge, Waste Oil/Organic Solvents, TCLP Extracts and Biological Tissue Matrices by DigiBloc".

#### **10. Instrument Operating Conditions**

Before using this method, the following procedure is followed to optimize plasma conditions. The analyst should follow Thermo Electron's instructions unless other conditions provide better performance.

10.1 Before lighting the plasma, make sure the following settings are in place:

Auxiliary gas- low Nebulizer flow rate  $\sim 0.60 L/min$  Pump Rate  $\sim 140 rpm$ 

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Internal standard solution with a buffer

It is important to ensure that there is no pulsing in any of the lines.

- 10.2 After lighting the plasma, make sure the above settings have not changed and verify that the RF power setting is at 950 W.
- 10.3 Allow the plasma to become stable. At a minimum, wait 30 minutes before proceeding.
- 10.4 Optically profile the spectrometer by aspirating a 5ppm arsenic solution. The spectrum shifter must be between -0.05 and +0.05.

#### 11. Sample Analysis

- 11.1 Configure the instrument settings to those in Section 10.
- 11.2 Fill in the sample ID file.
- 11.3 After the plasma has become stable, standardize the instrument using the mixed calibration standard solution (Section 7.2.2) and the calibration blank (Section 7.2.3.1). The average of three readings is to be used. Flush the system with the rinse blank for a minimum of 60 seconds between each standard.
- 11.4 After the completion of the initial requirements, samples should be analyzed in the same operational manner used in the standardization routine with a rinse blank also being used between all sample solutions, LFBs/LCSs-Aqueous, LFMs/MSs, and check solutions.
- 11.5 During the analysis of samples, the laboratory must comply with the required quality control in Sections 14. Only for the "direct analysis" of drinking water is the sample digestion step of the LRB/PB, LFB/LCS-Aqueous, and LFM/MS not required.
- 11.6 Sample analysis consists of the following:

Calibration Blank

Mixed Standard

IPC/ICV

IPB/ICB (Calibration Blank solution)

Low Check Sample/ICV/50, ICV/20 and ICV/10 (either solution at or below the analyte MCL excluding Al, Fe, Mg, and Na)

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LRB/PB
LFBs/LCSs
Samples
LFM/MS
IPC/CCV -must be analyzed at a minimum of every 10 samples
IPB/CCB
ICV/50, ICV/20, ICV/10
SIC/IOS

Note: The IPC/CCV and IPB/CCB must be analyzed at a minimum of every 10 analyses and at the end of each analysis run.

- 11.7 Determined sample analyte concentrations that are 90% or more of the upper limit of the analyte LDR must be diluted with reagent grade water that has been acidified in the same manner as the calibration blank and reanalyzed. See Section 11.4.7. of EPA Method 200.7, Rev. 4.4.
- 11.8 Report Data as directed in Section 12.

#### 12. Data Analysis and Calculations

- 12.1 Refer to Appendix A for detailed instructions for data workup and/or upload into LabWorks.
- 12.2 Aqueous Samples Undigested

All dilution factors required as a result of dilutions made during analysis are applied at the instrument. Therefore, all of the aqueous sample results generated from the analysis (in ug/L) can be reported directly from the instrument. All results are reported to two significant figures and, in most cases, are reported using reporting limits listed in Table 2.

#### 12.3 Aqueous Samples - Digested

Dilution factors required as a result of dilutions made during analysis are also applied at the instrument. Therefore, all of the aqueous sample results, in ug/L, generated from the analysis can be reported directly from the instrument. All results are reported to two significant figures and, in most cases, are reported down to the standard reporting limit listed in Table 2.

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#### 12.4 Non-Aqueous Samples

All dilution factors required as a result of dilutions made during analysis are applied at the instrument. Therefore, all of the results, in ug/L, generated from the analysis can be used directly from the instrument. These "ug/L" results must then be converted to "mg/Kg" results. The ug/L result is multiplied by the final digestate volume in Liters, usually 0.050 L, and divided by the sample mass in grams, usually 0.50 g (the specific sample volume and mass are recorded in the metals sample preparation log book). For dry weight calculation, the mg/Kg results must be divided by the decimal version of the percent solids, e.g., 90% is 0.90. Refer to to SOP G-23 for Percent Dry Solids.

All mg/Kg results are reported to two significant figures and, in most cases, are reported using the reporting limits listed in Table 2, adjusted for percent solids correction for dry weight basis.

#### 13. Method Performance

An initial demonstration of capability (DOC) must be performed each time there is a significant change in the chemistry of the method, a major modification to an existing instrument, or a new instrument is installed. A DOC is performed by each analyst designated to analyze samples using this method. An annual check must subsequently be performed and documented for each analyst using this method.

#### 13.1 Accuracy and Precision

#### 13.1.1 Initial Demonstration of Capability

An initial demonstration of capability study must be conducted for this method for each analyst using this method. The study consisted of the analysis of four standards which are from a source independent of the standard curve. The results of the standards must be within the acceptance criteria supplied by the manufacturer or within 10% if none are specified. The % RSD should be within 20%. The results of the accuracy and precision study (true value, % recovery, standard deviation and % RSD) are maintained by the Quality Assurance Officer for each analyst and are located in the Central Branch File.

#### 13.1.2 Continuing Demonstration of Capability

An annual continuing demonstration of capability study must be performed and documented. It may consist of either successfully analyzing a PT sample or analyzing

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2 sets of AQC standards to within control limits as stated in section 13.1.1. The results of the continuing accuracy and precision study (true value, % recovery, standard deviation and % RSD or final report from the PT provider) are maintained by the Quality Assurance Officer for each analyst and are located in the Central Branch File.

#### 13.2 Method Detection Limit (MDL)

An MDL Study must be conducted for this method. The study is based on the requirements listed in 40 CFR Part 136 Appendix B. Specific procedures for conducting an MDL study can be found in SOP # G-8. The MDL Study comprised the analysis of seven reagent grade water samples fortified at a level between 2-3x the detection limit. The results of the MDL determination (true value, average concentration, standard deviation and calculated MDL) are maintained by the Quality Assurance Officer for each method and are located in the Central Branch File.

#### 13.3 Linear Dynamic Range (LDR)

The LDR must be determined by generating a normal linear calibration curve followed by the analysis of successively higher standard solutions. The results of these standard solutions are used to calculate % recovery. This is conducted until the % recovery fell below 90%. The last standard that had a % recovery of at least 90% is identified as the LDR limit

The results of the LDR Study are maintained in a file next to the instrument. The LDR results must be below or equal to that listed as the upper range in EPA Method 200.7.

#### 14. Quality Control

#### 14.1 Calibration Curve

Acceptance Criteria - A calibration blank and one mixed standard are used to standardize the instrument. After standardization, the ICV and ICB are used to determine acceptance.

Corrective Action - If the results of the ICV or ICB are unacceptable, analysis must be discontinued, the cause determined and/or in the case of drift the instrument re-calibrated.

#### 14.2 Initial Calibration Verification (ICV).

Acceptance Criteria - Analyze the ICV solution from a separate identifiable source (different lot number or vendor from that of calibration standards) immediately following

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the calibration. The result of the ICV solution must be within  $\pm 5\%$  of the true value for NPDES compliance monitoring samples and  $\pm 10\%$  for all other samples.

Corrective Action - If the calibration cannot be verified within the specified limits, reanalyze the ICV solution. If the results of the second analysis of the IPC/ICV solution is not within the acceptance limits for both types of samples (NPDES compliance monitoring samples and other samples), the analysis must be evaluated and the cause determined and the instrument re-calibrated. If the results of the second analysis of the IPC/ICV solution is not within the acceptance limits of NPDES requirements but within the acceptance limits for other program samples, a case narrative must be issued for samples that are non-compliant.

#### 14.3 Continuing Calibration Verification (CCV)

Acceptance Criteria - Analyze the CCV solution, from the same source as that used for the ICV, after a maximum of ten samples and at the end of the sample run. The results of each CCV solution must be within  $\pm 10\%$  of the true value for NPDES compliance monitoring samples ( $\pm 20\%$  for all other samples).

Corrective Action - If the calibration cannot be verified within the specified limits, reanalyze the CCV solution. If the results of the second analysis of the CCV solution is not within the acceptance limits, the analysis must be discontinued, the cause determined and the instrument re-calibrated. All samples following the last acceptable CCV solution must be reanalyzed.

#### 14.4 Initial Calibration Blank/Continuing Calibration Blank (ICB/CCB)

Acceptance Criteria - Analyze the calibration blank immediately following each calibration and after every CCV. All ICB/CCBs results must be < the |Reporting Limit|.

Corrective Action - If the result of the ICB/CCB is > |Reporting Limit|, the analysis should be stopped the problem identified, and the ICB/CCB reanalyzed. If the ICB/CCB results remain > |Reporting Limit|, the instrument must be recalibrated.

#### 14.5 Preparatory Blank (PB)/Laboratory Reagent Blank(LRB)

Acceptance Criteria - Analyze an LRB/PB per 20 samples or less per matrix. The PB/LRB results must be < the |Reporting Limit|.

Corrective Action - If the result of the PB/LRB is > |Reporting Limit|, then all associated samples with a concentration of  $\le 10x$  the amount found in the PB/LRB should be

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reprepared and reanalyzed. If the samples cannot be reprepared, then all affected sample results must be either: qualified accordingly, or the Reporting Limit is raised to the amount found in the sample. Check with the team leader/section chief to determine which option should be used.

Sample results  $\ge 10x$  the amount found in the PB/LRB are not considered to be affected by the blank contamination or drift, so no corrective action is needed.

14.6 Laboratory Fortified Blank (LFB)/Laboratory Control Samples (LCS)

#### 14.6.1 Aqueous LCS

Acceptance Criteria - Analyze two aqueous LFB/LCS samples with each batch of aqueous samples of 20 or less. Calculate accuracy as percent recovery using the following equation:

$$Recovery = \frac{Average \ of \ 2 \ LFB/LCS's}{s} \ X \ 100$$

where: LFB/LCS = laboratory fortified blank/laboratory control sample s = concentration of analytes added to fortify the LFB/LCS solution

The % recovery of the aqueous LFB/LCS must be within  $\pm 15\%$  of the true value for NPDES wastewater compliance monitoring samples and within  $\pm 20\%$  of the true value for all other environmental samples. The RPD of two LCSs should be <20%.

#### 14.6.2 Solid LCS

Acceptance Criteria - Analyze two solid LCS samples with each batch of solid samples of 20 or less. Calculate accuracy as percent recovery using the following equation:

The % recovery of the solid LCS must be within  $\pm 25\%$  of the true value or within the limits established by the vendor. The relative percent difference (RPD) of the duplicates should not exceed 25% for solid samples.

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Corrective Action for 14.6.1 and 14.6.2 - If the % recovery or %RPD results are outside the required control limits, the affected samples should be reprepared and reanalyzed. If the samples cannot be reprepared, then all affected sample results must be qualified accordingly.

#### 14.7 Laboratory Fortified Matrix (LFM)/Matrix Spike(MS) Recovery

Acceptance Criteria - Fortify a known amount of analytes to one sample per matrix per project per batch of 20. The LFM/MS aliquot must be a duplicate of the aliquot used for sample analysis. When possible, the concentration should be the same as that added to the aqueous LFB/LCS, but should not exceed the midpoint concentration of the calibration curve. Calculate the percent recovery, corrected for background concentration measured in the unfortified sample aliquot, and compare these values to the control limits to the designated matrices recovery ranges:  $\pm 20\%$  for aqueous samples;  $\pm 25\%$  for solid samples (soils, sediment, and NAPL); and  $\pm 50\%$  for sludge and biological tissue samples. Percent recovery is calculated using the following equation:

$$R = \frac{C_s - C}{s} \times 100$$

where:

R = percent recovery,

Cs = fortified sample concentration,

C = sample background concentration, and

s = conc. equivalent of metal added to sample.

Corrective Action - If % recovery of the MS is outside the required control limits, and the laboratory performance is shown to be in control, the recovery problem encountered is judged to be matrix related, not system related. The native sample result of the sample used to produce the MS must be qualified accordingly.

Note: The % recovery of the MS is not evaluated if the result of the unfortified sample concentration is ≥1.0x the level used to fortify the sample.

#### 14.8 Serial Dilution Test

Acceptance Criteria - Analyze a 20% dilution of the MS sample(s). The serial diluted sample result(s), adjusted for the dilution, should agree with the MS result(s) to within 20% RPD.

Corrective Action - If the % RPD is outside the required control limits, and the

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laboratory performance is shown to be in control, the precision problem encountered is judged to be matrix related, not system related, and the sample should be qualified accordingly.

#### 14.9 Low-Level Checks - (ICV/50, ICV/20 and ICV/10)

Acceptance Criteria - Analyze the ICV/50, ICV/20 and ICV/10 standards, from a separate identifiable source other than the calibration standards, immediately following the ICV and ICB. The ICV/50, ICV/20 and ICV/10 should also be analyzed after every CCV. The % recovery of the ICV/50, ICV//20 and ICV/10 must be within  $\pm 30\%$  of the true value for all analytes of interest.

Corrective Action - If the ICV/50, ICV/20 and ICV/10 cannot be verified within the specified limits, analysis must be evaluated, the cause determined and/or in the case of drift the instrument re-calibrated. If the ICV/50 and ICV/20 are not within the specified limits for the elements of interest but the ICV/10 is within the required limits, then the Reporting Limit is raised up to the ICV/10 level as long as the sample project requirement allows.

14.10 Spectral Interference Check (SIC)/Inter-Element Correction (IEC) Solution (formerly known as Interferents Only Solution - IOS)

Acceptance Criteria - All metal results (required by the project(s), except for Al, Fe, Ca, K, Mg, and Na, should be below the established Reporting Limits listed in Table 2

Corrective Action - If a required metal result is > the |Reporting Limit|, the individual interferent metals must be analyzed independently to assess which metal is causing the interference. Once identified, the appropriate inter-element correction factor(s) and background correction point(s) must be reviewed, and where appropriate, adjusted. After the adjustment is completed, the IEC standard must be re-analyzed. Once a successful IEC is analyzed, the analysis can commence.

A corrective action is not required if one of the following conditions are met:

- 1. If the metal that is "affected" by the interferent is not required for the project(s) in question;
- 2. If the concentration of the metal that is "affected" by the interferent is < the |Reporting Limit|;
- 3. If the concentration of the metal causing the interference in the "affected" environmental sample(s) is at a trace level, i.e., <10000 ug/L (the level used in our mixed calibration standard, section 7.2.1).

#### 14.11 Triplicate Integrations

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Acceptance Criteria - Each analysis consists of three separate integrations or readings. This includes the calibration standards, quality control samples and all associated environmental samples. The average of the three measurements is used for reporting results. The RSD must be  $\leq 20\%$  for all results that are  $\geq$  the reporting limit.

Corrective Action - If the RSD for a calibration standard, quality control sample and environmental sample is outside the control limits, the analysis must be repeated. If the RSD is still outside the control limits, the analysis must be terminated, and repeated after correcting the problem. If the RSD is still outside the control limits, and the laboratory performance, i. e. CCV, is shown to be in control, the RSD problem encountered is judged to be matrix related, not system related, and the sample should be qualified accordingly.

#### 15. Reporting and Validation

15.1 Reporting Limits - The reporting limits are calculated based on the concentration of the lowest calibration standard analyzed. The reporting limits are matrix and dilution dependent. All results are reported to 2 significant figures.

#### 15.2 Sample Data Package

The sample data package should include but not be limited to the following:

- ICAP-AES QA/QC Checklist with all relevant information entered;
- Copies of Log Book entries of Analysis Run Log; Sample Digestion Log, and if required, Sample Percent Solids Log and/or pH Log;
- Calibration Report;
- Summary Analysis Form;
- QC Summary Forms; and
- Instrument generated Sample Data
- 15.3 Laboratory Information Management System (LIMS) The analyst enters the data on the LIMS under the appropriate analytical codes.
- 15.4 Data Validation The data package is given to the reviewer. The review is done by a peer who was not involved in the analysis. Upon completion of the review, including validation of all the appropriate codes in the LIMS for the particular project(s), the data reviewer will sign and date the QA/QC Checklist.
- 15.5 Data Records All project records associated with the data package are filed under one designated project file. All other projects associated with the data package are

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referenced to this designated project file via a "cross reference form". The "cross reference form" is placed in each of the project files that were associated with the data package.

The data package is placed in the bin identified for the designated project file. The records for this designated project file are filed in our locked record cabinets once all data from the project, e.g., non-metal inorganic data, organic data, microbiology data, etc. has been reviewed by the appropriate staff.

#### 16. Pollution Prevention

- 16.1 Pollution prevention encompasses any technique that reduces or eliminates the quantity or toxicity of waste at the point of generation. Numerous opportunities for pollution prevention exist in laboratory operation. The EPA has established a preferred hierarchy of environmental management techniques that places pollution prevention as the management option of first choice. Whenever feasible, laboratory personnel should use pollution prevention techniques to address their waste generation. When wastes cannot be feasiblely reduced at the source, the Agency recommends recycling as the next best option.
- 16.2 The quantity of chemicals purchased should be based on expected usage during its shelf life and disposal cost of unused material. Actual reagent preparation volumes should reflect anticipated usage and reagent stability.
- 16.3 For information about pollution prevention that may be applicable to laboratories and research institutions, consult *Less is Better: Laboratory Chemical and Management for Waste Reduction*, available from the American Chemical Society's Department of Government Relations and Science Policy, 1155 16th Street N.W., Washington D.C. 20036, (202)872-4477.

#### 17. Waste Management

The USEPA requires that laboratory waste management practices be conducted consistent with all applicable rules and regulations. Excess reagents, samples and method process waste should be characterized and disposed of in an acceptable manner. The Agency urges laboratories to protect the air, water, and land by minimizing and controlling all releases from hoods and bench operations, complying with the letter and spirit of any sewer discharge permits and regulations, and by complying with all solid and hazardous waste regulations, particularly the hazardous waste identification rules and land disposal restrictions. For

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further information on waste management consult the Region 2 SOP G-6, "Disposal of Samples and Hazardous Wastes".

#### 18. REFERENCES

- U. S. Environmental Protection Agency. "Determination of Metals and Trace Elements in Water and Wastes by Inductively Coupled Plasma-Atomic Emission Spectrometry," Method 200.7, Revision 4.4, May 1994.
- U. S. Environmental Protection Agency, Region 2, SOP G-6 "Disposal of Samples and Hazardous Wastes."
- U. S. Environmental Protection Agency, Region 2, SOP G-8 "Laboratory Policy for the Determination of Method Detection Limits (MDLs)."
- U. S. Environmental Protection Agency, Region 2, SOP G-15 "Laboratory Definitions and Data Qualifiers."
- U. S. Environmental Protection Agency, Region 2, SOP G-23 "Percent Dry Solids."
- U. S. Environmental Protection Agency, Region 2, SOP C-116 "Preparation of Aqueous, TCLP Extracts, Soil/Sediment/Sludge, Waste Oil/Organic Solvents, and Biological Tissue Matrices by Block Digestion."
- Method 2340 B "*Hardness by Calculation*"\_Standard Methods for the Examination of Water and Wastewater, 20<sup>th</sup> Edition-1998.
- U. S. Environmental Protection Agency, Solid Waste 846, Test Methods for Evaluating Solid Waste, Physical/Chemical Methods, Method 6010C "Inductively Coupled Plasma Atomic Emission Spectrometry" Laboratory Manual, Revision 3, November 2000.

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#### APPENDIX A

#### DATA WORKUP

- Copy data database (e.g., 121604.dbf) and the sample data file (samples.dbf) to a rewritable CD. The file nomenclature is based on the date of analysis, e.g., 121604.dbf 12 is month of December, 16 is the date and 04 is the year 2004.
- Login to LABWORKS
- Select Results from the main menu
- Select Instrument Conversion
- Select file name by recalling the data file, e.g., 121604.dbf
- Click OK. After OK is clicked a file is created converting the samples to .grf files
- Upload the correct .grf file into each individual sample by selecting the sample number with the corresponding test codes in the project.
- Right click on the results cell
- Select Modify Results
- Select Load Results
- Select the correct file from L:\Labwork ES\LWDATA5\INTRFACE\TJA DBF drive
- Click OK
- Check results to ensure that they are correct
- Delete any test codes in the project LIMS codes that are not required
- Make sure the reporting limits reflect analytical and prep dilutions, if any.

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**Table 1. Standard Solutions Preparation:** 

Standard/Solution Name	Concentration Required
Std 1 - Calibration Blank/ ICB/Rinse Blank/CCB	Reagent grade water
Std 2	1000 ppb all elements except 10,000 ppb for Al, Ca, Fe, Mg, K, Na and Si
ICV/CCV	200 ppb for all elements except 5000 ppb Al, Ca, Fe, Mg, K, Na and Si
ICV/50	4 ppb all elements except 100 ppb for Al, Ca, Fe, Mg, K, Na and Si
ICV/20	10 ppb all elements except 250 ppb for Al, Ca, Fe, Mg, K, Na and Si
ICV/10	20 ppb all elements except 500 ppb for Al, Ca, Fe, Mg, K, Na and Si
IEC Solution (previously known as IOS)	300,000 ppb Al, Ca, Fe, Mg and 60,000 ppb Na
Profile Solution	5,000 ppb As
Internal Standard	2,000,000 ppb Li 10,000 ppb Y

Note : The diluent used in preparing all the above standard solutions must be reagent grade water with 2% HNO $_3$  and 5% HCl.

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Table 2. Reporting Limits - Aqueous and Soil/Sediment

Element	MDL, ug/L	Reporting Limit, ug/L	Reporting Limit, mg/Kg
Ag	1.6	6	0.6
Al	48.5	200	200
As	2.3	8	0.8
В	2.3	8	0.8
Ba	1.6	6	0.6
Be	1.5	5	0.5
Ca 396.8	51.1	1000	100
Ca 317.9	51.4	1000	100
Cd	1.3	4	0.4
Со	2.3	8	0.8
Cr	1.8	6	0.6
Cu	2.7	10	1
Fe 259.9	42.5	200	20
Fe 271.4	55	200	20
K	126.0	1000	100
Mg 285.2	47.0	1000	100
Mg 279.0	54.8	1000	100
Mn	1.4	5	0.5
Mo	2.1	8	0.8
Na	589.0	1000	100
Ni	1.4	5	0.5
Pb	2.1	7	0.7
Se	1.9	7	0.7
Sb	3.7	14	1.4
Si	81.4	300	30
Sn	2.5	9	0.9
Sr	1.7	6.0	0.6
Ti	2.2	8	0.8
Tl	5.5	20	2
V	2.7	10	1
Zn	2.2	8	0.8

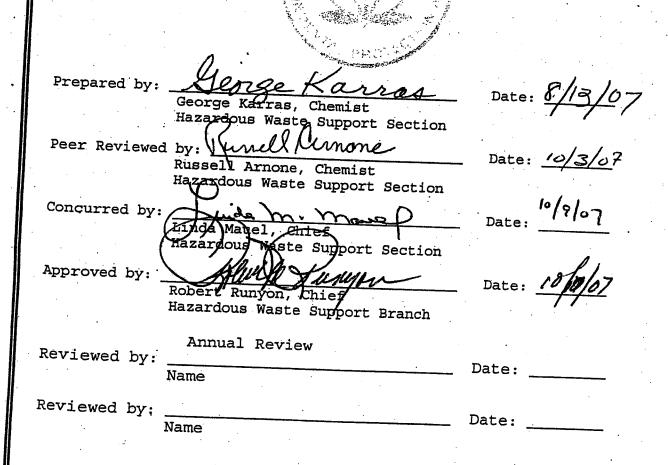
- Notes: 1) The IDL results were obtained using the analysis of seven ICV/40 standards analyzed on separate days
  - 2) The IDLs for all elements, except K and Si, were obtained by multiplying the standard deviation of the seven analysis by 3.14
  - 3) The Reporting Limits were obtained by multiplying the IDLs by 3.6 (1.2x3) and rounding to 2 significant figures

# **APPENDIX F**

# STANDARD OPERATING PROCEDURE HW-37/AROCLOR

SOP HW-37 Revision 1 August 2007

# SOP NO. HW-37/Aroclor Validation of Data USEPA Contract Laboratory Program Statement of Work for Organic Analysis of Low/Medium Concentration of Aroclor Organic Compounds SOM01.2



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#### INTRODUCTION

#### Scope and Applicability

This SOP offers detailed guidance in evaluating laboratory data generated according to the method in the "USEPA Contract Laboratory Program Statement of Work for Organics Analysis Multi-Media, Multi-Concentration, SOM01.2, February 2007". The validation procedures and actions discussed in this document are based on the requirements set forth in the "USEPA Contract Laboratory Program National Functional Guidelines for Superfund Organic Methods Data Review, July 2007". This document attempts to cover technical problems specific to low/Medium concentration of Aroclor compounds. Situations may arise where data limitations must be assessed based on the reviewer's own professional

In addition to technical requirements, contractual requirements may also be covered in this document. While it is important that instances of contract non-compliance be addressed in the Data Assessment, the technical criteria are always used to qualify the analytical data.

#### Summary

To ensure a thorough evaluation of each result in a data case, the reviewer must complete the checklist within this SOP, answering specific questions while performing the prescribed "ACTIONS" in each section. Qualifiers (or flags) are applied to questionable or unusable results as instructed. The data qualifiers discussed in this document are as follows:

#### Data Qualifiers

- U The analyte was analyzed for, but was not detected above the reported sample quantitation limit.
- J The analyte was positively identified; the associated numerical value is the approximate concentration of the analyte in the sample.
- N The analysis indicates the presence of an analyte for which there is presumptive evidence to make a "tentative identification."
- JN The analysis indicates the presence of an analyte that has been "tentatively identified" and the associated numerical value represents its approximate concentration.

- UJ The analyte was not detected above the reported sample quantitation limit. However, the reported quantitation limit is approximate and may or may not represent the actual limit of quantitation necessary to accurately and precisely measure the analyte in the sample.
- R The sample results are rejected due to serious deficiencies in the ability to analyze the sample and meet quality control criteria. The presence or absence of the analyte cannot be verified.

#### Lab Qualifiers:

- D The positive value is the result of an analysis at a secondary dilution factor.
- B The analyte is present in the associated method blank as well as in the sample. This qualifier has a different meaning when validating inorganic data.
- E The concentration of this analyte exceeds the calibration range of the instrument.
- P Pesticide/Aroclor target analytes when the % Difference between the analyte concentrations obtained from the two dissimilar GC columns is greater than 25%.

The reviewer must prepare a detailed data assessment to be submitted along with the completed SOP checklist. The Data Assessment must list all data qualifications, reasons for qualifications, instances of missing data and contract non-compliance.

#### Reviewer Qualifications:

Data reviewers must possess a working knowledge of the USEPA Statement of Work SOM01.2 and National Functional Guidelines mentioned above.

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YES NO N/A

PACKAGE COMPLETENESS AND DELIVERABLE	
CASE NUMBER: 36799 LAB: Data Chem	Laboratories. Inc
SITE NAME: Vineland Chemical SDG No(s) .: B41	= 1
1.0 Chain of Custody and Sampling Trip Reports	12/12/57
1.1 Are the Traffic Reports/Chain-of-Custody Reco present for all samples?	ords
ACTION: If no, contact RSCC, or the TOPO to obtain replacement of missing or illegible copies from the lab.	
1.2 Is the Sampling Trip Report present for all samples?	<u> </u>
ACTION: If no, contact either RSCC or ask the TOPO obtain the necessary information from the p contractor.	to rime
2.0 Data Completeness and Deliverables	
2.1 Have any missing deliverables been received and added to the data package?	IN
ACTION: Contact the TOPO to obtain an explanation of resubmittal of any missing deliverables from If lab cannot provide them, note the effect review of the data package in the Contract Problems/Non-compliance section of the Data Assessment.	n the lab
2.2 Was SMO/CLASS CCS checklist included with the package?	14

Date: August 2007 USEPA Region II SOP HW-37/Aroclor, Revision 1 Method: CLP/SOW, SOM01.2/Aroclor YES NO N/A Are there any discrepancies between the Traffic 2.3 Reports/Chain-of-Custody Records, and Sampling Trip Report? ACTION: If yes, contact the TOPO to obtain an explanation or resubmittal of any missing deliverables from the laboratory. 3.0 Cover Letter SDG Narrative Is the SDG Narrative or Cover Letter Present? 3.1 Are case number, SDG number and contract number 3.2 contained in the SDG Narrative or cover letter (see SOW, Exhibit B, section 2.5.1)? EPA sample numbers in the SDG, detailed documentation of any quality control, sample, shipment, and/or analytical problems encountered in processing the samples? Corrective action taken? Does the Narrative contain the following 3.3 information SOM01.1, page B-12, section 2.5.1)? column used, storage of samples, case#, SDG#, analytical problems, and discrepancies between field and lab weights. Did the contractor record the temperature of the 3.5 cooler on the Form DC-1, Item 9 - Cooler Temperature, and in the SDG Narrative? Does the Case Narrative contain the "verbatim" statement (page B-12, section 2.5.1 of the SOM)? If "No", to any question in this section, ACTION: contact the TOPO to obtain necessary If unavailable, document resubmittals. under the Contract Problems/ Non-Compliance section of the Data Assessment.

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YES NO N/A

# 4.0 Data Validation Checklist

- 4.1 Check the package for the following (see SOM reporting requirements, section 2.1, page B-10):
  - a. Is the package paginated in ascending order starting from the SDG narrative?
  - b. Are all forms and copies legible?
  - c. Assembled in the order set forth in the SOW?
  - d. All Aroclor Data present?

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# PART A: Low/Medium Aroclor Analyses

# 1.0 Sample Conditions/Problems

Do the Traffic Reports/Chain-of-Custody Records, Sampling Trip Report or Lab Narrative indicate any problems with sample receipt, condition of samples, analytical problems or special circumstances affecting the quality of the data?

19

ACTION: If samples were not iced or the ice was melted upon arrival at the laboratory and the temperature of the cooler was > 10° C, then flag all positive results with a "J" and all non-detects "UJ".

#### 2.0 Holding Times

2.1 Have any Aroclor technical holding times, determined from date of collection to date of analysis, been exceeded?

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2.2 Preservation: <u>Aqueous</u> and <u>Non-aqueous</u> samples must be cooled at 4°C ± 2°C.

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YES NO N/A

ACTION: Qualify sample results according to the following table.

# Holding Time Actions for Low/Medium Aroclor Analyses

			Act:	lon
Matrix Preserved		Criteria	Detected Associated Compounds	Non-Detected Associated Compounds
	. No	<pre>&lt; 7 days (extraction) &lt; 40 days (analysis)</pre>	J*	UJ*
Aqueous	No	> 7 days (extraction) > 40 days (analysis)	J	IJ
	Yes	<pre>≤ 7 days (extraction) ≤ 40 days (analysis)</pre>	No qualification	
	Yes	> 7 days (extraction) > 40 days (analysis)	J	ບປ
	Yes/No	> 28 Days (extraction)	J	R
	No	<pre>≤ 14 days (extraction) ≤ 40 days (analysis)</pre>	J*	UJ*
Non-aqueous	No	> 14 days (extraction) > 40 days (analysis)	J	บั
Yes		<pre>≤ 14 days (extraction) ≤ 40 days (analysis)</pre>	No quali	fication
	Yes	> 14 days (extraction) > 40 days (analysis)	J	ບປ
	Yes/No	> 28 Days (extraction)	J	R

<sup>\*</sup> Only if cooler temperature exceeds 10°C (see ACTION in Section 1.1 above). No action required if temperature  $\leq$  10°C.

# 3.0 Surrogate Recovery (Form II ARO-1, Form II ARO-2, Form VIII ARO)

3.1 Are the Aroclor Recovery Summary Forms present?

ACTION: Contact the TOPO to obtain an explanation/resubmittal from the lab. If missing deliverables are unavailable, document the effect in the Data Assessment.

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YES NO N/A

3.2 Were the two surrogates, tetrachloro-m-xylene (TCX) and decachlorobiphenyl (DCB) added to all samples, MS/MSD, LCS, blanks including standards?

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ACTION: If no, use professional judgment in qualifying data as missing surrogate analyte may not directly apply to target analytes.

3.3 Were outliers marked with an asterisk on Form II?

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ACTION: Circle all outliers with a red pencil.

If yes, were effected samples re-analyzed?

The RTs of the surrogates in each mid-point Aroclor standards used for continuing calibration verification, all samples, including MS/MSD, LCS and all blanks must be within the calculated RT window. TCX must be within  $\pm$  0.05 minutes and DCB must be within  $\pm$  0.10 minutes of the mean retention time (RT) determined from the initial calibration and tabulated in Form VIII Pest.

Were any outliers marked with an asterisk on Form VIII ARO?

ACTION: Circle all outliers with a red <u>pencil</u>. If any Surrogate is outside the required limits, qualify their associated target

compounds (See Table below) as follows:

# Surrogate Compound Recovery Action for Aroclors

	Action		
Criteria	Detected Target Compounds	Non-Detected Target Compounds	
%R > 200%	J	No qualification	
150% < %R < 200%	J	No qualification	
30% ≤ %R ≤ 150%	No quali	<del> </del>	
10% ≤ %R < 30%	J ·	UJ	
%R < 10% (sample dilution not a factor)	J	R	
%R < 10% (sample dilution is a factor)	J	Use Professional Judgement	
RT out of RT window	Use professional judgment		
RT within RT window	No qualification		

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YES NO N/A

#### Note: Blank analysis having surrogates out of specification:

The reviewer must give special consideration to the validity of associated samples. Basic concern is whether the blank problems represent an isolated problem with the blank alone or whether there is a fundamental problem with the analytical process. For example, if one or more samples in the batch show acceptable surrogate recoveries, the reviewer may choose to consider the blank problem to be an isolated occurrence.

ACTION: Note in the Data Assessment under Contract Problems/ Non-Compliance if the Lab did not perform reanalysis and reviewer's judgment regarding blank problem.

3.5 Are there any transcription/calculation errors between raw data and Form IIs?

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ACTION:

If large errors exist, ask the TOPO to obtain an explanation/resubmittal from the lab, make any necessary corrections and note errors in the data assessment.

#### 4.0 Matrix Spike/Matrix Spike Duplicate Recovery (Form III)

Note: Data for MS/MSD will not be present unless requested.

- 4.1 Are the MS/MSD Recovery Forms (Form III ARO) present?
- 4.2 Was the MS/MSD analyzed at the required frequency (once per SDG, or every 20 samples, whichever is more frequent)?

ACTION: If any MS/MSD data are missing, take action as specified in section 3.1 above.

ACTION: No action is taken on MS/MSD data <u>alone</u>. However, using professional judgement, the validator may use the MS and MSD results in conjunction with other QC criteria and determine the need for some qualification of the data. If Any MS/MSD % recovery or RPD is out of specification, qualify data to include the consideration of the existence of interference in the raw data. Consideration include, but not limited to the following "Action":

#### Matrix Spike/Matrix Spike Duplicate Action for Aroclor

	Action	
Criteria	Detected Spike Compounds	Non-detected Spike Compounds
%R or RPD > Upper Acceptance Limit	J	No qualification
20% ≤ %R < Lower Acceptance Limit	J	UJ

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YES NO N/A

%R < 20%	J	Use professioan judgement
Lower Acceptance Limit ≤ %R; RPD ≤ Upper Acceptance Limit	No	qualification

Note: If it can be determined that the results of the MS/MSD affects only the sample spiked, limit qualification to only this sample. However, use professional judgment when it is determined through the MS/MSD results that the laboratory is having systematic problem in the analysis of one or more analytes that affect all associated samples.

#### 5.0 Blanks (Form IV)

5.1	Is the Ar	roclor Method	d Blank Summary	(Form IV ARO)	present
	for aqueo	ous and soil	samples?	·	

Frequency of Analysis: For the analysis of AROCLOR, has a method blank been analyzed for each SDG or every 20 samples, whichever is more frequent?

ACTION: If any blank data are missing, take action as specified above in section 3.1. If blank data is not available, reject "R" all associated positive data. However, using professional judgement, the data reviewer may substitute field blank data for missing method blank data.

A separate Form IV should be present if part of an extraction batch required sulfur removal. In such cases some samples will be listed on two blank summary forms once under the method blank, and once under the sulfur clean-up blank (PCBLK). Was this additional blank raw data and Form IV submitted when required?

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ACTION: If Form IV sulfur clean-up blank is missing, take action as specified in section 3.1 above.

5.4 Has a Aroclor instrument blank been analyzed at the beginning of every 12 hr. period following the initial calibration sequence (minimum contract requirement)?

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ACTION: If any blank data are missing, take action specified in Section 3.1.

5.5 Was the correct identification scheme used for all Aroclor blanks? (See page B-39, section 3.3.7.3 of SOM01.1 for further information)

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ACTION: Contact the TOPO to obtain resubmittals or make the required corrections on the forms.

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YES NO N/A

Document in the Data Assessment under Contract Problems/Non-Compliance all corrections made by the validator.

5.6 <u>Chromatography</u>: Review the blank raw data chromatogram, quant. Reports and data system printout. Is the chromatographic performance (baseline stability) acceptable for each instrument?



ACTION: Use professional judgement to determine the effect on the data.

5.7 Are all detected hits for target compounds in method, and field blanks less than the CRQL?



ACTION: IF no, an explanation and laboratory's corrective actions must be addressed in the case SDG narrative. Contact TOPO to request from Lab. revised narrative and make a note in the Contract Problems/Non-Compliance section of the Data Assessment.

#### 6.0 Contamination

NOTE: "Water blanks", "drill blanks", and distilled water blanks" are validated like any other sample, and are <u>not</u> used to qualify data.

Do not confuse them with the other QC blanks discussed below.

6.1 Do any method/reagent or cleanup blanks contain positive hits for target Aroclor compounds with values greater than the CRQL for that analyte?



Note: The concentration of each target compound in the instrument blank must be less than the CRQL for that analyte.

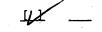
ACTION: Make note in data assessment under Contract Problems/Non-Compliance if any blank contains hit above the CRQLs.

6.2 Do any instrument blanks contain positive Aroclor results with values greater than CRQLs?



ACTION: Take the action specified in section 6.1.

6.3 Do any field/rinse blanks have positive Aroclor results?



NOTE: All field blank results associated with a particular group of samples (may exceed one per case) must be used to qualify data. Blanks may not be qualified because of contamination in another blank. Field blanks must be qualified for system monitoring compound, instrument performance criteria, spectral or calibration QC problems.

ACTION: Follow the directions in the table below to qualify results due to contamination. Use the largest value from all the associated

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YES NO N/A

blanks. If any blanks are grossly contaminated, all associated sample data should be qualified unusable (R).

#### Blank Action for Aroclor Analyses

Blank Type	Blank Result	Sample Result	Action for Samples
	Detects	Not detected	No qualification required
	< CRQL	< CRQL	Report CRQL value with a U
		≥ CRQL	No qualification required
	= CRQL	< CRQL	Report CRQL value with a U
Method, Field,		≥ CRQL	No qualification required
Sulfur Cleanup,		< CRQL	Report CRQL value with a U
Instrument	> CRQL	≥ CRQL and < blank contamination	Report concentration of sample with a U
en en en en en en en en en en en en en e		≥ CRQL and ≥ blank contamination	No qualification required
	Gross contamination	Detects	Qualify results as unusable R

NOTE: Analytes qualified "U" for blank contamination are treated as "hits" when qualifying for calibration criteria.

Note: When applied as described in the table above, the contaminant concentration in the blank are multiplied by the sample dilution factor.

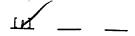
6.4 Are there field/rinse/equipment blanks associated with every sample?

ACTION: Note in data assessment if there's no associated field/rinse/equipment blank.

Exception: samples taken from a drinking water tap do not have associated field blanks.

# 7.0 Aroclor Initial and Continuing Calibration

- 7.1 Are the following Forms, chromatograms and data system printouts present?
  - a.) Form VI ARO-1/Aroclor Initial Calibration (Multipoint)



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	YES NO N	/A
b.) Form VI ARO-2/Aroclor Initial Calibration (Multipoint)		
c.) Form VI ARO-3/Aroclor Initial Calibration (Singlepoint)	[M _	
d.) Form VII ARO/Aroclor Calibration Verification	14	
e.) Form VIII ARO/Aroclor Analytical Sequence	т _	
f.) Form X ARO/Identification Summary for Multicomponent Analysis	тд —	

#### 7.2 Initial Calibration

7.2.1 Was the following contract required initial calibration sequence provided by the laboratory?

	Initial Calibration Sequence
1.	Aroclor 1221 CS3 (400ng/ml)
2.	Aroclor 1232 CS3 (400 ng/ml)
3.	Aroclor 1242 CS3 (400 ng/ml)
4.	Aroclor 1248 CS3 (400 ng/ml)
5.	Aroclor 1254 CS3 (400 ng/ml)
6.	Aroclor 1262 CS3 (400 ng/ml)
7.	Aroclor 1268 CS3 (400 ng/ml)
8.	Aroclor1016/1260 (100 ng/ml) CS1
9.	Aroclor1016/1260 (200 ng/ml) CS1
10.	Aroclor1016/1260 (400 ng/ml) CS1
11.	Aroclor1016/1260 (800 ng/ml) CS1
12.	Aroclor1016/1260 (1600 ng/ml) CS1
13.	Instrument Blank

ACTION: If initial calibration is not performed or not performed in the proper sequence, notify the TOPO and make a note in the data assessment.

7.3 Are there any transcription/calculation errors between raw data and the Forms? \_ m \_

ACTION: If large errors exist, take action specified in section 3.1 above.

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YES NO N/A

#### 7.4 Mean Retention Time (RT) and RT Window

Were the following mean RT and RT window met:

- 14 \_\_\_
- a.) The mean RT of each of the three to five major peaks were determined from the five-point initial calibration for all Aroclors
- b.) RT window was calculated as  $\pm$  0.07 for each of the three to five major peaks and  $\pm$  0.05 and  $\pm$  0.10 for the surrogates tetrachloro-m-xylene and decachlorobiphenyl, respectively.

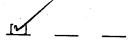
ACTION: If no, follow the action as specified in section 3.1.

7.5 Was at least one chromatogram from each of the Aroclor standards yield peaks that give deflection between 50-100% of full scale?

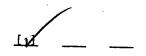


ACTION: IF no, take action as specified in section 3.1.

7.6 Was the mean Calibration Factor (CF) calculated for the three to five major peaks of each Aroclor, as well as for the surrogates, over the initial calibration range?



7.7 Were the Percent Relative Standard Deviation (%RSD) of the Calibration Factor for the three to five major peaks < 20% of each of the Aroclor compounds and surrogates?



ACTION: If no, take action as specified in the following Table.

# Initial Calibration Action for Aroclor Analyses

	Action	
Criteria	Detected Associated Compounds	Non-Detected Associated Compounds
Initial calibration is not performed or not performed in proper sequence	Use Professional Judgment and notify Contract Lab Program (CLP) Project Officer	
%RSD exceeds allowable limits *	J	UJ UJ
%RSD within allowable limits *	No qualification	

<sup>\* %</sup>RSD < 20.0% for Aroclors and surrogates (tetrachloro-m-xylene and decachlorobiphenyl.

# 7.8 <u>Continuing Calibration Verification (CCV) (Form VII)</u>

Were the Absolute Retention Time (RT) for each Aroclor and surrogate in the mid-point concentration (CS3) of

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YES NO N/A

the Standard used for CCV must be within the RT window determined from the initial calibration?

- 7.9 For opening CCV, or closing CCV that is used as an opening CCV for the next 12-hour period, the Percent Difference (%D) between the CF of each of the three to five peaks used to identify an Aroclor and surrogates in the mid-point concentration (CS3) of the Aroclor standards and the CF from the initial calibration must be within ±15.0%.
- 7.10 For a closing CCV, the %D between the CF of each of the three to five peaks used to identify an Aroclor and surrogates in the mid-point concentration (CS3) of the Aroclor standards and the CF from the initial calibration must be within ±50.0%.
- 7.11 No more than 14 hours may elapse from the injection of the instrument Blank that begins an analytical sequence (opening CCV) and the injection of the last mid-point concentration (CS3) of the Aroclor standards that ends an analytical sequence (closing CCV).
- 7.12 No more than 12 hours may elapse from the injection of the instrument blank that begins an analytical sequence (opening CCV and the injection of the last sample or blank that is part of the same analytical sequence.

Were sections 7.8 to 7.12 met?

ACTION: If no, use the following table to qualify Aroclor data:

## Continuing Calibration Verification (CCV) Action for Aroclor Analyses

	Action			
Criteria	Detected Associated Compounds	Non-Detected Associated Compounds		
RT out of RT Window	Use professional	l Judgment *		
Percent Difference not within limits $\pm$ 15% as specified in section 7.9 above	J	บัง		
Percent Difference not within limits $\pm$ 50% as specified in section 7.10 above	J	IJ		
Time elapsed is greater than acceptable limits as specified in section 7.11 & 7.12 above		R		
Percent Difference, time elapsed and RT are within acceptable limits	No qualification			

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YES NO N/A

\* For non-detected target compounds in the affected samples, check to see if the sample chromatogram contain any peak that are close to the expected RT window of the Aroclor of interest.

If no peaks are present, consider the non-detected values to be valid and no qualification of the data is necessary.

If any peaks are present close to the expected RT window of the Aroclor of interest, qualify the non-detected values as presumptively present "N".

For <u>detected compounds</u> in the affected samples, if the peaks are within the RT window, no qualification of the data is necessary. If the peaks are close to the expected RT window of the Aroclors of interest, the reviewer may take additional effort to determine if sample peaks represent the compound of interest.

For example, the reviewer can examine the data package for the presence of three or more standards containing the Aroclor of interest that were run within the analytical sequence during which the sample was analyzed. If three or more such standards are present, the RT window can be re-evaluated using the mean RT of the standards.

If the peaks in the affected sample fall within the revised window, qualify the detected Aroclor as "JN".

If the reviewer cannot do anything with the data to resolve the problem of concern, qualify all non-detects as unuseable "R".

## 8.0 Analytical Sequence Check (Form VIII-ARO)

8.1 Is Form VIII-Pest present and complete for each column and each period of analyses?

ACTION: If no, take action as specified in section 3.1

8.2 Was the proper analytical sequence followed for each initial calibration and subsequent analyses, and all standards analyzed at the required frequency for each GC/ECD instrument used?

ACTION: If no, use professional judgment to determine the severity of the effect on the data and qualify accordingly. Generally, the effect is negligible unless the sequence was grossly altered and/or the calibration was out of QC limits.

8.3 Are the surrogate retention time (RT) from the initial calibration for TCX and DCB provided on Form VIII-Pest?

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YES NO N/A

ACTION: If no, take action as specified in section 3.1

Was the asterisk (\*) applied to the RT of any blanks, samples, standards, MS/MSD, and LCS that did not meet the QC Limits of ± 0.05 minutes for TCX (tetrachloro-m-xylene) and ± 0.10 minutes for DCB (decachlorobiphenyl)?

ACTION: If any data are missing, take action specified in 3.1 above.

If no, use professional judgment to determine the severity of the effect on the data and qualify accordingly. Document in the data assessment under Contract Problems/Non-Compliance.

# 9.0 Sulfuric Acid and Gel Permeation Chromatography (GPC) Cleanup Procedures

9.1 Was sulfuric acid added to all extracts?

Note: Sulfuric acid cleanup is mandatory for all extracts

ACTION: If no, take action specified in section 3.1

9.2 <u>Gel Permeation Chromatography (GPC</u>

GPC is an optional cleanup procedure for both aqueous and non-aqueous samples that contain high molecular weight compounds that interfere with Aroclor analysis.

- 9.3 If GPC cleanup was performed on samples, GPC calibration is acceptable if the two UV traces meet the following requirements.
  - a. Peaks must be observed and should be symmetrical for all compounds in the calibration solution.
  - b. Corn oil and phthalate peaks should exhibit greater than 85% resolution.
  - c. The phthalate and Methoxychlor peaks should exhibit greater than 85% resolution.
  - d. Methoxychlor and perylene peaks should exhibit greater than 85% resolution.
  - e. Perylene and sulfur peaks must be saturated and should exhibit greater than 90% baseline resolution.

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YES NO N/A

- f. The RT shift is less than 5% between UV traces for bis(2-ethylhexylphthalate and perylene.
- 9.4 Were all above criteria met?

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ACTION: If no, examine the raw data for the presence of high molecular weight contaminants. Examine the subsequent sample data for unusual peaks and use professional judgment in qualifying the data.

## 10.0 Laboratory Control Samples (LCSs)

10.1 LCSs provide information on the accuracy of the analytical method and laboratory performance.

## Aroclor Laboratory Control Sample Recovery - Aqueous and Non-Aqueous

Compound	% Recovery QC Limit	
Aroclor 1016	50 - 150	
Aroclor 1260	50 - 150	
Tetrachloro-m-xylene (surrogate)	30 - 150	
Decachlorobiphenyl (surrogate)	30 - 150	

10.2 Were the above recoveries met?

ACTION: If no, qualify the sample data as follows:

	ACTION		
Criteria	Detected Associated Compound	Non-Detected Associated Compound	
%R> Upper Acceptance Limit	J	No qualification	
%R< Lower Acceptance Limit	J	R	
Lower Acceptance Limit < %R < Upper Acceptance Limit	No qualifi	cation	

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YES NO N/A

# 11.0 Aroclor Identification (Form X ARO/Identification Summary for Multicomponent Analysis

11.1 Is Form X (ARO) complete for every sample in which Aroclor was detected?

ACTION: Take action as specified in section 3.1 above.

11.2 The identification of a Multi component Aroclor by GC method is based primarily on RT data and pattern recognition. Were the following requirements met:

- a.) A Minimum of 3 major peaks were selected for each Aroclor. If more than one Aroclor is observed in a sample, a peak common to other Aroclor(s) must not be used to quantitate other Aroclor. Lab must choose different peaks to quantitate each Aroclor.
- b.) If a chromatogram is replotted electronically to meet these requirements, the scaling factor used must be displayed on the chromatogram, and both the initial chromatogram and the replotted chromatogram must be submitted in the data package.
- c.) The Retention Time (RT) of both of the surrogates and reported target compounds must be within the calculated RT window of both columns.
- d.) When no analytes are identified in the sample, the chromatograms of the sample extract must use the same scaling factor used for the low-point standard of the initial calibration associated with those samples.
- e.) Chromatogram must display the largest peak of any Aroclor detected in the sample at less than full scale.
- f.) If an extract must be diluted, chromatograms must display Aroclor peaks between 25-100% of full scale.
- ACTION: If retention times (RT) or peak apex cannot be verified, contact TOPO to obtain rescaled chromatograms from the lab.

If data reviewer identifies a peak in both GC columns that fall within the appropriate RT windows, but was reported as

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YES NO N/A

non-detect, the compound may be false negative. If necessary, contact TOPO to instruct laboratory to re-evaluate the chromatograms.

11.3 Are there any transcription/calculation errors in Form I and Form X ARO?

14

ACTION: Take action as specified in section 3.1 above.

11.4 Are the RTs of Aroclor peaks within the established RT window for analyses on both columns?

4

11.5 Was the GC/MS confirmation provided for Aroclor concentration > 10 ug/ml in final extract?

NOTE: Laboratory is required to contact SMO to determine if GC/MS confirmation is required. Check the semivolatile TIC data for presence of Aroclors.

11.6 Is the per cent difference (%D) calculated for positive results on both columns < 25%?

Action: Reviewer must check columns for peak interferences for the positive hits. Qualify the Arclor (s) according to the following Table:

## Action on Qualifying Positive Aroclor Results

Percent Differences	Qualifier
0 - 25%	None
26 - 70%	"Ј″
71 - 100%	. "JN"
101 - 200% (No Peak Interferences)	"R"
101 - 200% (Interferences detected)*	"ИС"

USEPA Region II Method: CLP/SOW, SOM01.2/Aroclor Date: August 2007 SOP HW-37/Aroclor, Revision 1

YES NO N/A

> 50% (Aroclo	value < CRQL)**	<b>"</b> υ"
> 200%		"R"

- \* When interferences is detected on either column, qualify the data as "JN"
- \*\* When the Aroclor value is below CRQL and %D > 50%, raise the value to CRQL and qualify "U", undetected.

## 12.0 Target Aroclor List (TCL)

12.1 Are the Aroclor Analysis Data Sheets (Form I ARO) present with required header information on each page for samples, MS/MSD (if required), method and instrument blanks (per column & analysis)? тл \_ \_

12.2 Is the chromatographic performance acceptable with respect to baseline stability, full-scale attenuation, peak shape/resolution?

m/ \_\_ \_

ACTION: If no, take action specified in section 3.1 above.

#### 13.0 Compound Quantitation and Reported Detection Limits

13.1 Are there any transcription/calculation errors in the Form I results? Check at least two positive results. Were any errors found?

ц 🗸 \_

ACTION: If errors were found, take action as specified in section 3.1 above.

13.2 Are the contract required quantitation limits (CRQL) adjusted to reflect sample dilution?

ACTION: If errors exist, take action as specified in section 3.1 above.

ACTION: When a sample is required to be diluted, the lowest CRQL is used (unless a QC exceedance dictates the use of the higher CRQL from the diluted sample). Replace concentration which exceed the calibration range in the original analysis by crossing out the "E" value on the original Form I and substituting it with the result from the diluted sample. Specify which Form I to use.

USEPA Region II Method: CLP/SOW, SOM01.2/Aroclor

Date: August 2007 SOP HW-37/Aroclor, Revision 1

YES NO N/A

Use a red pencil and draw a red "X" across the entire page of all Form I's that should not be used, including those in the data summary package.

At the top or bottom of the Forms, write with red pencil, "DO Not Use".

Note: If the sample dilution factor (DF) is greater than 10, an additional 10 times more concentrated than the diluted sample extract must be analyzed and reported with the sample data. If the DF is less or equal to 10, but greater than 1, the results of the original undiluted analysis must also be reported (see SOM01.1/section 10.3.3.4/page D-44/ARO).

ACTION: If the above requirement was not met, contact the TOPO to obtain an explanation/resubmittal from the lab and make a note in the Data Assessment under Contract Problems/Non-Compliance section.

13.3 For non-aqueous samples, were the percent moisture < 70%?

Action: If the % moisture  $\geq$  70.0% and < 90.0%, qualify detects as "J" and non-detects as approximated "UJ" If the % Moisture  $\geq$  90%, qualify detects as "J" and non-detects as "R"

[ ]

## 14.0 Field Duplicates

14.1 Were any field duplicates submitted for Aroclor analysis?

ACTION: Compare the reported results for field duplicates and calculate the relative percent difference.

ACTION: Any gross variation between duplicate results must be addressed in the reviewer narrative. If large differences exist, contact the TOPO to confirm identification of field duplicates with the sampler.

USEPA Region II Method: CLP/SOW, SOM01.2/Aroclor Date: August 2007 SOP HW-37/Aroclor, Revision 1

YES NO N/A

USEPA Region II

Method: CLP/SOW, SOM01.2/Aroclor

Date: August 2007 SOP HW-37/Aroclor, Revision 1

YES NO N/A

#### Definitions

ARO - Aroclor

CCS - contract compliance screening

CF - Calibration Factor

CLASS - Contract Laboratory Analytical Services Support

CLP - Contract Laboratory Program

CRQL - Contract Required Quantitation Limit

GC/ECD - Gas Chromatography/Electron Capture Detector

kg - kilogram

μg - microgram

ℓ - liter

me - milliliter

QC - quality control

RAS - Routine Analytical Services

RPD - Relative Percent Difference

RRF - Relative Response Factor

RRF - Average Relative Response Factor (from initial

calibration)

RRT - Relative Retention Time

RSD - Relative Standard Deviation

RT - Retention Time

RSCC - Regional Sample Control Center

SDG - Sample Delivery Group

SOP - standard operating procedure

SOW - Statement of Work

TCL - Target Compound List

TCLP - Toxicity Characteristics Leachate Procedure

TIC - Tentatively Identified Compound

TPO - Technical Project Officer

VTSR - Validated Time of Sample Receipt

TOPO - Task Order Project Officer

USEPA Region II Method: CLP/SOW, SOM01.2/Aroclor Date: August 2007 SOP HW-37/Aroclor, Revision 1

YES NO N/A

#### References

 USEPA Contract Laboratory Program of Work for Organic Analysis Multi-Media, Multi-Concentration, SOW/CLP/SOM01.2, February 2007.

2. National Functional Guidelines for Superfund Organic Methods Data Review July 2007.

## **APPENDIX G**

# HISTORICAL ARSENIC DATA RESULTS (YEAR 1992 AND YEARS 1994 THROUGH 1999)

(USEPALERT 6,1999)

FINAL REPORT
Vineland Chemical Site
Field Investigation
Vineland, Cumberland County, NJ
May 1999

U.S. EPA Work Assignment No.: 3-195 WESTON Work Order No.: 03347-143-001-3195-01 U.S. EPA Contract No.: 68-C4-0022

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1	Results of the Arsenic Analysis in Soil/Sediment
2	Results of the Arsenic Analysis in Water
3	In Situ Water Quality Parameters
4	Concentrations of Arsenic in Sediment, Soil and Water From 1992-1999

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No.	Title
1	Sample Location Map
2	Arsenic Concentrations in Sediment From 1992-1999
3	Arsenic Concentrations in Soil From 1994-1999
4	Arsenic Concentrations in Water From 1992-1999

#### 1.0 INTRODUCTION

#### 1.1 Objective

The objective of this study was to collect sediment, soil and water data to assess the public health hazard for arsenic contamination at three beaches along the Maurice River, two beaches along Union Lake, a potable water well at the Union Lake Sailing and Tennis Club, and potable water wells at two houses adjacent to the Vineland Chemical site. The data was evaluated against human health risk based action levels. This study was part of an annual monitoring program performed at beaches along the Maurice River and Union Lake.

#### 1.2 Site Background

The Vineland Chemical site is a 54-acre manufacturing facility located in Vineland, Cumberland County, NJ. The facility was involved in the production of arsenical herbicides, fungicides, and biocides since 1949. Arsenical feedstock compounds were historically stored in unprotected piles, a practice that has since been discontinued. This resulted in soil and groundwater contamination in the vicinity of the site. In addition, runoff during storm events and recharge of arsenic-bearing groundwater has contaminated the adjacent watershed, including the Blackwater Branch, Maurice River, and Union Lake. Arsenic-contaminated groundwater, process water, non-contact cooling water, and storm water runoff are currently treated on site. Effluent containing approximately 0.7 milligrams per liter (mg/L) arsenic is discharged from the wastewater treatment facility to an unlined lagoon where it percolates into the ground.

Previous studies have investigated the extent and magnitude of arsenic contamination in the Maurice River watershed (Faust et al. 1983, Weston 1988). Data available concerning arsenic contamination indicates that both the water and sediment are contaminated downstream of the site. The maximum arsenic concentrations detected in surface water, sediment and interstitial water were 2,780 micrograms per Liter (µg/L), 14,000 milligrams per kilogram (mg/kg) and 12.5 mg/L, respectively. The New Jersey Department of Environmental Protection (NJ DEP) and the United States Environmental Protection Agency (U.S. EPA) standards for arsenic in drinking water are 0.05 mg arsenic/L (U.S. EPA 1993). The extent of arsenic contamination ranges from the Blackwater Branch in the vicinity of the Vineland Chemical facility to a point approximately 26.5 miles downstream from the site.

Sediment, soil and surface water samples were collected at four beaches downstream of the site as part of an annual monitoring program (Weston 1992, 1995a, 1995b, 1996, 1997, 1998). In addition, potable water samples were collected from a well downstream of the site in 1996 and 1997. In 1998, additional potable water samples were collected at two houses adjacent to the Vineland Chemical property. Arsenic was not detected in any of the well samples.

In May 1999, the U.S. EPA Region II requested that the U.S. EPA/Environmental Response Team Center (ERTC) collect sediment, soil and water samples from three beaches along the Maurice River, two beaches at Union Lake, and potable water samples from the Union Lake Sailing and Tennis Club and from two houses adjacent to the site.

#### 2.0 METHODOLOGY

#### 2.1 Field Sampling Design

The sampling was conducted to assess the potential human exposure to arsenic and should not be interpreted as a comprehensive extent of contamination. The specific sampling locations were

determined by the U.S. EPA Region II Remedial Project Manager (RPM), Matthew Westgate (Figure 1). On 29 April 1999, two sediment, one soil and one disturbed water sample were collected at each of the five previously sampled beaches. The disturbed water sample was used to simulate potential human exposure to arsenic contaminated surface water and sediment during beach use. Potable water samples were collected from Union Lake Sailing and Tennis Club and from two houses adjacent to the site. The eight locations sampled were as follows:

Sampling Location	Description of Sampling Location
Alliance Beach	Upstream of Almond Beach, unmaintained public day-use area.
Almond Beach	Publicly maintained beach area approximately 100-150 feet long.
BA Beach	Downstream of Almond Beach, consisting of an unmaintained public day-use area.
Union Lake Beach	Privately maintained beach, downstream of the site.
Union Lakes Sailing and Tennis Club	Privately maintained club, downstream of the site, potable water well.
South End of Union Lake Beach	Publicly maintained beach at the southern end of Union Lake.
House #1	1618 Wheat Rd., across from the site.
House #2	1509 Wheat Rd., adjacent to the site.

All beach sampling areas were characterized by shallow depth (less than three feet), gradual slope and sluggish flow. The sediment was sandy near the shore with coarser sand and small to medium gravel in deeper areas. Thin deposits of black silt were evident at depositional areas along the bottom.

A Horiba U-10® Water Quality Monitoring Instrument was used at each sample location to measure temperature, pH, dissolved oxygen, conductivity, and salinity. The Horiba U-10® was operated according to the manufacturer's operating manual.

#### 2.2 Sediment, Soil and Water Sampling

Two sediment samples were collected from each of the five beaches sampled using a decontaminated Ponar dredge according to ERTC/REAC SOP #2016, Sediment Sampling. One sediment sample was collected upstream of the beach and one downstream of the beach. Sediment was collected from depositional areas where there was evidence of black silt. The dredge contents were composited into an aluminum tray, homogenized and transferred to a labeled 8-ounce glass jar.

Surface soil samples (0-4 inches below ground surface) were collected from each beach area using plastic trowels according to ERTC/REAC SOP #2012, Soil Sampling. The soil sample was composited into an aluminum tray, homogenized and transferred to a labeled 8-ounce glass jar.

Surface water samples were collected as per modification of ERTC/REAC SOP #2013, Water Sampling. The modification included collecting samples directly into a 1-L polyethylene bottle while

Submersed

disturbing the adjacent sediments. The samples were collected at a depth of 6 to 12 inches below the surface and approximately 6 inches above the bottom. The disturbed water sample was used to simulate potential human exposure to arsenic contaminated surface water and sediment during beach use.

The potable water samples were collected according to ERTC/REAC SOP #2051, *Potable Water Sampling*. All water samples were preserved after collection using 40 percent nitric acid to a pH of less than 2.

After each sample was collected, the labeled sample jars were placed in a resealable plastic bag and stored in a sample cooler on wet ice [4 degrees Celsius (°C)]. Field documentation (field logbook notes, and chain of custody forms) are located in Appendix A. The samples were delivered to the REAC Inorganic Laboratory in Edison, New Jersey, on 29 April 1999. The final analytical results are located in Appendix B.

#### 3.0 RESULTS

#### 3.1 Sediment Samples

Arsenic was detected in all sediment samples at concentrations ranging from 1.7 to 11 mg/kg (Table 1). The maximum concentration of arsenic (11 mg/kg) was detected in the upstream sample from the South end of Union Lake and the minimum concentration of arsenic (1.7 mg/kg) was detected in the downstream sample from BA Beach. Sediment grain size was qualitatively characterized as containing predominantly large grain sizes such as sand and small to medium gravel. Sediment collected from all five beaches contained a fine layer of highly suspendible silt.

#### 3.2 Soil Samples

Arsenic was detected in three of the five beach soil samples at concentrations ranging from 0.43 to 2.6 mg/kg (Table 1). The maximum concentration of arsenic (2.6 mg/kg) was detected at Union Lake and the minimum concentration of arsenic (0.43 mg/kg) was detected at Almond Beach. Arsenic was not detected at BA Beach and at South End of Union Lake Beach above the method detection limit (MDL) of 0.41 mg/kg. The soil at all five locations consisted primarily of sand and some small gravel.

#### 3.3 Water Samples

Arsenic was detected in all five surface water samples at concentrations ranging from 0.006 to 0.24 mg/L (Table 2). The maximum concentration of arsenic (0.24 mg/L) was detected at Alliance Beach and the minimum concentration of arsenic (0.006 mg/L) was detected at Union Lake. Arsenic was not detected (MDL = 0.002 mg/L) in Union Lake Sailing and Tennis Club, House #1 and House #2 potable water samples.

#### 3.4 In-Situ Water Quality

In-situ water quality parameters were consistent at all locations (Table 3). Temperature ranged from 13.2 to 16.9 °C and dissolved oxygen ranged from 8.66 to 11.5 mg/L. The pH ranged from 5.7 to 8.69 standard units. Conductivity ranged from 0.061 to 0.095 millimhos per centimeter (mmhos/cm). Salinity was measured at zero parts per thousand (ppt) at all locations.

#### 3.5 Field Blanks

Arsenic was not detected at concentrations above the MDL (0.002 mg/L for water and 0.48 mg/kg for soil and sediment) in any field blanks. As such, it is unlikely that any contamination can be attributed to improper sample collection and handling.

#### 4.0 DISCUSSION AND SUMMARY

Arsenic was detected in sediment samples collected at all locations at concentrations ranging from 1.7 to 11 mg/kg. The sediment in the Maurice River and Union Lake contains a high content of organic matter. Arsenic is typically adsorbed to the organic portion of sediment (Eisler 1988). Therefore, the arsenic may have been bound to the fine organic matter that was observed in the sediment.

Arsenic was detected in three of the five soil samples collected at concentrations ranging from 0.43 to 2.6 mg/kg. The soil samples were composed of coarse sand and some small gravel with very little organic matter. Arsenic typically binds to the organic matter in soil (Eisler 1988). The lack of organic matter in these samples implies a lack of adequate binding sites for arsenic. Therefore, rain or other inputs may cause the leaching of arsenic out of the soil and may account for the low concentrations of arsenic in these soil samples.

Arsenic was detected in all five surface water samples at concentrations ranging from 0.006 mg/L to 0.24 mg/L. Arsenic concentrations were above the U.S. EPA drinking water standard of 0.05 mg/L (U.S. EPA 1993) at two of the five beaches sampled (Alliance Beach and Almond Beach). Water samples were collected while disturbing the sediments so that particles of the highly suspendible fine silt layer were incorporated in the sample. Arsenic detection may be attributed to the organic fraction of the disturbed sediments present in the water samples. Riedel et al. (1988) found that the predominate component of detectable arsenic in water is associated with the sediment solids. Arsenic was not detected (MDL = 0.002 mg/L) in the potable well water samples taken at the Union Lake Sailing and Tennis Club, House #1 or House #2.

Concentrations of arsenic in the surface water collected from the Maurice River tended to be higher than the concentrations in surface water collected from Union Lake, with the exception of the BA Beach sample. Several reasons for this appear to exist. First, the Maurice River is directly downstream of the point-source of contamination and flows into Union Lake. Second, other tributaries flowing into Union Lake may cause the dilution of arsenic concentrations detected at these locations.

There are a number of factors that affect arsenic concentrations, including storm events, groundwater flow, flooding, grain size and sampling technique. Historic data for sediment, soil and water were compiled in Table 4 and Figures 2-4. No trends are discernable in the data as the concentration of arsenic in all matrices have remained relatively constant over time.





#### 5.0 REFERENCES

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## Table 1. Results of the Arsenic Analysis in Soil/Sediment Vineland Chemical Site Vineland, Cumberland County, New Jersey May 1999

## (Results reported in mg/kg)

Sample ID	Matrix	Location	Conc	MDL	
A19255	Soil	Alliance Beach	0.59	0.41	
A19256	Sediment upstream	Alliance-U	10	0.42	
A19257	Sediment downstream	Alliance-D	3.4	0.41	
A19251	Soil	Almond Beach	0.43	0.39	
A19252	Sediment upstream	Almond-U	3.4	0.44	
A19253	Sediment downstream	Almond-D	1.9	0.41	
A19259	Soil	BA Beach	Ū	0.47	
A19260	Sediment upstream	BA-U	6.6	0.41	
A19261	Sediment downstream	BA-D	1.7	0.44	
	T				
A19267	Soil	Union Lake	2.6	0.41	
A19268	Sediment upstream	Union Lake-U	6.0	0.47	
A19269	Sediment downstream	Union Lake-D	2.8	0.42	
A19263	Soil	So. End Union Lake	U	0.48	
A19264	Sediment upstream	So. End Union Lake-U	11	0.38	
A19265	Sediment downstream	So. End Union Lake-D	5.1	0.42	

 $\begin{array}{l} MDL \text{ - method detection limit} \\ U \text{ - not detected} \end{array}$ 

mg/kg - milligram per kilogram

## Table 2. Results of the Arsenic Analysis in Water Vineland Chemical Site Vineland, Cumberland County, New Jersey May 1999

## (Results reported in mg/L)

Sample ID	Location	Conc.	MDL
A19258	Alliance Beach	0.24	0.002
A19254	Almond Beach	0.11	0.002
A19262	BA Beach	0.01	0.002
A19270	Union Lake	0.006	0.002
A19271	Union Lake Yact Club House	U	0.002
A19266	South End Beach, Union Lake	0,02	0.002
A19272	House #1	U	0.002
A19273	House #2	U	0.002

MDL - method detection limit

U - not detected

mg/L - milligrams per Liter

Table 3. In-Situ Water Quality Parameters
Vineland Chemical Site
Vineland, New Jersey
May 1999

Location	Temperature (C)	Dissolved Oxygen (mg/L)	pН	(mmhos/cm)	Salinity (ppt)
Alliance Beach	15.1	8.91	6.30	0.079	Ø.00
Almond Beach	13.5	8.66	6.40	0.077	/ 0.00
B-A Beach	14.6	9.65	5.83	0.079	0.00
Union Lake	16.9	9.65	5.70	0.091	0.00
Union Lake Yact Club	14.8	11.50	7.00	0.061	0.00
So. End Union Lake	15.3	9.40	5.90	0.093	0.00
House #1	14.0	11.30	6.10	0.095	0.00
					<b>\</b>
House #2	13.2	11.24	8.69	0.085	0.00

C - degrees Celsius
mg/L - milligrams of dissolved oxygen per liter of water
mmhos/cm - micromhos per centimeter
NTU - nephelometric turbidity units
ppt - parts per thousand

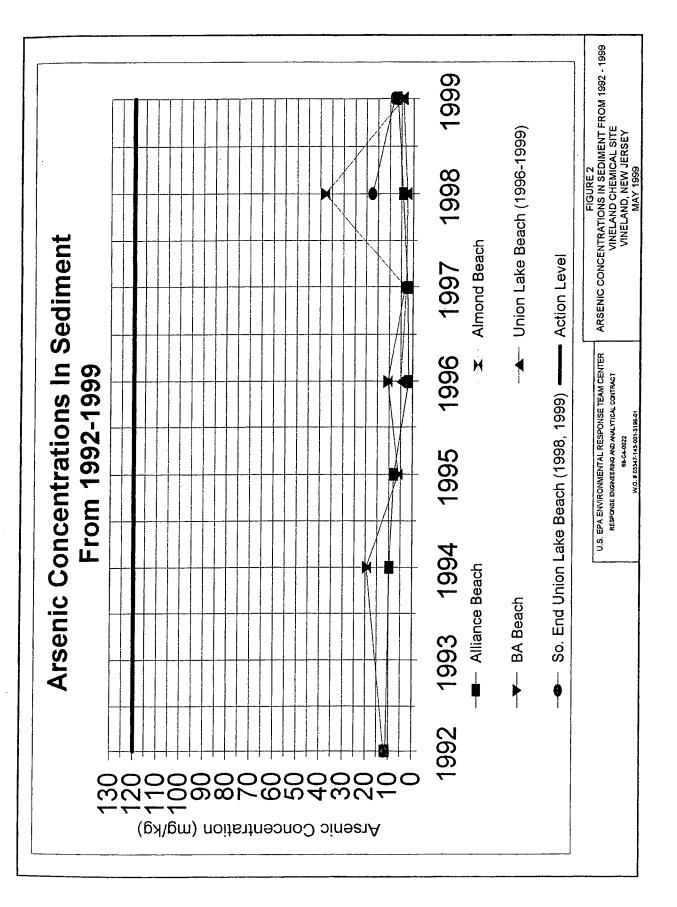
Table 4. Concentrations of Arsenic in Sediment, Soil and Water from 1992-1999
Vineland Chemical Site
Vineland, New Jersey
May 1999

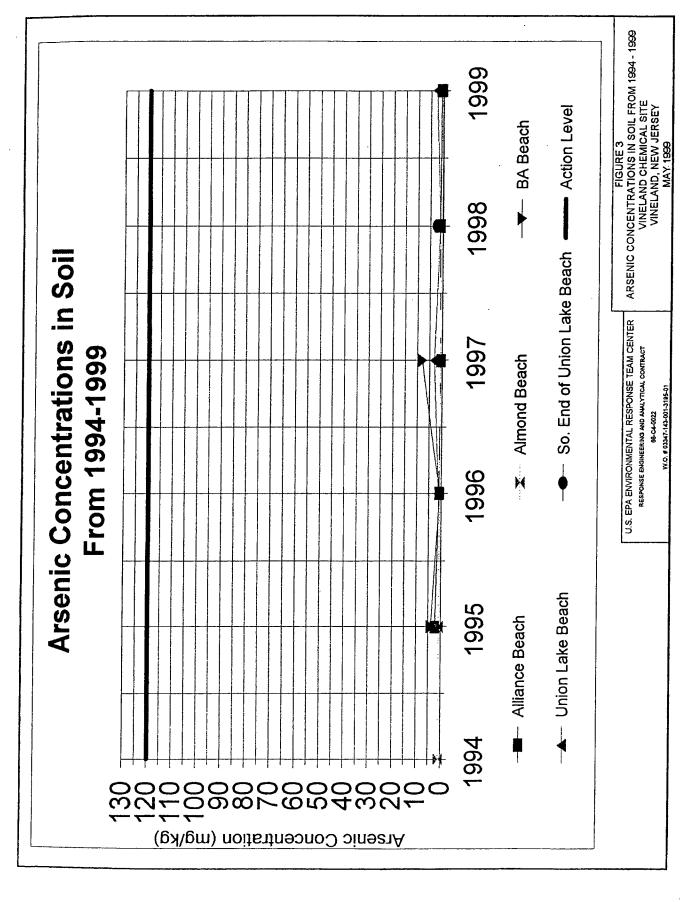
	1999			8661			1997			1996			1995			1994			1992	)		Year Sampled
Sediment	Soil	Water	Sediment	Soil	Water	Sediment	Soil	Water	Sediment	Soil	Water	Sediment	Soil	Water	Sediment	Soil	Water	Sediment	Soil	Water	441	Matrix
(mg/kg)	(mg/kg)	(mg/L)	(mg/kg)	(mg/kg)	(mg/L)	(mg/kg)	(mg/kg)	(mg/L)	(mg/kg)	(mg/kg)	(mg/L)	(mg/kg)	(mg/kg)	(mg/L)	(mg/kg)	(mg/kg)	(mg/L)	(mg/kg)	(mg/kg)	(mg/L)	,	Units
6.7	0.59	0.24	4.2	1.1	0.12	2.2	0.49	0.11	1.6	0.89	0.04	7.75	2.2	0.1	9.65	U (0.45)	0.023	11.1	NS	0.52		Alliance Beach
2.7	0.43	0.11	17	<b>P-1</b>	0.054	4.35	0.46	0.02	5.3	0.76	0.11	3.85	0.86	0.2	2	0.92	0.023	7.9	NS	0.15		Almond Beach
4.2	U (0.47)	0.01	38	U (0.41)	0.099	3.25	8.1	0.021	10.5	0.67	0.085	5.95	4	0.077	19.3	U (0.44)	0.039	11.8	NS	0.036		BA Beach
4.4	2.6	0.006	2.5	1.2	0.0088	2.65	3.1	0.0066	5	1.3	0.016	SN	SN	NS	SN	SN	SN	SN	SN	NS	Beach	Union Lake
8.1	U (0.48)	0.02	17.5	2.3	0.026	NS	NS	SN	SN	NS	NS	NS	SN	SN	SN	SN	NS	SN	SN	SN	Lake Beach	South End Union

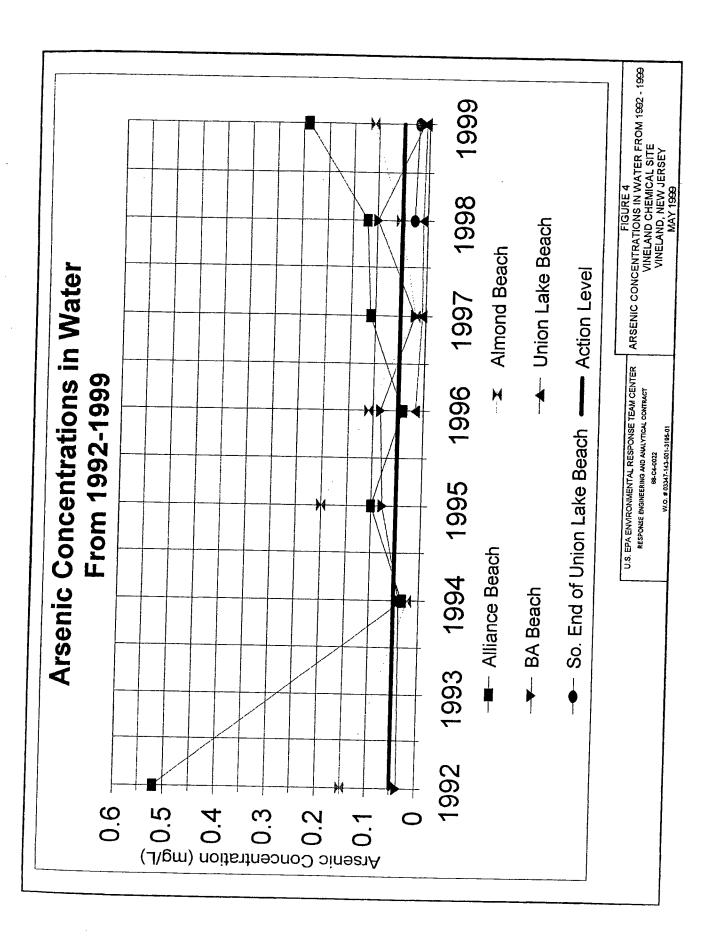
NS - Not Sampled Note: Sediment concentrations reflect a mean of the upstream sample and the downstream sample

U - not detected at indicated concentration Action Limit: Soil/Sediment - 120 mg/kg, Water - .05 mg/L mg/L - milligrams per Liter

mg/kg - milligram per kilogram







APPENDIX A
Field Documentation
Vineland Chemical Site
Final Report
May 1999

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APPENDIX B
Analytical Report
Vineland Chemical Site
Final Report
May 1999

#### ANALYTICAL REPORT

Prepared by Roy F. Weston, Inc.

Vineland Chemical Site
Vineland, Cumberland County, New Jersey

May 1999

EPA Work Assignment No. 3-195
WESTON Work Order No. 03347-143-001-3195-01
EPA Contract No. 68-C4-0022

Submitted to M. Sprenger EPA-ERTC

Analysis by:

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Analytical Section Leader | S/7/66 | Reviewed by:

E. Gilardi Date M. Barkley

Project Manager

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#### Introduction

REAC in response to WA #3-195, provided analytical support for environmental samples collected from the Vineland Chemical Site located in Vineland, Cumberland County, New Jersey as described in the following table. The support also included QA/QC, data review, and preparation of an analytical report containing a summary of analytical methods, results, and QA/QC results.

The samples were treated with procedures consistent with those specified in SOP #1008.

Chain of Custody	Number of Samples	Sampling Date	Date Received	Matrix	Analysis	Laboratory
03503	5	4/29/99	4/30/99	Water	Arsenic	REAC
	15			Soil		
03504	4			Water		
	2			Soil		

#### **CASE NARRATIVE**

#### Data Package I181

The arsenic (As) percent recoveries for soil samples A19256MS (46%) and A19256MSD (48%) exceeded the QC limits. The As results for soil samples A19251, A19252, A19253, A19255, A19256, A19257, A19259, A19260, A19261, A19263, A19264, A19265, A19267, A19268, A19269, A19274 and A19275 are considered estimated.

#### Summary of Abbreviations

AA	Atomic Absorption		
В	The analyte was found in	the blank	
BFB	Bromofluorobenzene		
BPQL	Below the Practical Quar	ntitation Limit	
BS `	Blank Spike		
BSD	Blank Spike Duplicate		
C	Centigrade		
Ď	(Surrogate Table) this val	lue is from a dilute	ed sample and was not calculated
_	(Result Table) this result	was obtained from	a diluted sample
CLP	Contract Laboratory Prot		
COC	Chain of Custody	••••	•
CONC	Concentration		
CRDL	Contract Required Detect	tion Limit	•
CROL	Contract Required Quant		
DFTPP	Decafluorotriphenylphos		
DL	Detection Limit	PILLIO	
E	The value is greater than	the highest linear	standard and is estimated
EMPC	Estimated maximum poss	ible concentration	
I I	The value is below the me		
ICAP	Inductively Coupled Argo		
IDL	Instrument Detection Lim		
ISTD	Internal Standard	146	•
MDL	Method Detection Limit		
	Method Quantitation Lim	<b>:</b> +	
MQL MI	Matrix Interference	It	
MRL	Method Reporting Limit		
MS.			
MSD	Matrix Spike Matrix Spike Duplicate		
MW	Molecular Weight		
NA	either Not Applicable or I	Jot Available	
NC	Not Calculated	tot Available	
NR NR	Not Requested		
NS	Not Spiked		
% D	Percent Difference		
% REC	Percent Recovery		
PQL	Practical Quantitation Lin		
PPBV	Parts per billion by volum	C	
QL	Quantitation Limit	••	
RPD	Relative Percent Difference		
RSD	Relative Standard Deviation	on	
SIM	Selected Ion Mode		
U	Denotes not detected	- 41	
W		•	ld be regarded as estimated.
m	cubic meter	kg	kilogram
L	liter	g	gram
dL	deciliter	cg	centigram
mL	milliliter	mg	milligram
$\mu$ L	microliter	$\mu$ g	microgram
ng	nanogram	pg	picogram

denotes a value that exceeds the acceptable QC limit

Abbreviations that are specific to a particular table are explained in footnotes on that table

Revision 7/9/98

#### Analytical Procedure for Arsenic in Water

#### Sample Preparation

A representative 45 mL aliquot of each sample was mixed with 5.0 mL concentrated nitric acid, placed in an acid rinsed Teflon container, capped with a Teflon lined cap, and digested according to SW-846, Method 3015 in a CEM MDS-2100 microwave oven, which was programmed to bring the samples to 160 +/- 4°C in 10 minutes (first stage) and slowly rise to 165-170°C in the second 10 minutes (second stage). After digestion, samples were allowed to cool to room temperature and were transferred to polyethylene bottles. Samples were analyzed for all metals, except mercury, by US EPA SW-846, Method 7000 Atomic Absorption (AA) or Method 6010 Inductively Coupled Argon Plasma (ICAP) procedures.

A reagent blank and a blank spike sample were carried through the sample preparation procedure for each analytical batch of samples processed. One matrix spike (MS) and one matrix spike duplicate (MSD) sample were also processed for each analytical batch or every 10 samples.

Analysis and Calculations

The AA and ICAP instruments were calibrated and operated according to SW-846, Method 7000/7470/6010 and the manufacturer's operating instructions. After calibration, initial calibration verification (ICV), initial calibration blank (ICB), and QC check standards were run to verify proper calibration. The continuing calibration verification (CCV) and continuing calibration blank (CCB) standards were run after every 10 samples to verify proper operation during sample analysis.

The metal concentrations in solution, in micrograms per liter (µg/L) were read directly from the read-out systems of the instruments. ICAP and Mercury results were taken directly from instrument read-outs. The ICAP results were corrected for digestion volume (45 mL sample + 5 mL nitric acid) prior to instrument read-out; AA read-outs (excluding Mercury) were externally corrected for digestion volume (1.1111 \* AA read-out).

For samples that required dilution to fall within the instrument calibration range:

 $\mu$ g/L metal in sample = A [ (C+B) / C ]

where:

A = direct read-out (ICAP and Mercury)

A = corrected read-out (AA)

B = acid blank matrix used for dilution, mL

C = sample aliquot, mL

Results of the analyses are listed in Table 1.1.

#### Analytical Procedure for Arsenic in Soil

#### Sample Preparation

A representative 1-2 g (wet weight) sample, weighed to 0.01 g accuracy, was mixed with 10 mL 1:1 nitric acid, placed in a clean beaker and digested in nitric acid and hydrogen peroxide according to SW-846, Method 3050. The final reflux was either nitric acid or hydrochloric acid depending on the metals to be determined. After digestion, the samples were allowed to cool to room temperature and transferred to 100 mL volumetric flasks and diluted to volume with ASTM Type II water. The samples were analyzed for all metals, except mercury, by USEPA SW-846, Method 7000 (Atomic absorption) or Method 6010 (Inductively Coupled Argon Plasma-ICAP) procedures.

A separate sample was used to determine total solids.

A reagent blank and a blank spike sample were carried through the sample preparation procedure for each batch of samples processed. One matrix spike (MS) and one matrix spike duplicate (MSD) were analyzed for each batch or for every ten samples.

Analysis and Calculations

The instruments were calibrated and operated according to SW-846, Method 7000/7471/6010 and the manufacturers operating instructions. After calibration, initial calibration verification (ICV), initial calibration blank (ICB) and quality control check standards were run to verify proper calibration. The continuing calibration verification (CCV) and continuing calibration blank (CCB) were run after every ten samples to assure proper operation during sample analysis.

The metal concentrations in solution, in micrograms per liter (µg/L) were taken from the read-out systems of the Atomic Absorption instruments. The results were converted to milligrams per kilogram (mg/kg) by correcting the reading for the sample weight and percent solids. The ICAP results (mg/kg) were corrected for sample weight prior to instrument read-out; the instrument read-out was then corrected for percent solids.

Final concentrations, based on wet weight are given by:

```
mg metal/kg sample = [(AxV)/W]xDFxCF

where:

A = Instrument read-out (µg/L, AA; mg/kg, ICAP)

V = final volume of processed sample (mL, AA; 1.00 ICAP)

W = weight of sample (g, AA; 1.00 ICAP)

DF = Dilution Factor (1.00 for no dilution)

CF = conversion factor (0.001, AA; 1.00, ICAP)
```

For samples that required dilution to be within the instrument calibration range, DF is given by:

```
DF = (C+B)/C

where:

B = acid blank matrix used for dilution (mL)

C = sample blank aliquot (mL)
```

Final concentrations, based on dry weight, are given by:

```
mg/kg(dry) =[mg/kg (wet)x100] /S
where
S = percent solids
```

The results are listed in Table 1.2.

Table 1.2 Results of the Analysis for Arsenic in Soil WA# 3195 Vineland Chemical Site Results Based on Dry Weight

Parameter: Analysis Method:

Arsenic AA-Furnace

,,	-		•		
Client ID	Location	Percent Solids	Conc mg/kg	MDL mg/kg	
Method Blank	Lab	NA	U	0.50	
A19251	Almond-S	82.91	0.43	0.50 0.39	<i>;</i>
A19252	Almond-Sd-U	76.51	3.4	0.39 0.44	
A19253	Almond-Sd-D	80.00	1.9	0.41	•
A19255	Alliance-S	87.68	0.59	0.41	
A19256	Alliance-Sd-U	82.49	10	0.42	
A19257	Alliance-Sd-D	85.09	3.4	0.42	
A19259	BA Beach-S	99.53	Ü	0.47	
A19260	BA Beach-Sd-U	79.54	6.6	0.41	
A19261	BA Beach-Sd-D	77.98	1.7	0.44	
A19263	So. End Union Lake-S	99.57	ີ	0.48	
A19264	So. End Union Lake-Sd-U	74.95	11	0.38	
A19265	So. End Union Lake-Sd-D	71.90	5.1	0.42	
A19267	Union Lake-S	86.25	2.6	0.41	
A19268	Union Lake-Sd-U	77.54	6.0	0.47	•
A19269	Union Lake-Sd-D	82.51	2.8	0.42	
A19274	Field Blank-S	100.00	Ü	0.48	
A19275	Field Blank-Sd	100.00	Ü	0.48	

Table 1.1 Results of the Analysis for Arsenic in Water WA# 3195 Vineland Chemical Site

Parameter: Analysis Metho	od:	Arsenic AA-Furna	Arsenic AA-Furnace		
Client ID	Location	Conc ug/L	MDL ug/L		
Method Blank	Lab	U	2.2		
A19254	Almond-SW	110	2.2		
A19258	Alliance-SW	240	2.2		
A19262	BA Beach-SW	. 11	2.2		
A19266	So. End Union Lake-SW	15	2.2		
A19270	Union Lake-SW	5.8	2.2		
A19271	Union Lake-P	U	2.2		
A19272	House #1-P	U	2.2		
A19273	House #2-P	U	2.2		
A19276	Field Blank-SW	υ	2.2		

#### QA/QC for Arsenic

#### Results of the OC Standard Analysis for Arsenic in Water

The QC standard TMAA#1 was used to check the accuracy of the calibration curve. The percent recovery for the arsenic found in the QC standard listed in Table 2.1, was 97 and within the 95% confidence interval limit.

#### Results of the MS/MSD Analysis for Arsenic in Water

Sample A19258 was chosen for matrix spike/matrix spike duplicate (MS/MSD) analysis. The percent recoveries, listed in Table 2.2, wer 76 and 99. Both recoveries were within QC limits. The relative percent difference (RPD), also listed in Table 2.2, was 27 and outside the QC limits.

#### Results of the Blank Spike Analysis for Arsenic in Water

The percent recovery for the blank spike arsenic, listed in Table 2.3, was 99 and within QC limits.

#### Results of the QC Standard Analysis for Arsenic (Soil)

The QC standard TMAA#1 was used to check the accuracy of the calibration curve. The percent recovery for the arsenic found in the QC standard listed in Table 2.4, was 96 and within the 95% confidence interval limits

#### Results of the MS/MSD Analysis for Arsenic in Soil

Samples A19255, A19256, and A19257 were chosen for matrix spike/matrix spike duplicate (MS/MSD) analysis. The percent recoveries, listed in Table 2.5, ranged from 46 to 83. Four out of 6 recoveries were within QC limits. The relative percent differences (RPDs), also listed in Table 2.5, ranged from 2 to 10. All 3 RPDs were within QC limits.

#### Results of the Blank Spike Analysis for Arsenic in Soil

The percent recovery for the blank spike arsenic, listed in Table 2.6, was 95 and within QC limits.

Table 2.1 Results of the QC Standard Analysis for Arsenic in Water WA# 3195 Vineland Chemical Site

Metal	Date Analyzed	Quality Control Standard	Conc. Recovered ug/L	Certified Value ug/L	95 % Confidence Interval	% Recovery
Arsenic	04/30/99	TMAA #1	48.53	· 50	41.9-55.9	97

### Table 2.2 Results of the MS/MSD Analysis for Arsenic in Water WA# 3195 Vineland Chemical Site

Sample ID:	A19258	Sample Conc µg/L	MS Spike Added µg/L	MS Conc µg/L	MS % Rec	MSD Spike Added µg/L	MSD Conc µg/L	MSD % Rec	RPD	Recomm QC Li % Rec	
Arsenic		244	55.6	299	99	55.6	286	76	27 •	75-125	20

Table 2.3 Results of the Blank Spike Analysis for Arsenic in Water WA# 3195 Vineland Chemical Site

Metal	Spiked Conc ug/L	Recovered Conc. ug/L	% Recovery	Recommended QC Limit % Rec	
Arsenic	55.6	55.1	99	75-125	٠.

### Table 2.4 Results of the QC Standard Analysis for Arsenic (Soil) WA# 3195 Vineland Chemical Site

Metal	Date Analyzed	Quality Control Standard	Conc. Recovered ug/L	Certified Value ug/L	95 % Confidence Interval	% Recovery
Arsenic	05/03/99	TMAA #1	48	50	41.9-55.9	96

## Table 2.5 Results of the MS/MSD Analysis for Arsenic in Soil WA# 3195 Vineland Chemical Site Results Based on Dry Weight

Sample ID:	A19255	Sample Conc mg/kg	MS Spike Added	MS Conc mg/kg	MS % Rec	MSD Spike Added mg/kg	MSD Conc mg/kg	MSD % Rec	RPD	Recommended QC Limits	
Metal			mg/kg							%Rec	RPD
Arsenic .		0.587	4.04	3.61	75	4.19	3.79	76	2	, 75-125	20

## Table 2.5 (cont.) Results of the MS/MSD Analysis for Arsenic in Soil WA# 3195 Vineland Chemical Site Results Based on Dry Weight

Sample ID:	A19256	Sample Conc mg/kg	MS Spike Added mg/kg	MS Conc mg/kg	MS % Rec	MSD Spike Added mg/kg	MSD Conc mg/kg	MSD % Rec	RPD	Recommo QC Lin %Rec	
Arsenic		10.4	4.36	12.4	46 •	4.39	12.5	48 *	• 4	75-125	20

## Table 2.5 (cont.) Results of the MS/MSD Analysis for Arsenic in Soil WAlf 3195 Vineland Chemical Site Results Based on Dry Weight

Sample ID:	A19257	Sample Conc mg/kg	MS Spike Added mg/kg	MS Conc mg/kg	MS % Rec	MSD Spike Added mg/kg	MSD Conc mg/kg	MSD % Rec	RPD	Recomm QC Lir %Rec	
Arsenic	<del>*************************************</del>	3.44	4.29	6.99	83	4.32	6.66	75	10 .	75-125	20

Table 2.6 Results of the Blank Spike Analysis for Arsenic in Soil WA# 3195 Vineland Chemical Site

Metal	Spiked Conc mg/kg	Sand Blk Conc. mg/kg	Recovered Conc. mg/kg	% Recovery	Recommended QC Limit % Rec
Arsenic:	4.95	U	4.71	95	75-125

REAC, Edison, NJ EPA Contract 68-C4-0022 (908) 321-4200

## CHAIN OF CUSTODY RECORD

Project Number: Project Name: 13347-143-001-3195-01 Lucland

NO.

Matrix: SD -DC -V -D430% REAC # 125 6 123 120 121 2 109 Sediment
Drum Solids
Drum Liquids
Other 119261 A19256 4BC 19258 A19260 419259 A19255 419257 H19251 A19270 419264 H19264 419257 #1926 B 872614 295611 119263 119254 119269 85rb/ Sample No. A19267 82 % SW-SW-SW-BA Brach - Sw BA Bush - Sw BA Beach - Sd-D BA Brown-Su- U Allians C-Sd. D Alliance - Sd- U Alliance - 5 Almond - Sa- U Alment - Sd-1 0-15- may said lano Cale-Sd-U Elmond. S Sampling Location End Union lake Soft End Union Lake-Sy-U End Haimlake Potable Water Groundwater Surface Water Studge Sample Identification نعاد: علما Watrix 8 8 1V 25 8/8/2 58 馬克 کیا SE >050 Date Collected Soil Water Air 4/29/99 RFW Contact: Special Instructions: \* Ms/mso # of Bottles 8 of Kt 15004 BOX Var Container/Preservative 120 5.t. 14.1 20 6001 antiasta) 20.11 ASE HIVO A Segue A Se Huo 14.0 7.4 200 Phone: 732 - 494 - 4004 Hrsen, C CUSTODY # FROM CHAIN OF FOR SUBCONTRACTING USE ONLY **Analyses Requested** SHEET NO. LOF 2 03503 00016

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EPA Contract 68-C4-0022 REAC, Euson, NJ (908) 321-4200

# CHAIN OF CUSTODY RECORD

Project Number: Project Name: 03347-145.001 3145-01 liseland Chemica

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RFW Contact:\_ Jun Rosse \_Phone:\_ 732-494-4004

Matrix: SD. DS. 043099-REAC # 129 30 2000 હ Sediment
Drum Solids
Drum Liquids
Other A19275 A19276 A19272 A18273 A19274 Sample No. 1:1271 SW-SW-SW-Sampling Location Eield Blant-S Unicaloke - P Field Blant-Sd Field Blank - Su House # 1 - P Yourse #2- P Potable Water Groundwater Surface Water Studge Sample Identification Matrix رين E 以公 **>** 0 **≥** s Date Collected Soil Water Oil 4/29/19 Special Instructions: # of Bottles Container/Presegrative 5.0200 (Lange Har) 1400 Rusenic **Analyses Requested** SHEET NO & OF -

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FOR SUBCONTRACTING USE ONLY

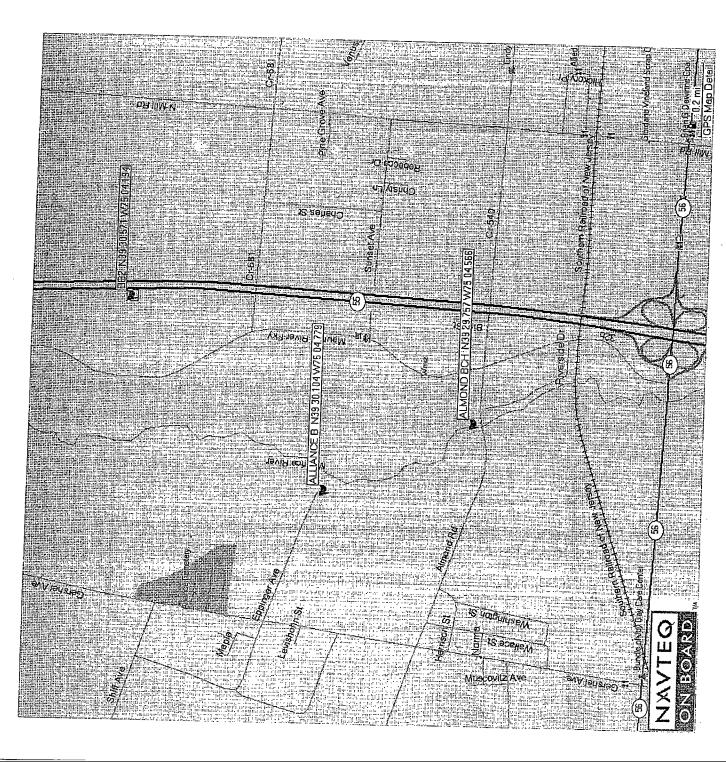
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